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Expedition 368X methods supplement¹

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Introduction

Site location

An acoustic positioning beacon was not deployed during International Ocean Discovery Program (IODP) Expedition 368X, but one was prepared for immediate deployment if required.

Core handling and analysis

Sediment

During Expedition 368X, a 5 cm whole-round sample for paleontological (PAL) analysis was collected from the core catcher of most sediment cores by the IODP *JOIDES Resolution* Science Operator (JRSO) technical staff. The sample was vacuum sealed and placed in cold storage for postcruise analysis. Paleontological samples were not collected from cores with exceptionally low recovery or from sediments that were not soft or semilithified. Paleontological samples were distributed postcruise to micropaleontologists of Expeditions 367 and 368. Postcruise processing of paleontological samples is described in [Biostratigraphy](#).

The archive half of each core was preliminarily described on board by two structural geologists and a petrologist. Shipboard descriptions were primarily macroscopic except for seven smear slides and six thin sections. Descriptions of each core were completed during the subsequent description party at the IODP Gulf Coast Repository (GCR; Texas A&M University, College Station, Texas [USA]).

Authorship of chapters

The separate sections of the methods and site chapters were written during Expedition 368X, and the subsequent core description and editorial meeting was held at the GCR on 16–24 April

2019. Content was contributed by the following scientists (authors are listed in alphabetical order; see [Expedition 368X participants](#) for affiliation information):

- Operations: Childress and Midgley
- Lithostratigraphy, igneous and metamorphic petrology, and structural geology: Dadd, Huang, Nirrengarten, Peate, Sun, van der Zwan, and Zhong
- Biostratigraphy: Alvarez Zarikian, Li, C. Liu, and Xiang
- Paleomagnetism: Satolli and Yi
- Geochemistry: Childress
- Physical properties: Briais, Deng, Lin, Qiu, and Stock
- Downhole measurements and seismic correlation: Briais, Deng, Lin, and Qiu

Core description (lithostratigraphy, igneous and metamorphic petrology, and structural geology)

The lithostratigraphy, structural geology, and igneous petrology methods from Expedition 367/368 (see the [Expedition 367/368 methods](#) chapter [Sun et al., 2018]) were consistently followed by the core description group during Expedition 368X and the subsequent postcruise core description meeting at the GCR.

Visual core description

Expedition 368X graphic visual core descriptions (VCDs) include a simplified graphical representation of Site U1503 on a core-by-core basis. VCDs illustrate an interval-by-interval record of the primary lithologies contained within each core by pairing the prin-

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cipal lithology name assigned to each interval in DESClogik with a predetermined set of lithology patterns for sedimentary (Figure F1) and igneous (Figure F2) core material. See the [Expedition 367/368 methods](#) chapter (Sun et al., 2018) for detailed information.

Chemical analysis

X-ray fluorescence

During Expedition 368X, we expanded the use of the handheld portable X-ray fluorescence spectrometer (pXRF; Olympus Delta) to include sediment section halves. Although many factors can impact the accuracy of section-half pXRF measurements, the presence of pore water results in large relative standard deviation (RSD) values (5%–20%) that increase with increasing pore water content. This makes quantitative results from section-half measurements less reliable for sediment. However, pXRF measurements of section halves can provide qualitative data on the elemental composition of sediment, including relative percent abundances. pXRF measurements of sediment section halves provide complementary results to smear slide and X-ray diffraction (XRD) analyses.

By comparing the results of shipboard pXRF data and published values of United States Geological Society (USGS) rock standard

basalt (BHVO-2; e.g., Wei et al., 2014), we determined that discrete-powder pXRF results are reliable for elements with a content >10 ppm (Figure F3). Comparisons were then made between measurements of section-half surfaces, quarter-core wedges, and powder samples of material from Hole U1503A to better estimate error. Quarter-core wedges show a 1%–20% discrepancy with powders of the same material, especially for Ti, K, Sr, Zr, Ni, and Cu, whereas measurements of section-half surfaces usually show an even greater discrepancy with powder samples, especially for Mg, Mo, Ca, Si, Al, and Zn (Figure F4A). To produce the best-quality data, section halves were measured after their surfaces were dried for imaging. Surfaces that were flat and large enough for the detector (roughly 2–4 cm long) were selected to assure consistent measurements between each data point. Fractures, nonplanar surfaces, and areas of drilling disturbance were avoided. Measurements along sediment section halves were made every 20–40 cm, depending on the lithologic variation. Basalt rock measurements were done on centimeter-scale broken pieces. In contrast to the sediment section halves, there is a closer correspondence for most elements between measurements of basalt pieces and basalt powder (much closer to the diagonal line) than sediment section halves (Figure F4).

Figure F1. Lithology patterns and symbols used for visual core description of sedimentary intervals, Expedition 368X.

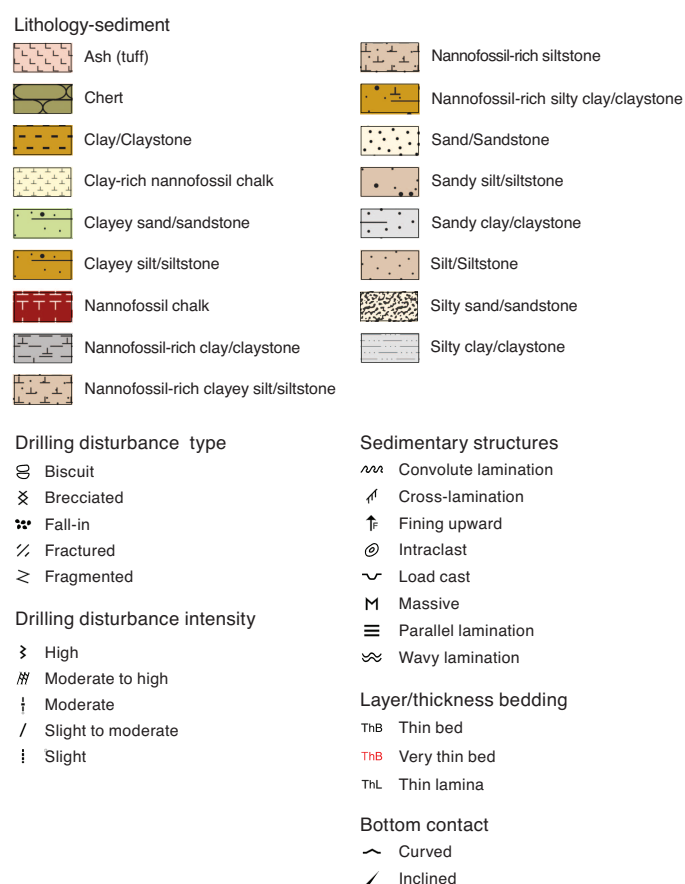


Figure F2. Lithology pattern and symbols used for visual core description of igneous intervals, Expedition 368X.

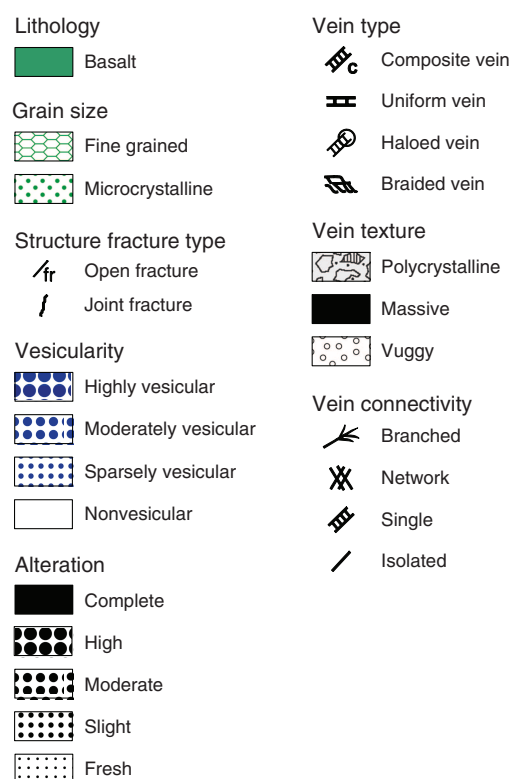


Figure F3. pXRF measurements versus published measurements of rock Standard BHVO-2 (e.g., Wei et al., 2014) used to test the accuracy and reproducibility of discrete powder pXRF data during Expedition 368X. Accuracy and reproducibility are good for most elements, but large discrepancies were observed for elements with values <10 µg/g. P, Cr, and V are also less consistent than published standard measurements, generally underreporting for pXRF measurements. A, B = repeated tests on the same standard vial, C–E = three separate BHVO-2 sample vials.

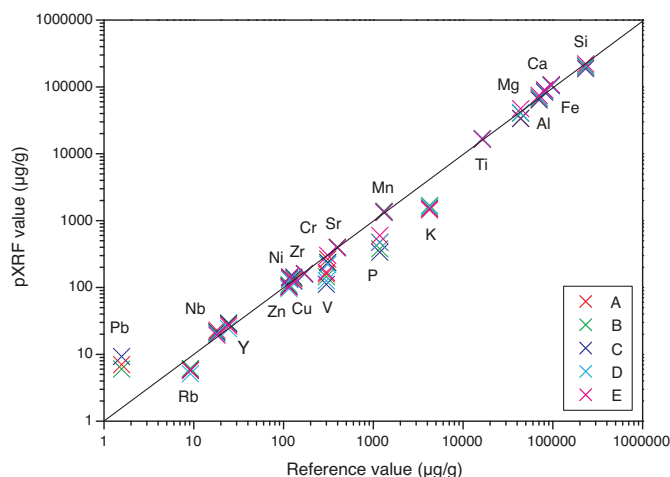
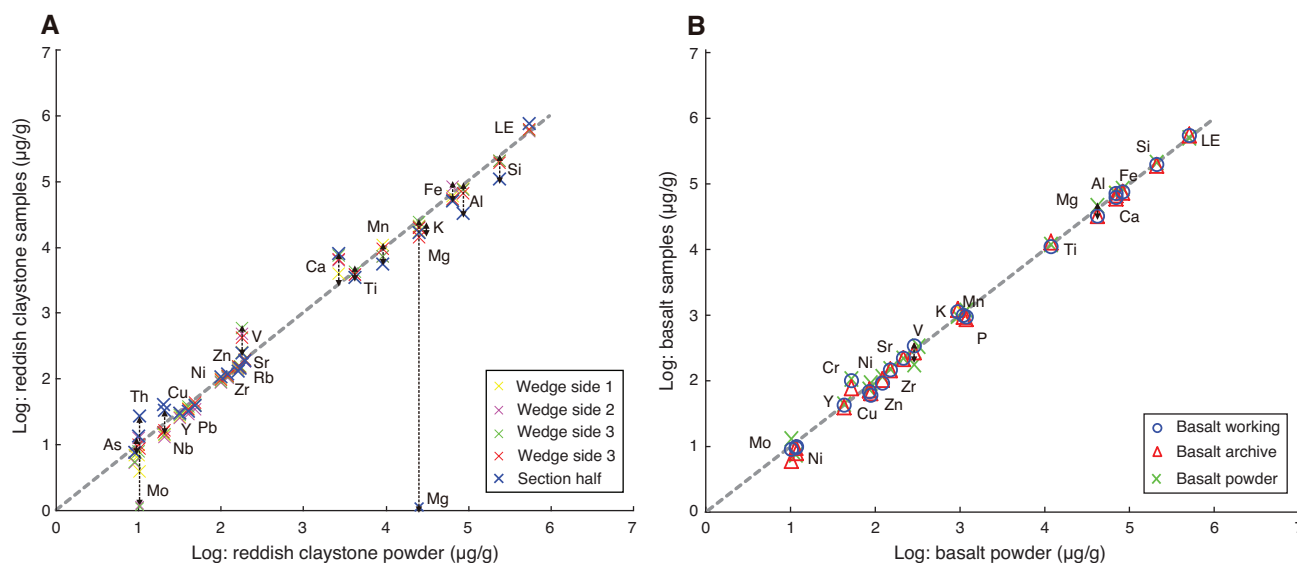


Figure F4. pXRF quarter-core wedges or core pieces versus powders of the same material, Hole U1503A. A. Sediment (56R-3, 12–14 cm). Three cut surfaces of the wedge sample were measured (one side in duplicate). Black dashed line with arrows = range of measurements with large differences between methods. B. Basalt (80R-3, 55–57 cm). A duplicate measure of the powdered basalt is also plotted.



Biostratigraphy

Calcareous nannofossils and planktonic foraminifers were studied from core catcher samples after Expedition 368X to build a biostratigraphic framework for Hole U1503A. Twenty-seven core catcher samples from Cores 368X-U1503A-2R through 48R were split at the GCR after Expedition 368X and distributed to members of the Expedition 367 and 368 micropaleontology team for examination at their home institutions. Additional samples from the core sections were examined during the Expedition 368X onshore core description and editorial meeting to refine age determinations at selected intervals. Biostratigraphic events, mainly the first appearance datum (FAD; or base) and last appearance datum (LAD; or top) of the diagnostic species are tied to the geomagnetic polarity timescale of Gradstein et al. (2012) (Figure F5).

Calcareous nannofossils

The calcareous nannofossil zonation is based on the schemes of Okada and Bukry (1980) and Martini (1971). Calibrated ages for bioevents are from Gradstein et al. (2012) and given in Table T1. The Gradstein et al. (2012) timescale assigns the Pleistocene/Pliocene boundary between the Gelasian and Piacenzian stages (2.59 Ma), the Pliocene/Miocene boundary between the Zanclean and Messinian stages (5.33 Ma), the late/middle Miocene boundary between the Tortonian and Serravallian stages (11.63 Ma), the middle/early Miocene boundary between the Langhian and Burdigalian stages (15.97 Ma), and the Miocene/Oligocene boundary between the Aquitanian and Chattian stages (23.03 Ma). For calcareous nannofossil biostratigraphy, the Pleistocene/Pliocene boundary now falls in Zone NN16 (Martini, 1971) between the LAD of *Discoaster surculus* (2.49 Ma) and *Discoaster tamalis* (2.8 Ma). The

[illegible]

Several species of the genus *Gephyrocapsa*, which are commonly used as Pleistocene biostratigraphic markers, often show a great range of variation in sizes and other morphological features, which causes problems in identification (e.g., Samtleben, 1980; Su,

Several *Reticulofenestra* species with different coccolith and central opening sizes have been used as Neogene and Quaternary biostratigraphic markers; however, these parameters show considerable variations within and between "species," making species differentiation difficult (e.g., Su, 1996; Young, 1998). In this study, we followed the definition of *Reticulofenestra pseudoumbilicus* by Young (1998) as having a maximum coccolith length $>7\ \mu\text{m}$ (similar to the size of its holotype), especially for specimens from its uppermost range in the early Pliocene. We distinguished *Reticulofenestra asanoi* from the similarly sized *Pseudoemiliania lacunosa* by the absence of slits on the shield (Su, 1996). In addition, we further

Table T1. Calcareous nannofossil events and scaled ages (Gradstein et al., 2012) used during Expedition 368X. GTS2012 = 2012 geologic timescale. T = top/last appearance; B = base/first appearance. Bold = age-diagnostic datum. (Continued on next page.) [Download table in CSV format.](#)

GTS2012 chronostratigraphy	Standard tropical–subtropical biozone (biochron)		Biohorizon (datum)	GTS2012 age (Ma)	
	CN zones (Okada and Bukry, 1980)	NN zones (Martini, 1971)			
0.126 Ma					
Ionian (middle Pleistocene)	CN15/CN14b	NN21/NN20	B <i>Emiliania huxleyi</i>	0.29	
	CN14b/CN14a	NN20/NN19	T <i>Pseudoemiliania lacunosa</i> T <i>Gephyrocapsa</i> sp. 3	0.44 0.61	
0.781 Ma					
Calabrian	CN14a	NN19	T <i>Reticulofenestra asanoi</i> (common)	0.91	
			T small <i>Gephyrocapsa</i> spp. dominance	1.02	
			B <i>Gephyrocapsa</i> sp. 3	1.02	
	CN14a/CN13b		B medium (>4 μm) <i>Gephyrocapsa</i> spp. reentrance (reemG event)	1.04	
			B <i>Reticulofenestra asanoi</i> (common)	1.14	
			T large (>5.5 μm) <i>Gephyrocapsa</i> spp.	1.24	
	CN13b	B small <i>Gephyrocapsa</i> spp. dominance	1.24		
		T <i>Helicosphaera sellii</i>	(1.26)		
		T <i>Calcidiscus macintyreii</i>	1.60		
		B large (>5.5 μm) <i>Gephyrocapsa</i> spp.	1.62		
	CN13b/CN13a	B medium (>4 μm) <i>Gephyrocapsa</i> spp. (= bmG event)	1.73		
	1.806 Ma	CN13a			
Gelasian	CN13a/CN12d	NN19/NN18	T <i>Discoaster brouweri</i>	1.93	
	CN12d	NN18	T <i>Discoaster triradiatus</i>	1.95	
			B acme <i>Discoaster triradiatus</i>	2.22	
	CN12d/CN12c	NN18/NN17	T <i>Discoaster pentaradiatus</i>	2.39	
	CN12c/CN12b	NN17/NN16	T <i>Discoaster surculus</i>	2.49	
2.588 Ma	CN12b	NN16			
Piacenzian	CN12b/CN12a		T <i>Discoaster tamalis</i> (subtop)	2.80	
			T <i>Sphenolithus</i> spp. (subtop)	3.54	
3.600 Ma	CN12a				
Zanclean	CN12a/CN11b	NN16/NN15	T <i>Reticulofenestra pseudumbilicus</i>	3.70	
	CN11b	NN15/NN14	T <i>Amaurolithus tricorniculatus</i>	(3.92)	
	CN11b/CN11a	NN14/NN13	B common <i>Discoaster asymmetricus</i>	4.13	
	CN11a/CN10c	NN13	T <i>Amaurolithus primus</i>	4.50	
			B <i>Reticulofenestra pseudumbilica</i> , <i>Discoaster ovata</i> (subbottom)	4.91	
			T <i>Ceratolithus acutus</i>	5.04	
		CN10c/CN10b	NN13/NN12	B <i>Ceratolithus rugosus</i>	5.12
		T <i>Triquetrorhabdulus rugosus</i>	5.28		
5.333 Ma	CN10b	NN12			
CN10b/CN10a			B <i>Ceratolithus acutus</i>	5.35	
CN10a/CN9d	NN12/NN11		T <i>Discoaster quinqueramus</i>	5.59	
CN9d/CN9c			T <i>Nicklithus amplificus</i>	5.94	
	CN9c/CN9b	NN11	B <i>Nicklithus amplificus</i>	6.91	
7.246 Ma	CN9b				
CN9b/CN9a			B <i>Amaurolithus primus</i> , <i>Amaurolithus</i> spp.	7.42	
			T <i>Discoaster loeblichii</i>	7.53	
Tortonian	CN9a	NN10	B common <i>Discoaster surculus</i>	7.79	
			B <i>Discoaster quinqueramus</i>	(8.12)	
	CN9a/CN8		NN11/NN10	B <i>Discoaster berggrenii</i>	8.29
			T <i>Minylitha convallis</i>	8.68	
	CN8	NN10	B <i>Discoaster loeblichii</i>	8.77	
			B paracme <i>Reticulofenestra pseudumbilicus</i>	8.79	
			T <i>Discoaster bollii</i>	9.21	
			B common <i>Discoaster pentaradiatus</i>	9.37	
	CN8/CN7	NN10/NN9	T <i>Discoaster hamatus</i>	9.53	
	CN7	NN9	T <i>Catinaster calyculus</i>	9.67	
			T <i>Catinaster coalitus</i>	9.69	
			B <i>Minylitha convallis</i>	9.75	
			B <i>Discoaster bellus</i>	10.40	
			B <i>Discoaster neohamatus</i>	10.52	
			B <i>Discoaster hamatus</i>	10.55	
			B common <i>Helicosphaera stalis</i>	10.71	
			T common <i>Helicosphaera walbersdorfensis</i>	10.74	
	CN6	NN8	B <i>Discoaster brouweri</i>	10.76	
			B <i>Catinaster calyculus</i>	10.79	
B <i>Catinaster coalitus</i>			10.89		
T <i>Coccolithus miopelagicus</i>			10.97		
CN5b	NN7	T <i>Calcidiscus premacintyreii</i>	11.21		
		T common <i>Discoaster kuqleri</i>	11.58		

Table T1 (continued).

GTS2012 chronostratigraphy	Standard tropical–subtropical biozone (biochron)		Biohorizon (datum)	GTS2012 age (Ma)		
	CN zones (Okada and Bukry, 1980)	NN zones (Martini, 1971)				
11.608 Ma	CN5b	NN7				
Serravallian	CN5b/CN5a	NN7/NN6	T <i>Cyclicargolithus floridanus</i>	11.85		
	CN5a	NN6	B common <i>Discoaster kugleri</i>	11.90		
			T <i>Coronocyclus nitescens</i>	12.12		
			T regular <i>Calcidiscus premacintyre</i>	12.38		
			B common <i>Calcidiscus macintyre</i>	12.46		
			B <i>Reticulofenestra pseudoumbilicus</i>	12.83		
B <i>Triquetrorhabdulus rugosus</i>			13.27			
		T common <i>Cyclicargolithus floridanus</i>	13.28			
		B <i>Calcidiscus macintyre</i>	13.36			
	CN5a/CN4	NN6/NN5	T <i>Sphenolithus heteromorphus</i>	13.53		
13.82 Ma	CN4	NN5				
Langhian	CN4/CN3	NN5/NN4	T <i>Helicosphaera ampliaperta</i>	14.91		
	CN3	NN4	T abundant <i>Discoaster deflandrei</i> group	15.80		
B <i>Discoaster signus</i>			15.85			
15.97 Ma						
Burdigalian			B <i>Sphenolithus heteromorphus</i>	17.71		
	CN3/CN2	NN4/NN3	T <i>Sphenolithus belemnus</i>	17.95		
	CN2/CN1c	NN3/NN2	T <i>Triquetrorhabdulus carinatus</i>	18.28		
(20.44 Ma)	CN1c	NN2	B <i>Sphenolithus belemnus</i>	19.03		
			B <i>Helicosphaera ampliaperta</i>	20.43		
			B common <i>Helicosphaera carteri</i>	22.03		
	CN1c/CN1a-b	NN2/NN1	B <i>Orthorhabdus serratus</i>	22.42		
			B <i>Sphenolithus disbelemnus</i>	22.76		
			B <i>Discoaster druggi</i> (sensu stricto)	22.82		
			T <i>Sphenolithus capricornutus</i>	22.97		
23.03 Ma	CN1a-b	NN1				
Chattian	CN1a-b/CP19b	NN1/NP25	T <i>Sphenolithus delphix</i>	23.11		
	CP19b	NP25	T <i>Reticulofenestra bisecta</i> (>10 μm)	23.13		
			B <i>Sphenolithus delphix</i>	23.21		
			T <i>Zygrhabdulus bijugatus</i>	23.76		
			T <i>Sphenolithus ciperoensis</i>	24.43		
			T <i>Cyclicargolithus abisectus</i> (common)	24.67		
			T <i>Chiasmolithus altus</i>	25.44		
	B <i>Triquetrorhabdulus carinatus</i> (common)	26.57				
	CP19b/CP19a	NP25/NP24	T <i>Sphenolithus distentus</i>	26.84		
			T <i>Sphenolithus predistentus</i>	26.93		
(28.09 Ma)	CP19a	NP24				
Rupelian			T <i>Sphenolithus pseudoradians</i>	28.73		
	CP19a/CP18	NP24/NP23	B <i>Sphenolithus ciperoensis</i>	29.62		
	CP18/CP17	NP23	B <i>Sphenolithus distentus</i>	30.00		
	CP17/CP16c	NP23/NP22	T <i>Reticulofenestra umbilicus</i> (low–mid latitude)	32.02		
	CP16c/CP16b	NP22/NP21	T <i>Coccolithus formosus</i>	32.92		
33.89 Ma	CP16b/CP16a	NP21	T <i>Clausicoccus subdistichus</i> (top of acme)	33.43		
	CP16a					
	CP16a/CP15	NP21/NP20-19	T <i>Discoaster saipanensis</i>	34.44		
	CP15	NP20-19	T <i>Discoaster barbadiensis</i>	34.76		
			T <i>Reticulofenestra reticulata</i>	35.40		
			B <i>Isthmolithus recurvus</i>	36.97		
		NP20-19/NP18	B <i>Chiasmolithus oamaruensis</i> (common)	37.32		
(37.75 Ma)						
Bartonian	CP15/CP14b	NP17	T <i>Chiasmolithus grandis</i>	37.98		
	CP14b		B <i>Chiasmolithus oamaruensis</i> (rare)	38.09		
			B <i>Reticulofenestra bisecta</i> (>10 μm)	38.25		
	CP14b/CP14a	NP17/NP16	T <i>Chiasmolithus solitus</i>	40.40		

distinguished three *Reticulofenestra* morphotypes, *Reticulofenestra ampla* (5–7 μm , with central opening), *R. bisecta* (5–10 μm , with a solid central plug), and *Reticulofenestra stavensis* (>10 μm , with a solid central plug), following Sato et al. (1991) and Young et al. (2014).

The LAD of *Sphenolithus* spp. (3.54 Ma) in Pliocene Zone NN16 was based on the LAD of *Sphenolithus abies* and *Sphenolithus neoabies* according to Raffi et al. (2006). Species concepts for other taxa mainly follow those of Perch-Nielsen (1985) and Bown (1998).

Methods

Calcareous nannofossil samples were prepared using standard smear slide techniques. In cases with sandy sediments, suspended aliquots of the raw sample were utilized for analysis. Samples were examined with a Zeiss microscope under crossed polarized and plane-transmitted or phase contrast light at 1000 $\times$ to 2000 $\times$ magnification. Preservation of nannofossils was noted as follows:

VG = very good (no evidence of dissolution and/or overgrowth).

G = good (slight dissolution and/or overgrowth; specimens are identifiable to the species level).

M = moderate (some etching and/or overgrowth; most specimens are identifiable to the species level).

P = poor (severely etched or with overgrowth; most specimens cannot be identified at the species and/or generic level).

The relative abundance of calcareous nannofossils in the sediment was visually estimated at 1000 $\times$ magnification by referring to the particle abundance charts in Rothwell (1989) and reported using the following abundance categories:

D = dominant (>90% of sediment particles).

A = abundant (>50%–90% of sediment particles).

C = common (>10%–50% of sediment particles).

F = few (1%–10% of sediment particles).

R = rare (<1% of sediment particles).

B = barren (no nannofossils present in 100 fields of view [FOVs]).

The relative abundance of individual calcareous nannofossil species or taxa groups was estimated at 1000 $\times$ magnification as follows:

D = dominant (>50%, or 100 specimens per FOV).

A = abundant (10%–50%, or 10,100 specimens per FOV).

C = common (1%–10%, or 110 specimens per FOV).

F = few (0.1%–1%, or 1 specimen per 110 FOVs).

R = rare (<0.1%, or <1 specimen per 10 FOVs).

Planktonic foraminifers

The planktonic foraminiferal zonation schemes of Blow (1969, 1979) and Berggren et al. (1995), as modified by Wade et al. (2011),

were used in this study. Calibrated ages for bioevents are from Gradstein et al. (2012), as given in Table T2. We adopted the use of the LAD of *Paragloborotalia nana* as an early Miocene (19.30–21.12 Ma; Leckie et al., 2018) biostratigraphic indicator. Taxonomic concepts for Neogene and Paleogene taxa mainly follow those of Kennett and Srinivasan (1983) and Bolli and Saunders (1985).

Core catcher samples were soaked in distilled water and washed over a 63 μm mesh sieve. Consolidated or lithified samples were cut into pieces and crushed to pea size, added to a hydrogen peroxide solution, heated in the oven at <50°C for several hours, and then sieved as above. All samples were dried in a low-temperature oven at ~50°C. The dried samples were sieved over a 150 μm sieve, retaining the <150 μm size fraction in a separate vial. To avoid contamination of foraminifers between samples, the sieves were thoroughly cleaned between samples, placed in a sonicator for at least 15 min, and then carefully checked for the presence of sediment particles. Species identifications for planktonic and benthic foraminifers were generally made on the >150 μm size fraction.

The total abundance of planktonic foraminifers was defined as follows:

A = abundant (>30% planktonic foraminifer specimens in total residue).

C = common (10%–30% planktonic foraminifer specimens in total residue).

R = rare (1%–10% planktonic foraminifer specimens in total residue).

P = present (<1% planktonic foraminifer specimens in total residue).

B = barren (no planktonic foraminifer specimens in total residue).

Individual planktonic foraminifers were recorded in qualitative terms based on an assessment of forms observed in a random sample of ~400 specimens from the >150 μm size fraction. Relative abundances were reported using the following categories:

D = dominant (>30% of the assemblage).

A = abundant (10%–30%).

F = few (5%–10%).

R = rare (1%–5%).

P = present (<1%).

Preservation of planktonic foraminifer assemblages was recorded using the following categories:

VG = very good (no evidence of breakage or dissolution).

G = good (>80% of specimens are unbroken with only minor evidence of diagenetic alteration).

M = moderate (30%–80% of the specimens are unbroken).

P = poor (strongly recrystallized or dominated by fragments and broken or corroded specimens).

Table T2. Planktonic foraminiferal events and scaled ages (Gradstein et al., 2012) used during Expedition 368X. GTS2012 = 2012 geologic timescale. T = top/last appearance; B = base/first appearance. Bold = age-diagnostic datum. (Continued on next two pages.) [Download table in CSV format.](#)

Standard tropical–subtropical biozone (biochron)						
GTS2012 chronostratigraphy	Indo-Pacific (Blow, 1969, 1979; Berggren et al., 1995)	Indo-Pacific (Berggren et al., 1995; Wade et al., 2011)	Biohorizon (datum)	GTS2012 age (Ma)	Error (My)	
Tarantian (late Pleistocene)	N22	PT1b	T <i>Globorotalia flexuosa</i>	0.07		
0.126 Ma			T <i>Globigerinoides ruber</i> pink	0.12		
Ionian (middle Pleistocene)			B <i>Globigerinella calida</i>	0.22		
			Bc <i>Globigerinoides ruber</i> pink	0.40		
			B <i>Globorotalia flexuosa</i>	0.40		
			B <i>Globorotalia hirsuta</i>	0.45		
			T <i>Globorotalia tosaensis</i>	0.61		
0.781 Ma			B <i>Globorotalia hessi</i>	0.75		
1.806 Ma			B <i>Globigerinoides ruber</i> pink	1.16		
			T <i>Globigerinoides obliquus</i>	1.3	±0.1	
		T <i>Neogloboquadrina acostaensis</i>	1.58	±0.03		
		T <i>Globoturborotalita apertura</i>	1.64	±0.03		
Gelasian (early Pleistocene)		PT1a	PT1a/PL6	T <i>Globigerinoides fistulosus</i>	1.88	±0.03
		PL6/PL5	T <i>Globigerinoides extremus</i>	1.98	±0.03	
			B <i>Pulleniatina finalis</i>	2.04	±0.03	
			T <i>Globorotalia multicamerata</i>	2.18		
			T <i>Globorotalia pertenuis</i>	2.30		
			T <i>Globoturborotalita woodi</i>	2.30	±0.02	
2.588 Ma		N22/N21		T <i>Globorotalia pseudomiocenica</i>	2.39	
Piacenzian (late Pliocene)	N21	PL5/PL4	T <i>Globoturborotalita decoraperta</i>	2.75	±0.03	
	N21/N19–N20		B <i>Globigerinoides fistulosus</i>	3.33		
	PL4/PL3		B <i>Globorotalia tosaensis</i>	3.35		
			T <i>Dentoglobigerina altispira</i>	3.47		
			B <i>Globorotalia pertenuis</i>	3.52	±0.03	
3.600 Ma			T <i>Sphaeroidinellopsis seminulina</i>	3.59		
Zanclean (early Pliocene)	PL3	T <i>Pulleniatina primalis</i>	3.66			
		T <i>Globorotalia plesiotumida</i>	3.77	±0.02		
		T <i>Globorotalia margaritae</i>	3.85	±0.03		
		T <i>Pulleniatina spectabilis</i>	4.21			
		B <i>Globorotalia crassaformis</i> sensu lato	4.31	±0.04		
	PL2/PL1	T <i>Globoturborotalita nepenthes</i>	4.37	±0.01		
		B <i>Globorotalia exilis</i>	4.45	±0.04		
		T <i>Sphaeroidinellopsis kochi</i>	4.53	±0.17		
		T <i>Globorotalia cibaoensis</i>	4.60			
		T <i>Globigerinoides seiglei</i>	4.72			
5.333 Ma						
Messinian (late Miocene)	N19–20/N18	PL1/M14	B <i>Sphaeroidinella dehiscens</i> sensu lato	5.53	±0.04	
	N18/N17b		B <i>Globorotalia tumida</i>	5.57		
	M14/M13b		B <i>Turborotalita humilis</i>	5.81	±0.17	
			T <i>Globoquadrina dehiscens</i>	5.92		
			B <i>Globorotalia margaritae</i>	6.08	±0.03	
		N17b/N17a	T <i>Globorotalia lenguaensis</i>	6.14		
	Tortonian (late Miocene)	M13b	B <i>Globigerinoides conglobatus</i>	6.20	±0.41	
			B <i>Pulleniatina primalis</i>	6.60		
			B <i>Globorotalia miotumida</i> (conomiozea)	7.89		
			B <i>Neogloboquadrina humerosa</i>	8.56		
B <i>Globorotalia plesiotumida</i>			8.58	±0.03		
N17a/N16		M13b/M13a	B <i>Globigerinoides extremus</i>	8.93	±0.03	
N16		B <i>Globorotalia cibaoensis</i>	9.44	±0.05		
		B <i>Globorotalia juanai</i>	9.69	±0.26		
		B <i>Neogloboquadrina acostaensis</i>	9.83	±0.06		
		T <i>Globorotalia challengerii</i>	9.99			
	N16/N15	M13a/M12	T <i>Paragloborotalia mayeri/siakensis</i>	10.46	±0.02	
N15/N14	M12/M11	B <i>Globorotalia limbata</i>	10.64	±0.26		
		T <i>Cassigerinella chipolensis</i>	10.89			
		B <i>Globoturborotalita apertura</i>	11.18	±0.13		
		B <i>Globorotalia challengerii</i>	11.22			
		B regular <i>Globigerinoides obliquus</i>	11.25			
11.608 Ma			B <i>Globoturborotalita decoraperta</i>	11.49	±0.04	
			T <i>Globigerinoides subquadratus</i>	11.54		

Table T2 (continued). (Continued on next page.)

GTS2012 chronostratigraphy	Standard tropical–subtropical biozone (biochron)		Biohorizon (datum)	GTS2012 age (Ma)	Error (My)
	Indo-Pacific (Blow, 1969, 1979; Berggren et al., 1995)	Indo-Pacific (Berggren et al., 1995; Wade et al., 2011)			
Serravallian (middle Miocene)	N14/N13	M11/M10	B Globoturborotalita nepenthes	11.63	±0.02
	N13/N12	M10/M9b	T Fohsella fohsi, Fohsella plexus	11.79	±0.15
			T Clavatorella bermudezi	12.00	
			B Globorotalia languanensis	12.84	±0.05
			B Sphaeroidinellopsis subdehiscens	13.02	
		M9b/M9a	B Fohsella robusta	13.13	±0.02
		M9a	T Cassigerinella martinezpicoi	13.27	
	N12/N11	M9a/M8	B Fohsella fohsi	13.41	±0.04
Langhian (middle Miocene)	N11	M8	B Neogloboquadrina nympha	13.49	
	N11/N10	M8/M7	B Fohsella praefohsi	13.77	
			T Fohsella peripheroronda	13.80	
	13.82 Ma	N10	M7		
			T regular Clavatorella bermudezi	13.82	
			T Globorotalia archeomenardii	13.87	
	N10/N9	M7/M6	B Fohsella peripheroacuta	14.24	
			B Globorotalia praemenardii	14.38	
Burdigalian (early Miocene)			T Praeorbulina sicana	14.53	
			T Globigerinella insueta	14.66	
			T Praeorbulina glomerosa sensu stricto	14.78	
			T Praeorbulina circularis	14.89	
	N9/N8	M6/M5b	B Orbulina suturalis	15.10	
			B Clavatorella bermudezi	15.73	
			B Praeorbulina circularis	15.96	
	15.97 Ma	N8	M5b		
Aquitanian (early Miocene)			B Globigerinoides diminutus	16.06	
			B Globorotalia archeomenardii	16.26	
		M5b/M5a	B Praeorbulina glomerosa sensu stricto	16.27	
		M5a	B Praeorbulina curva	16.28	
	N8/N7	M5a/M4b	B Praeorbulina sicana	16.38	
		M4b	T Globorotalia incognita	16.39	
		M4b/M4a	B Fohsella birnageae	16.69	
			B Globorotalia miozea	16.70	
			B Globorotalia zealandica	17.26	
		M4a	T Globorotalia semivera	17.26	
	N7/N6	M4a/M3	T Catapsydrax dissimilis	17.54	
			B Globigerinella insueta sensu stricto	17.59	
Chattian (late Oligocene)			B Globorotalia praescitula	18.26	
			T Globoquadrina binaiensis	19.09	
		M3/M2	B Globigerinatella sp.	19.30	
			B Globoquadrina binaiensis	19.30	
			B Globigerinoides altiapturus	20.03	
	20.44 Ma	N6–N5	M2		
			T Tenuitella munda	20.78	
			B Globorotalia incognita	20.93	
Rupelian (early Oligocene)			T Globoturborotalita angulisuturalis	20.94	
	N5/N4b	M2/M1b	T Paragloborotalia kugleri	21.12	
			T Paragloborotalia pseudokugleri	21.31	
			B Globoquadrina dehiscens forma spinosa	21.44	
			T Dentoglobigerina globularis	21.98	
	N4b/N4a	M1b/M1a	B Globoquadrina dehiscens	22.44	
			T Globigerina ciperoensis	22.90	
			B Globigerinoides trilobus sensu lato	22.96	
Rupelian (early Oligocene)	N4a/P22	M1a/O7	B Paragloborotalia kugleri	22.96	
	23.03 Ma				
			T Globigerina euapertura	23.03	
			T Tenuitella gemma	23.50	
			B common Globigerinoides primordius	23.50	
		O7/O6	B Paragloborotalia pseudokugleri	25.21	
		O6	B Globigerinoides primordius	26.12	
	P22/P21	O6/O5	T Paragloborotalia opima sensu stricto	26.93	
Rupelian (early Oligocene)	28.09 Ma	P21 (N2)	O5/O4		
			T common Chiloguembelina cubensis	28.09	
	P21/P20	O4/O3	B Globigerina angulisuturalis	29.18	
			B Tenuitellinata juvenilis	29.50	
Rupelian (early Oligocene)		P20	O3		
			T Subbotina angiporoides	29.84	
	P20/P19	O3/O2	T Turborotalia ampliapertura	30.28	

Table T2 (continued).

Standard tropical–subtropical biozone (biochron)					
GTS2012 chronostratigraphy	Indo-Pacific (Blow, 1969, 1979; Berggren et al., 1995)	Indo-Pacific (Berggren et al., 1995; Wade et al., 2011)	Biohorizon (datum)	GTS2012 age (Ma)	Error (My)
Rupelian (early Oligocene)	P19	O2	B <i>Paragloborotalia opima</i>	30.72	
	P19/P18	O2/O1	T <i>Pseudohastigerina nagewichiensis</i>	32.10	
		O1	B <i>Cassigerinella chipolensis</i>	33.89	
33.89 Ma	P18	O1/E16			
Priabonian (late Eocene)			T <i>Hantkenina</i> spp.	33.89	
		T common <i>Pseudohastigerina micra</i>	33.89		
	P18/P17	E16	T <i>Turborotalia cerroazulensis</i>	34.03	
	P17/P16		T <i>Cribohantkenina inflata</i>	34.22	
		E16/E15	T <i>Globigerinatheka index</i>	34.61	
	P16	E15	T <i>Turborotalia pomeroli</i>	35.66	
			B <i>Turborotalia cunialensis</i>	35.71	
	P16/P15		B <i>Cribohantkenina inflata</i>	35.87	
(37.75 Ma)	P15	E15/E14	T <i>Globigerinatheka semiinvoluta</i>	36.18	
Bartonian					
		E14	T <i>Acarinina</i> spp.	37.75	

Table T3. Sampling protocol for hard rock discrete samples, Expedition 368X. AMS = anisotropy of magnetic susceptibility, NRM = natural remanent magnetism, AF = alternating field, TSB = thin-section billet, XRD = X-ray diffraction, ICP = inductively coupled plasma, Pmag = paleomagnetism. [Download table in CSV format.](#)

Sample type	Team	Treatment
CUBE	Paleomagnetism	AMS, NRM, and AF demagnetization
CUBE	Petrophysics	P-wave velocity on wet sample Split to two residues
RESIDUE 1 (upper half)	Core describers or Paleomagnetism	Shipboard TSB, XRD, ICP, or postcruise Pmag
RESIDUE 2 (lower half)		
If XRD, ICP were not run on RESIDUE 1	Petrophysics	Measured for density and porosity
If XRD, ICP were run on RESIDUE 1		Split to two residues
RESIDUE 2a	Petrophysics	Measured for density and porosity
RESIDUE 2b	Paleomagnetism	Postcruise

Paleomagnetism

Shipboard paleomagnetic investigations were conducted following the same methods applied during Expeditions 367 and 368 (see the [Expedition 367/368 methods](#) chapter [Sun et al., 2018]). During Expedition 368X, some differences in methods for discrete samples, including the sampling space and the demagnetization steps, were used and are therefore described here.

Usually, only one discrete cube sample was collected from each core to perform alternating field (AF) demagnetization, depending on recovery rate and lithology. No samples were collected for thermal demagnetization because of the time necessary to complete those measurements and the limited duration of Expedition 368X.

Basalt samples were shared with the petrophysics and core description teams to minimize the number of samples collected, optimize data processing time, and maximize the number of residuals (e.g., not heating residuals for rock magnetic analysis). A protocol modified after Expeditions 367 and 368 (see the [Expedition 367/368 methods](#) chapter [Sun et al., 2018]) was adopted (Table T3).

The characteristic remanent magnetization (ChRM) directions were calculated by principal component analysis (PCA; Kirschvink, 1980) using the Remasoft 3.0 software. Data visualizations (demag-

netization plots; Zijderveld, 1967) and equal area projections were plotted using the PuffinPlot software (version 1.03; 23 April 2015; Lurcock and Wilson, 2012).

Magnetic measurements on archive section halves

Measurement of archive section halves was conducted using the superconducting rock magnetometer (SRM) and IMS-SRM software (version 10.2). A reduced sample area of 13.4 cm² was assumed for conversion to volume-normalized magnetization units (A/m) of cores sampled with the rotary core barrel (RCB) system. We performed successive AF demagnetization using the SRM in-line AF demagnetizer on all archive section halves except for cores with <2% recovery (only the core catcher) and intervals that were too highly fractured to collect accurate measurements.

Natural remanent magnetization (NRM) measurements were made every 2.5 cm for sedimentary material and every 2.0 cm for igneous rocks. After NRM measurements, sediment cores were subjected to stepwise in-line AF demagnetization at 5, 10, 15, 20, and 25 mT. For igneous rocks, we adopted the same set of AF demagnetization steps used during Expeditions 367 and 368, which includes using narrower AF steps at an incremental rate of 2 mT for NRM up to 10 mT and at a rate of 5 mT from 10 to 25 mT.

Magnetic measurements on discrete samples

Incremental AF demagnetization was performed with the DTech AF demagnetizer (model D-2000) for the JR-6A spinner magnetometer (steps at 2.5, 5, 7.5, 10, 15, 20, 25, 30, 35, 40, 50, 60, 70, and 80 mT). The demagnetization axes were inverted at each step to avoid the gyroremanent magnetization documented during Expeditions 367 and 368 (see the [Expedition 367/368 summary](#) chapter [Larsen et al., 2018]). For basalts, the same set of AF demagnetization steps used for archive section halves was adopted, with further steps at an incremental rate of 5 mT up to 40 mT until the demagnetization was completed.

Geochemistry

The shipboard geochemistry program for Expedition 368X included measurements of headspace gas content; sedimentary geochemistry, including total inorganic carbon, total carbon, total nitrogen, and major and minor element content; and igneous and metamorphic rock geochemistry (major and minor element content).

Our analytical procedures followed those described in the [Expeditions 367/368 methods](#) chapter (Sun et al., 2018). Our analyses were conducted to satisfy routine shipboard safety and pollution prevention requirements, characterize sediment and rock geochemistry for shipboard interpretation, and provide a basis for sampling for subsequent shore-based research.

Headspace gas analysis

During Expedition 368X, headspace gas samples were frequently taken from the first or second section of cores because of low recovery or lack of a third section.

Interstitial water chemistry

No interstitial water samples were obtained during Expedition 368X.

Sediment geochemistry

Sediment sulfur, nitrogen, and inorganic and organic carbon contents

Sulfur analyses were carried out during Expeditions 368 and 368X. Total organic and inorganic carbon, total nitrogen, and total sulfur were measured during Expedition 368X.

Elemental analysis of bulk sediment/sedimentary rock by ICP-AES

The elemental compositions of sediment/sedimentary rock were only determined during Expedition 368 because the bead maker used for preparing inductively coupled plasma–atomic emission spectroscopy (ICP-AES) samples did not work during Expedition 367. During Expedition 368X, discrete samples were collected for shipboard XRD analysis and collocated samples were retained for shore-based analysis for concentrations of major elements and several trace elements by ICP-AES.

Source rock analysis

During Expedition 368X, source rock analyses were conducted to identify the type and stage of maturation of organic matter, estimate total organic carbon (TOC), and detect petroleum potential in sediments. Using source rock pyrolysis, free and adsorbed hydrocarbons released during programmed heating of a sample are re-

corded in a pyrogram as the first peak (S1) under low temperature. The second peak (S2) represents hydrocarbons released by kerogen cracking. The temperature at the maximum of the S2 peak ($T_{\max}$) is an indicator of rock maturity. CO₂ (third peak; S3) is also generated by kerogen degradation. When these components are normalized to the TOC content, the S2 peak becomes the hydrogen index ($HI = S2 \times 100/TOC$) and S3 becomes the oxygen index ($OI = S3 \times 100/TOC$) (Tissot and Welte, 1984).

Source rock pyrolysis and TOC were determined by a Weatherford source rock analyzer. A crucible sample containing no material was included as the first sample of any sequence. After this calibration blank, 90–100 mg of standard material was used to calibrate the instrument signals and allow us to monitor instrument accuracy and precision. A pyrolysis program starting at 300°C with a heating rate of 25°C/min was used as a standard mode for the analysis of sediment with low maturity.

Physical properties

Measurements of physical properties in the Expedition 368X cores followed the same methodology as for Expeditions 367 and 368 (see the [Expedition 367/368 methods](#) chapter [Sun et al., 2018]). Only a few procedures were adjusted during Expedition 368X, and they are described below.

Hard rock cores

Recovered hard rock sections were run through the Whole-Round Multisensor Logger (WRMSL) and Natural Gamma Radiation Logger (NGRL) after the core sections reached equilibrium with laboratory temperature, which typically took 2 h during Expedition 368X because of the great depth of the cores below seafloor.

Samples from igneous basement were shared for both paleomagnetic and moisture and density measurements. The flow of sample sharing from Expeditions 367 and 368 was optimized for Expedition 368X (see [Paleomagnetism](#); Table T3).

Thermal conductivity measurements

During Expedition 368X, we measured thermal conductivity on samples from working section halves using contact probes of different sizes to optimize the measurements according to the size of the piece of rock. After the rock and probe were equilibrated together in a bath of seawater at room temperature in a cooler insulated with extruded polystyrene foam, the calibrated heat source of the probe was turned on and the increase in temperature was recorded over 80 s for the standard probe and 60 s for the small probe.

Thermal conductivity was calculated from the rate of temperature rise while the heater current was flowing. Temperatures measured during the first 80 or 60 s of the heating cycle were fitted to an approximate solution of a constantly heated line source (for details, see Kristiansen, 1982, and Blum, 1997). Measurement errors were 5%–10%. Thermal conductivity measurements were routinely taken in one section per core (usually the third).

Discrete sample moisture and density measurements

Discrete samples were collected from the working section halves to determine wet and dry bulk density, grain density, water content, and porosity. In hard rock, cubes were extracted from the working section halves for physical property measurements. Samples from igneous basement were shared for paleomagnetic measurements.

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