

Phylogeny and taxonomy of *Acer* powdery mildews, including genera *Sawadaea* and *Takamatsuella* (Erysiphaceae, Ascomycota)

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Abstract: *Acer* (Sapindaceae) is a major genus of broadleaf trees dominating deciduous forests in the Northern Hemisphere, with Asia exhibiting the highest species diversity. Many economically important *Acer* species are cultivated for ornamental or timber purposes. *Acer* powdery mildew, caused by fungi in the tribe Cystothecae, poses significant global economic and ecological threats. The pathogenicity spectrum remains unclear due to taxonomic uncertainties in its primary causal genera, *Sawadaea* and *Takamatsuella*. This study presents a comprehensive phylogenetic-taxonomic analysis of the two genera across East Asia, Europe, and North America. Using 75 ITS and 58 28S rDNA newly obtained sequences, we resolved 12 *Sawadaea* species and one *Takamatsuella* species into nine monophyletic clades, revealing marked cryptic diversity (three new species: *S. acerina*, *S. aceris-arguti*, *S. taii*) and two paraphyletic groups (*S. bifida*/*S. negundinis*). Taxonomic revisions include: *S. bicornis* split into two *formae* (*f. bicornis* and *f. polyphaga f. nov.*) with distinct host preferences; *S. tulasnei* (sensu stricto) restricted to Europe/North America, invalidating previous Asian records; *S. nankinensis* and *S. koelreuteriae* form two basal lineages. Phylogenetic positioning confirmed *Takamatsuella* as a distinct genus sister to *Sawadaea*, supported by an ITS1 26 bp deletion. Host specificity analysis revealed narrow host ranges (primarily *Acer*) with two evolutionary host expansions to *Koelreuteria*, *Aesculus*, and *Liquidambar*. This study also newly describes the asexual morphs of four species (*S. aesculi*, *S. bifida*, *S. bomiensis* and *S. kovaliana*) and establishes a molecular framework for disease management through clarified phylogeny and taxonomy. Our findings provide critical insights into fungal evolution, host-pathogen interactions, and strategies for mitigating powdery mildew impacts in forest ecosystems.

Key words: Diversity, Erysiphaceae, *formae*, Helotiales, new species.

Taxonomic novelties: New *forma*: *Sawadaea bicornis f. polyphaga* M. Bradshaw & U. Braun. New species: *Sawadaea acerina* G.X. Guan & S.Y. Liu, *Sawadaea aceris-arguti* S. Takam. & U. Braun, *Sawadaea taii* G.X. Guan & S.Y. Liu.

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INTRODUCTION

Acer (Sapindaceae, incl. Aceraceae) is an important broad leaves genus that prominent deciduous forests of the Northern Hemisphere, and comprises about 125 to 156 species based on the applied species concepts. Asia harbours the highest species diversity, where multiple species are important economically in that they are planted as ornamental trees or for timber (Gao *et al.* 2020). East Asia, especially China is considered the present centre of diversification for *Acer* (Suh *et al.* 2000). The taxonomy of *Acer* species is well documented (Li *et al.* 2006, Grimm *et al.* 2007), whereas Li *et al.* (2019) provided a modern classification of *Acer* into sections, substantiated by phylogenetic data, which

largely coincides with previous subgeneric concepts (de Jong 2002). Microfungi associated with *Acer* species have diverse host-plant interactions. Among these phytopathogenic ascomycetes, including powdery mildews is one of the important diseases on acer plants (Johnston *et al.* 2019, Haelewaters *et al.* 2021).

Powdery mildews are obligate biotrophic fungi forming characteristic, conspicuous symptoms caused by whitish superficial mycelial patches or films of the sexual morph (Braun & Cook 2012). They are cosmopolitan in distribution and occur on more than 10,000 plant species, including numerous important crops (Amano 1986, Braun & Cook 2012; Bradshaw *et al.* 2022). The currently accepted generic system of powdery mildews in the world containing five tribes, including 17 sexual morph genera and two asexual morph

Ucinula circinata *Dodonaea viscosa* *Sapindus*
drummondii *S. marginatus*
Sapindus
 deposited at BPI, have been examined by U. Braun and confirmed
E. flexuosa
Takamatsuella *Sawadaea* and its affiliation to tribe *Cystothecaceae*
 have been insufficiently examined so far, and require further
 research. Therefore, the clarifications of these problems have been
Sawadaea
Takamatsuella
Sawadaea
Takamatsuella
Sawadaea *Takamatsuella*

MATERIALS AND METHODS

Fungal materials

countries. In total, 123 specimens from five countries (China,
 Of these, 62 specimens of *Sawadaea* *Takamatsuella*

Morphological examination

DNA extraction, PCR amplification and sequencing

spacer (ITS) regions, including the 5.8S rDNA, were amplified
 amplified with primers pairs LSU1/LSU2 (Scholin *et al.*
 components were 2 µL of total genomic DNA, 2.5 µL 10× PCR buffer
 (2.5 mM each), 1 µL each primer (20 ng/µL), 0.1 µL *Taq*
 (TaKaRa, Japan) (5 U/µL) and sterile ddH₂O up to a final volume
 of 25 µL. The amplification reactions were conducted under the
 52–56 °C for annealing, and 30 s at 72 °C for extension, and a final
 gel in 0.5× TBE buffer. The amplicons were sent to Sangon Biotech

Phylogenetic analyses

additions to find the global optimum tree. All gaps were treated as
 both analyses. For BI analysis, the best-fit substitution models
 and trees were sampled every 100 generations. The first 25 %

RESULTS

The phylogeny of *Sawadaea* and *Takamatsuella*

Seventy-five ITS sequences including the ITS1-5.8S rDNA-ITS2 and 58 28S rDNA sequences including D1/D2 domains were generated in this study and were aligned with sequences retrieved from the NCBI GenBank nucleotide database. The ITS+28S rDNA sequence alignment matrix consisted of 203 sequences including 950 characters, of which 168 (17.7 %) were parsimony informative

and 18 (1.2 %) were parsimony-uninformative. *Cystotheca castanopsidis* and *C. wrightii* were selected as outgroups. The maximum parsimony (MP) and maximum likelihood (ML) tree with the highest likelihood value of more than 70 % and posterior probabilities value greater than 0.90 for Bayesian Inference (BI) are shown in Fig. 1. Phylogenetic trees generated from ML and BI analyses are almost identical to the MP tree, which is, therefore, not shown.

The tree topology revealed the molecular phylogeny among species in *Sawadaea* and a much higher species diversity than



Fig. 1. Phylogenetic analysis of the ITS+28S rDNA regions. Nodes are labelled with bootstrap values from Parsimony bootstrap/Maximum likelihood/Bayesian posterior probabilities values. Posterior probabilities greater than bootstrap support values (> 70 %) by maximum parsimony (MP), maximum likelihood (ML) and 0.90 for Bayesian Inference (BI) methods are shown on the respective branches. The newly generated sequences in this study are in bold. *Cystotheca castanopsidis* and *C. wrightii* are used as outgroup taxa.

previously assumed, especially in Eastern Asia. Twelve species of *Sawadaea* and one species of *Takamatsuella* formed nine monophyletic clades (clades 1–5, 7, 9–11) and two non-monophyletic clades (clade 6 and clade 8). The insertion/deletion differences between sites 60 and 107 of the ITS1 region were summarized among these clades (Fig. 2). *Takamatsuella circinata* (Clade 1; MP = 100 %, ML = 100 %, BI = 1.00) formed a sister clade to genus *Sawadaea* with high support which is mainly caused by a 26 bp deletion between sites 64 and 89 of the ITS1 region. This result supports the treatment of *Takamatsuella* as an independent genus. The differences among *Sawadaea* species were caused mainly by a 6–14 bp insertion/deletion (indel) between sites 65 and 101 of

the ITS1 region. *Sawadaea nankinensis* (Clade 2), *S. koelreuteriae* (Clade 3), *S. aesculi* (Clade 4), *S. negundinis* (Clade 6), *S. bifida* (Clade 6), *S. tulasnei* (Clade 7), *S. taii* (Clade 8) and *S. kovaliana* (Clade 8) had 10 bp deletions. *Sawadaea nankinensis* formed a separate clade (Clade 2; MP = 100 %, ML = 100 %, BI = 1.00) with the other species of *Sawadaea*, which was mainly caused by the two bases insertion at sites 65 and 66 and one base deletion at site 89 (Fig. 2), and its distinguished unbranched uncinete-circinate appendages (Fig. 3). *Sawadaea koelreuteriae* formed a separate clade with high support (MP = 100 %, ML = 100 %, BI = 1.00). *Sawadaea aceris-arguti* has a 6 bp deletion between sites 93 and 98, and *S. acerina* has a 7 bp deletion between sites 94 and 100.

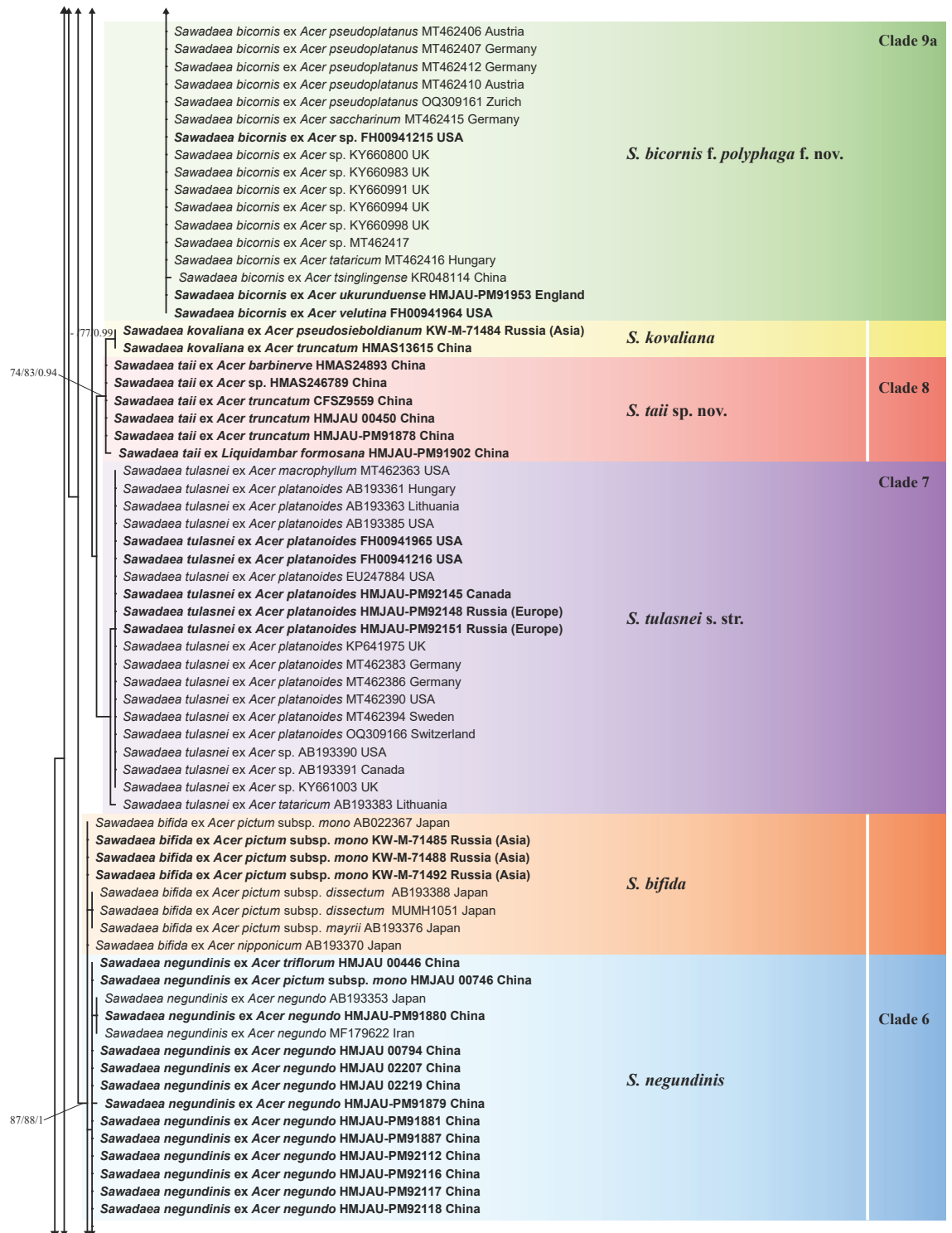


Fig. 1. (Continued)

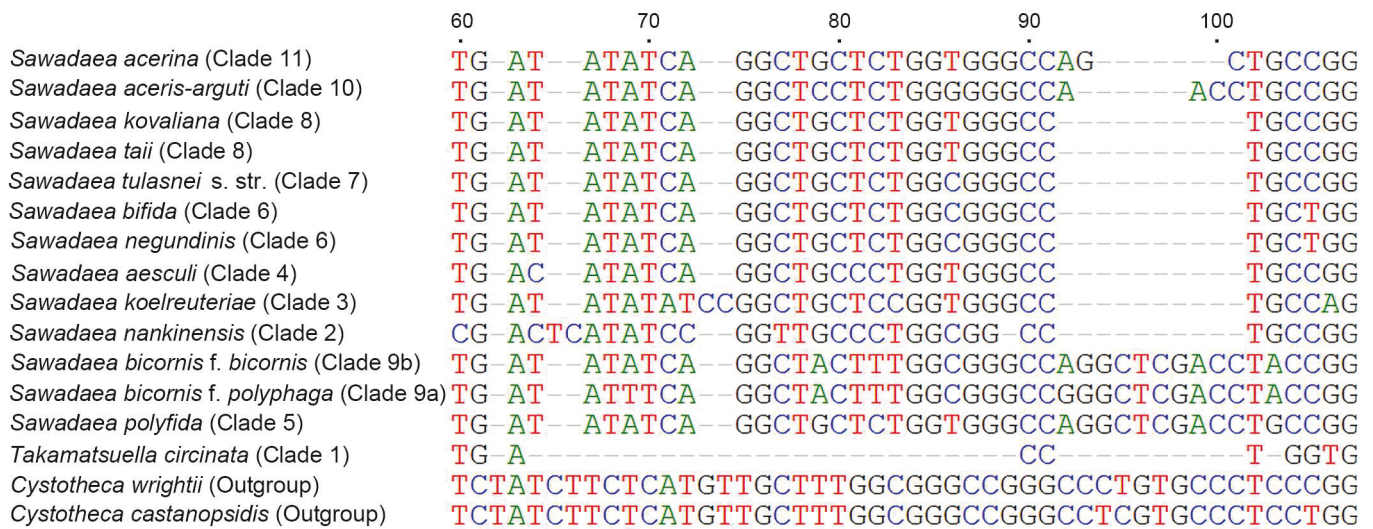


Fig. 2. Insertion/deletion (indel) found between sites 60 and 107 of ITS1 regions of *Sawadadea*, *Takamatsuella* and *Cystotheca*. The insertion or deletion is related to the formation of clades in the phylogenetic tree and the presentation of phylogenetic relationships.

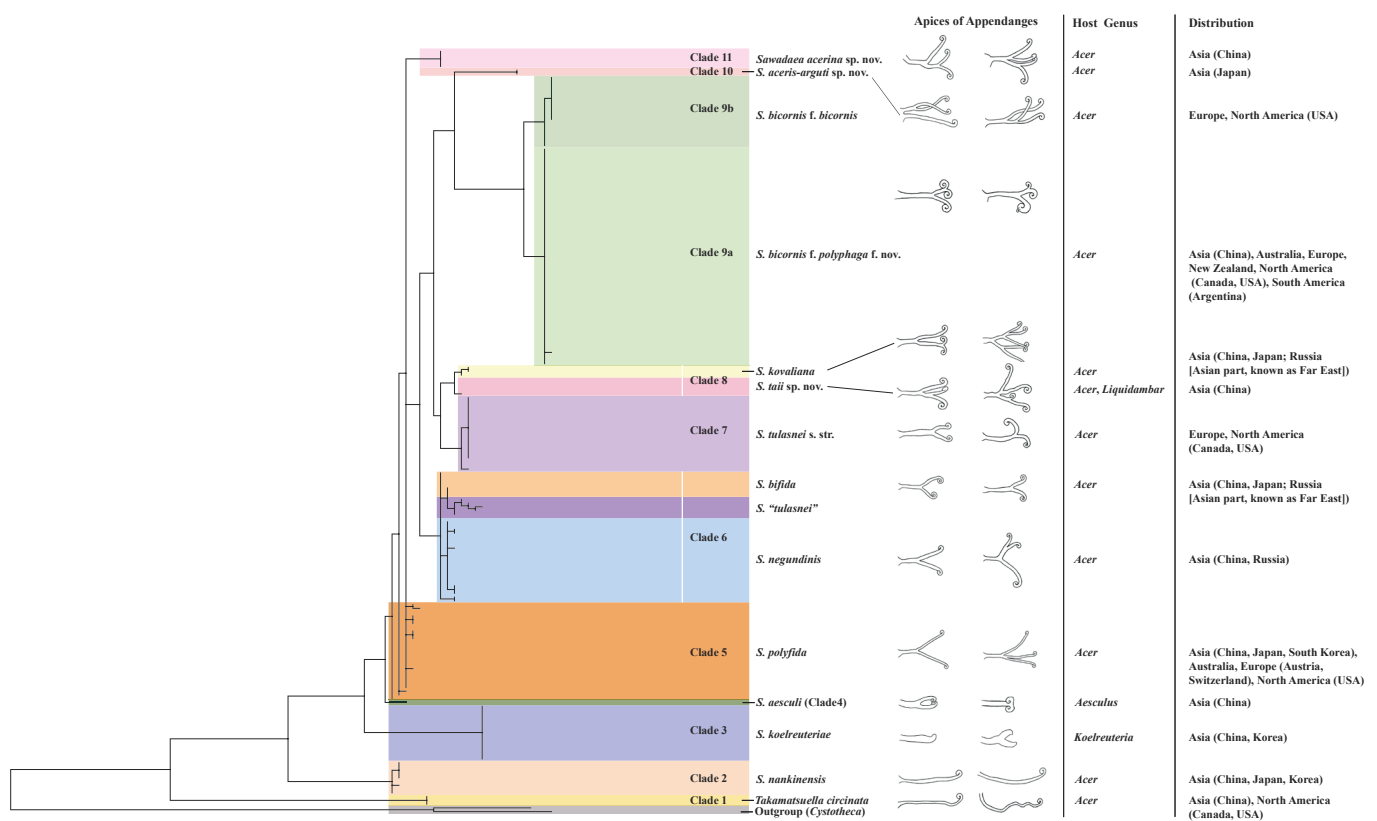


Fig. 3. Phylogenetic outline of the ITS+28S rDNA regions combining typical characteristics of appendages on chasmothecia.

Taxonomy

Twelve species of *Sawadadea* and one species of *Takamatsuella* were recognized, including three new species, namely *Sawadadea acerina* sp. nov., *Sawadadea aceris-arguti* sp. nov., and *Sawadadea taii* sp. nov. described from Asia. *Sawadadea bicornis* comprises two biological formae, *Sawadadea bicornis* f. *bicornis* and f. *polyphaga* f. nov., with different host preferences. Asexual morphs of *S. aesculi*, *S. bifida*, *S. kovaliana* and *T. circinata* have also been detected and described for the first time.

























































































Sawadadea Miyake, *Special Bull Agric. Exp. Stat. Formosa* **9**: 49. 1914.

Synonyms: *Uncinula* sect. *Sawadadea* (Miyake) U. Braun, *Feddes Rept.* **88**: 663. 1978.

Oidium subgen. *Octagoidium* R.T.A. Cook et al., *Mycol. Res.* **101**: 998. 1997 [type species: *Oidium aceris* Rabenh.].

Octagoidium (R.T.A. Cook et al.) R.T.A. Cook & U. Braun, in Braun & Cook, *Taxonomic manual of the Erysiphales (powdery mildews)*: 172. 2012.

Table 1. 


Fungal species ^a	Host	Specimen No. ^b	Collection locality	GenBank accession number	
				ITS	28S rDNA
Sawadaea acerina sp. nov.	<i>Acer tataricum</i> 				
	<i>A. tataricum</i> 			—	OQ866185
	<i>A. tataricum</i> 				
	<i>A. tataricum</i> 			—	OQ866186
	<i>A. tataricum</i> 			OQ875005	—
	<i>A. tataricum</i> 			OQ866170	—
	<i>A. tataricum</i> 			—	—
	<i>A. tataricum</i> 				
	<i>A. saccharinum</i>				
	<i>A. argutum</i>				—
S. aceris-arguti sp. nov.	<i>A. argutum</i>			—	—
	<i>A. argutum</i>				
	<i>A. argutum</i>			—	—
	<i>A. argutum</i>			—	—
	<i>A. argutum</i>			—	—
	<i>Aesculus chinensis</i> 			—	—
	<i>Aesculus chinensis</i>				
	<i>Aesculus chinensis</i> 			—	—
	<i>Acer campestre</i>			OQ866160	OQ866160
	<i>A. campestre</i>			OQ866171	—
S. bicornis f. <i>polyphaga</i> f. nov.	<i>A. campestre</i>			OQ866172	—
	<i>A. heldreichii</i> 			OR166375	—
	<i>A. negundo</i>			OQ866159	OQ866159
	<i>A. pseudoplatanus</i>		—	—	—
	<i>A. ukunduense</i>				
	<i>A. velutina</i>			OR166383	OR166383
	<i>Acer</i> 			OR166382	OR166382
	<i>Acer</i> 			—	OQ866189
	<i>A. pictum</i> 			OR166385	OR166385
	<i>A. bifida</i>			—	—

Fungal species^a

Fungal species ^a	Host	Specimen No. ^b	Collection locality	GenBank accession number	
				ITS	28S rDNA
S. bomiensis	A. pictum mono	W	W	OR166373	—
	A. pictum mono	W	W	OR166374	—
	A. pictum mono	W	W	—	—
	A. pictum mono	W	W	—	—
	A. pictum mono	W	W	—	—
	A. pictum mono	W	W	—	—
S. koelreuteriae	A. caudatum	W	W	—	—
	A. caudatum	W	W	—	—
S. kovaliana	Koelreuteria paniculata	W	W	—	—
	K. paniculata	W	W	—	—
	K. paniculata	W	W	—	—
	K. paniculata	W	W	—	—
	K. paniculata	W	W	—	—
	K. paniculata	W	W	—	—
	K. paniculata	W	W	—	—
	K. paniculata	W	W	—	—
	K. paniculata	W	W	—	—
	K. paniculata	W	W	—	—
	K. paniculata	W	W	—	—
	K. paniculata	W	W	—	—
	K. paniculata	W	W	—	—
	K. paniculata	W	W	—	—
S. nankinensis	A. pseudosieboidianum	W	W	—	—
	A. truncatum	W	W	—	—
	A. buergerianum	W	W	—	—
	A. buergerianum	W	W	—	—

Table 1. 8

Fungal species ^a	Host	Specimen No. ^b	Collection locality	GenBank accession number	
				ITS	28S rDNA
<i>S. negundinis</i>	<i>A. buergerianum</i>	1	6	—	—
	<i>A. buergerianum</i>	2	6	—	—
	<i>A. buergerianum</i>	3	6	—	—
	<i>A. negundo</i>	4	6	21	21
	<i>A. negundo</i>	5	6	21	21
	<i>A. negundo</i>	6	6	21	21
	<i>A. negundo</i>	7	6	—	—
	<i>A. negundo</i>	8	6	—	8
	<i>A. negundo</i>	9	6	QQ866163	QQ866163
	<i>A. negundo</i>	10	6	QQ866174	—
	<i>A. negundo</i>	11	6	QQ866162	QQ866162
	<i>A. negundo</i>	12	6	QQ866175	—
	<i>A. negundo</i>	13	6	QQ866176	—
	<i>A. triflorum</i>	14	6	QQ866161	QQ866161
	<i>A. triflorum</i>	15	6	QQ866177	—
	<i>A. negundo</i>	16	6	21	21
	<i>A. negundo</i>	17	6	21	21
	<i>A. negundo</i>	18	6	OR711904	21
	<i>A. triflorum</i>	19	6	OR708548	21
	<i>A. pictum</i> mono	20	6	OR711903	21
<i>S. polyfida</i>	<i>A. negundo</i>	21	6	OR708549	21
	<i>A. negundo</i>	22	6	—	—
	<i>A. amplum</i> scatalpifolium	23	6	—	—
	<i>A. amplum</i> scatalpifolium	24	6	—	QQ874975
	<i>A. circinatum</i>	25	2	OR166376	—
	<i>A. palmatum</i>	26	2	OR166379	—
	<i>A. palmatum</i>	27	2	OR166384	OR166384
	<i>A. palmatum</i>	28	6	21	21

Table 1. Fungal species^a

Host





































































Specimen No.^b

Collection locality

GenBank accession number

ITS

28S rDNA

<i>A. palmatum</i>	<i>A. palmatum</i>				
<i>A. palmatum</i>	<i>A. palmatum</i>			—	—
<i>A. palmatum</i>	<i>A. palmatum</i>			—	—
<i>A. palmatum</i>	<i>A. palmatum</i>			—	—
<i>A. palmatum</i>	<i>A. palmatum</i>			—	—
<i>A. palmatum</i>	<i>A. palmatum</i>			OR863613	OR863613
<i>A. palmatum</i>	<i>A. palmatum</i>			—	—
<i>A. palmatum</i>	<i>A. palmatum</i>			—	—
<i>A. palmatum</i>	<i>A. palmatum</i>			—	—
<i>A. palmatum</i>	<i>A. palmatum</i>			OR701408	OR701408
<i>A. palmatum</i>	<i>A. palmatum</i>			OR701409	OR701409
<i>A. palmatum</i>	<i>A. palmatum</i>			OR701411	OR701411
<i>Acer</i> 	<i>Acer</i> 			—	—
<i>Acer truncatum</i>	<i>Acer truncatum</i>				
<i>A. truncatum</i>	<i>A. truncatum</i>				
<i>A. truncatum</i>	<i>A. truncatum</i>			—	—
<i>A. truncatum</i>	<i>A. truncatum</i>			—	—
<i>A. truncatum</i>	<i>A. truncatum</i>			QO866158	QO866158
<i>A. barbinerve</i>	<i>A. barbinerve</i>			QO866169	—
<i>Liquidambar formosana</i>	<i>Liquidambar formosana</i>				
<i>Acer</i> 	<i>Acer</i> 			OR841072	OR841072
<i>A. platanoides</i>	<i>A. platanoides</i>			OR166380	OR166380
<i>A. platanoides</i>	<i>A. platanoides</i>			OR166381	OR166381
<i>A. platanoides</i>	<i>A. platanoides</i>			QO866165	QO866165
<i>A. platanoides</i>	<i>A. platanoides</i>			—	—
<i>A. platanoides</i>	<i>A. platanoides</i>			—	QO866193
<i>A. platanoides</i>	<i>A. platanoides</i>			QO866178	—
<i>A. platanoides</i>	<i>A. platanoides</i>			—	—

S. taii sp. nov.*S. tulasnei* 

Fungal species ^a	Host	Specimen No. ^b	Collection locality	GenBank accession number	
				ITS	28S rDNA
<i>Takamatsuella circinata</i>	<i>A. platanoides</i>	■	■	OQ866166	OQ866166
	<i>A. platanoides</i>	■	■	—	—
	<i>A. pycnanthum</i>	■	■	OR166378	—
	<i>A. rubrum</i>	■	■	—	—
	<i>A. spicatum</i>	■	■	OR166377	—
	<i>A. wilsonii</i>	■	■	—	—
<i>Acer</i>	<i>Acer p</i>	■	■	—	—
	<i>Acer p</i>	■	■	—	—
	<i>A. platanoides</i>	■	■	—	—
	<i>A. tataricum</i>	■	■	—	—
<i>A. tataricum</i>	<i>A. tataricum</i>	■	■	—	—

Type species: *Sawadaea aceris* (DC.) Miyake (\equiv *S. bicornis*)^W

Sawadaea we

***Sawadaea acerina* sp. nov.**

Etymology: *acer*,
+ Latin adjectival suffix -ina (belonging to).

Typus: China, 1978

Acer tataricum, *pinnale*, X. Guan

& J. Feng, *holotype*, *holotype*

Diagnosis: *Sawadaea polyfida*, 1978

Mycelium *diff*

hyphae *diff*

thin-walled, 27.0–69.5(–83.0) \times 2.5–6.5 μ m; *hyphal appressoria*

micro-conidiophores

59.5 \times 4.0–6.5 μ m, arising from the mother cell centrally or towards one septum, foot-cells cylindrical, straight, 12.5–29.0 \times 4.0–6.0 μ m, basal septum sometimes elevated, 7.0–16.5 μ m distant from

micro-conidia *diff*

ovoid, 5.5–9.5(–13.5) \times 5.0–7.5(–11.0) μ m, length/width ratio 1.0–1.5(–2.5) (av. 1.3), with fibrosin bodies; *macro-conidiophores* 25.5–79.0 \times 5.5–10.5 μ m, arising from the upper surface of the mother cells, foot-cells cylindrical, straight, 13.5–36.0 \times 5.5–10.5 μ m, basal septum sometimes elevated, 3.5–18.5 μ m distant from

macro-conidia *diff*

doliiform, fresh conidia 20.5–38.5(–54.5) \times 12.5–23.0 μ m, length/width ratio 1.0–4.0 (av. 1.8), with fibrosin bodies; *germ tubes*

orthotubus

Fibroidium *Chasmothecia*

(126.5–)141.5–211.5 μ m diam.; *peridium cells*

4.0–19.5 μ m diam., arranged \pm radially; *appendages*

branched, (31.5–)63.0–195.5 \times 4.5–14.0 μ m, 0.2–1.3 (av. 0.8) times

56.0–98.0 \times 35.0–53.0 μ m, length/width ratio 1.2–2.5 (av. 1.8),

wall up to 4.7 μ m, short-stalked or almost sessile, (6–)8-spored;

ascospores ellipsoid-ovoid, 12.5–26.5 \times 11.0–15.0 μ m, length/width

Additional materials examined: **China**, **Yunnan**

Acer tataricum **ginnala**

S. Takamatsu, V.N. Nguyen, J. Feng & W.W. Yan
PM91886, 28S rDNA GenBank No.: OQ866185; HMJAU-PM92100);

Acer tataricum

ginnala,

G.X. Guan & J. Feng

GenBank No.: OQ875005; HMJAU-PM92099, ITS rDNA GenBank
No.: OQ866170); Jilin Province, Changchun City, on *Acer tataricum*

ginnala

S.R. Tang, G.X. Guan & J. Feng

Acer tataricum

ginnala

Oct. 2010, W.T. Jian

Acer saccharinum

24 Oct. 2018, S.R. Tang & L. Liu

Acer tataricum

ginnala, 24 Oct. 2018,

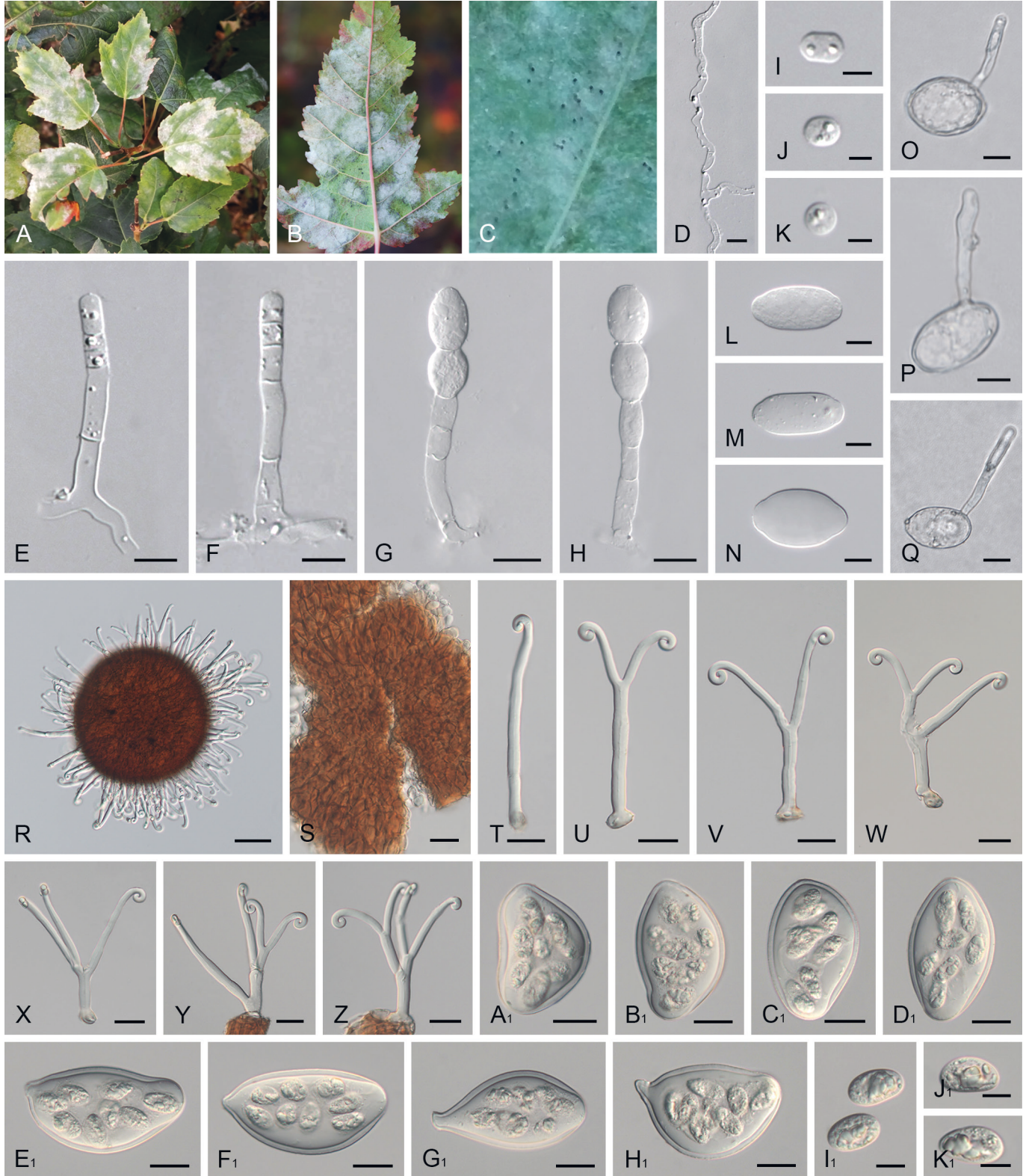


Fig. 4. *Sawadaea acerina* sp. nov. **Acer tataricum**

ginnala

L-N-Q-R-S-Z

R = 50 µm; G, H, S-H1 = 20 µm; D-F, L-Q, I1-K1 = 10 µm; I-K = 5 µm.

ginnala

E, F, H

G, H, I

I-K

A1-H1

I1-K1

S.R. Tang & L. Liu (HMJAU-PM91888, 28S rDNA GenBank No.: OQ866186).

Host range and distribution: On *Acer* (*saccharinum*, *tataricum* subsp. *ginnala*), *Sapindaceae*, Asia (China).

Notes: The new species can be morphologically readily distinguished from other species of *Sawadaea*: (1) from *S. koelreuteriae* and *S. nankinensis* by having branched appendages; (2) from *S. aesculi* and *S. bomiensis* by having a much larger number of chasmothecial appendages (58–142 vs < 50); (3) by having chasmothecial appendages branched in the middle or lower part (vs appendages branched in the middle or upper part of the appendages in *S. bicornis*); (4) by having larger chasmothecia (141–212 μm diam., vs 115–185 μm in *S. bifida*); (5) by having shorter macro-conidiophores (up to 79 μm vs 150 μm) and shorter foot-cells (21–31 μm vs 25–50 μm) compared with *S. negundinis*, but larger macro-conidia and micro-conidia compared with *S. negundinis* and *S. tulasnei* (macro-conidia 25–55 \times 13–21 μm vs 24–30 \times 15–19 μm and vs 16–28 \times 10–18 μm , micro-conidia 21–31 \times 13–23 μm vs 6–8 \times 5–7 μm and vs 2–11 \times 6–9 μm). The new species is morphologically similar to *S. polyfida*, but it differs in having mainly unbranched or 1–2 times branched chasmothecial appendages and it forms a separate

well-supported species clade in the molecular phylogenetic tree. In addition, we need a special explanation that there were only a few fully mature chasmothecia on the holotype specimen, due to the early collection season.

Sawadaea aceris-arguti S. Takam. & U. Braun, **sp. nov.** MycoBank MB 849117. Fig. 5.

Etymology: Epithet derived from the name of the host plant, *Acer argutum*.

Typus: Japan, Nagano Pref., Ueda-shi, Sugadaira, Tsukuba University, Sugadaira Research Station, on *Acer argutum*, 30 Sep. 2000, S. Takamatsu [**holotype** TNS-F-87577, ex-holotype sequence: GenBank No. AB193367 (ITS rDNA)].

Diagnosis: Morphologically close to *Sawadaea polyfida*, but chasmothecia only with 50–100 appendages, fewer asci, 7–13, and shorter and, above all, narrower ascospores, 17–22 \times 9–11 μm (vs 100–250 appendages, 8–38 asci, 12.5–33 \times 9.5–19 μm in *S. polyfida*). Genetically different from all other species within *Sawadaea* by forming a well-supported separate species clade.

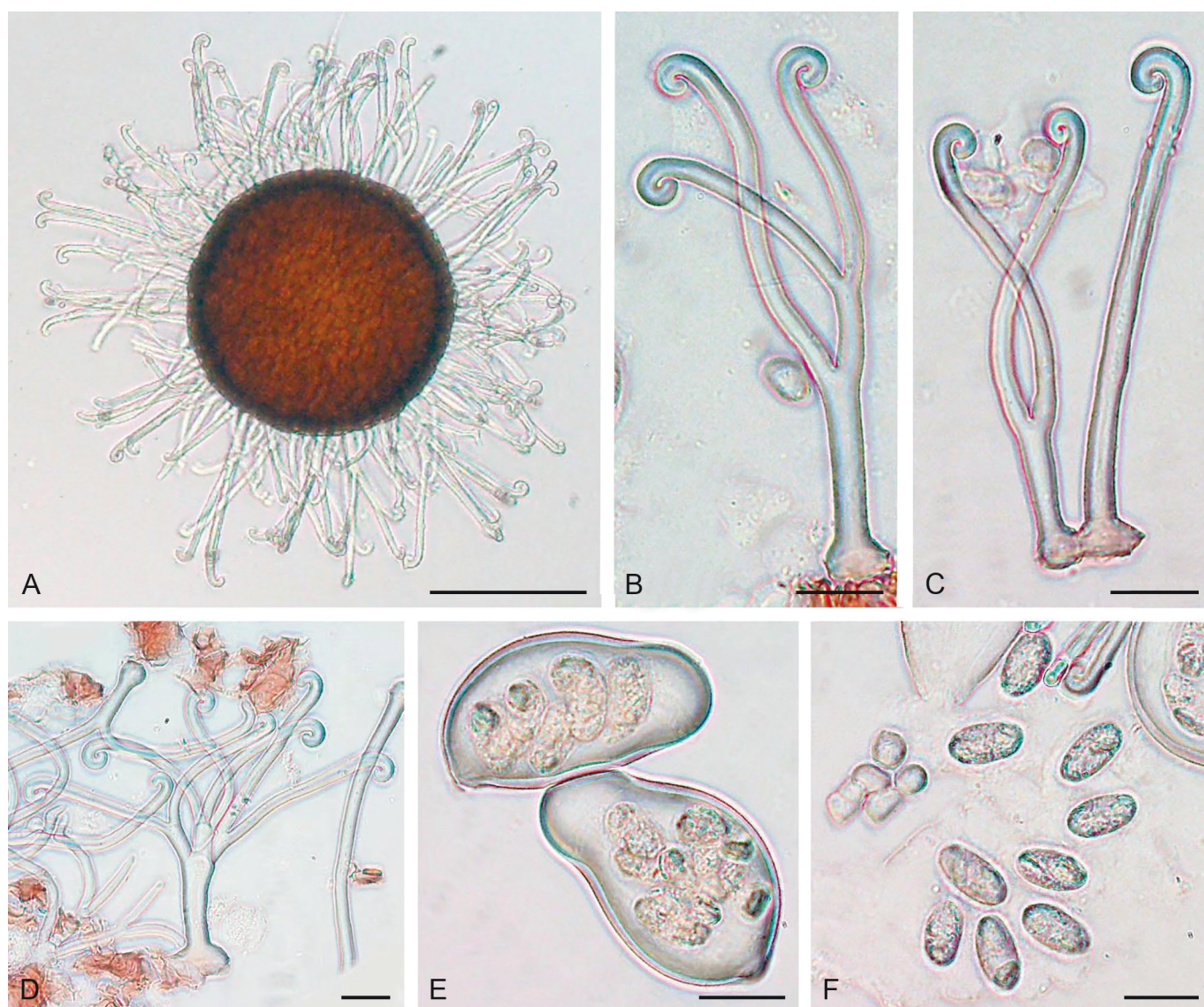


Fig. 5. *Sawadaea aceris-arguti* sp. nov. (TNS-F-87577, holotype). **A.** Chasmothecium. **B–D.** Appendages. **E.** Asci. **F.** Ascospores. Scale bars: A = 100 μm ; B–D = 50 μm ; E, F = 20 μm .

Mycelium hyphae $24.5\text{--}65.5 \times 3.0\text{--}8.0$ (-8.5) μm wide, hyaline, septate, thin-walled, smooth. *Chasmothecia* scattered, depressed globose, $(130.0\text{--})140.0\text{--}180.0 \mu\text{m}$ diam.; peridium cells appendages apices uncinately to circinate, $60.0\text{--}120.0 \mu\text{m}$ in length (0.3–0.7 times smooth to rough-walled, main axis $5.0\text{--}8.0 \mu\text{m}$ wide throughout; asci 7–13, saccate, $55.0\text{--}77.0 \times 32.0\text{--}47.0 \mu\text{m}$, short-stalked or almost ascospores ovoid, $17.0\text{--}22.0 \times 9.0\text{--}11.0 \mu\text{m}$, colourless.

Additional materials examined: Japan

o *Acer argutum*, 12 Oct. 1986, S. Tanda

Acer argutum, Takamatsu

o *Acer argutum*, C. Nakashima

Acer argutum, S. Tanda

Host range and distribution: On *Acer argutum*, Sapindaceae

Notes: *Sawadaea aceris-arguti*

S. polyfida

S. aceris-arguti

S. polyfida

Sawadaea

S. aceris-arguti

it was not possible to find any trace of asexual morphs, which

S. aceris-arguti

S. aceris-arguti

Sawadaea aesculi Acta Microbiol. Sin.

20

Typus: China, Beijing

Aesculus chinensis *wilsonii*,

N. Yu & Y.S. Xing holotype

Mycelium

hyphae sinuous, septate, branched, thin-walled, $24.5\text{--}65.5 \times 3.0\text{--}8.0$ (-8.5) μm ; hyphal appressoria micro-conidiophores $20.0\text{--}43.0 \times 5.0\text{--}7.0 \mu\text{m}$, arising from the cells cylindrical, straight, $7.0\text{--}21.5 \times 4.5\text{--}7.5 \mu\text{m}$, basal septum sometimes elevated, $2.5\text{--}11.5 \mu\text{m}$ from the junction with the mother micro-conidia variable in shape, ellipsoid-ovoid, $4.5\text{--}15.5 \times 4.5\text{--}11.0 \mu\text{m}$,

macro-conidiophores $85.5 \times 4.5\text{--}8.5 \mu\text{m}$, arising from mother cells, foot-cells cylindrical, straight, $(12.0\text{--})23.5\text{--}36.0$ (-56.0) $\times 4.5\text{--}9.0 \mu\text{m}$, basal septum sometimes elevated, $1.5\text{--}12.5 \mu\text{m}$ from the junction with the mother macro-conidia 32.0 (-48.5) $\times 10.0\text{--}18.0 \mu\text{m}$, length/width ratio 1.3–3.1 (-3.8), with fibrosin bodies; germ tubes

orthotubous fibroidium *Chasmothecia*

Additional materials examined: China

Aesculus chinensis

S.Q. Chen

Aesculus chinensis

S.R. Tang & J. Feng

Host range and distribution: On *Aesculus chinensis*, Sapindaceae,

Notes: *Sawadaea aesculi*

Aesculus chinensis

wilsonii

aesculi *Aesculus chinensis*

aesculi

aesculi

aesculi

aesculi

aesculi

Sawadaea bicornis Fac. Agric. Hokkaido Univ.

38

Basionym: *Alphitomorpha bicornis* Verh. Ges. Naturf.

Freunde Berlin

Synonyms: *Erysiphe bicornis* [Erysiphe]

6. nom. sanct.

Uncinula bicornis Ann. Sci. Nat., Bot., Sér. 3,

8

Erysiphe aceris Dyn. pl. Fl. gall.

Uncinula aceris Pyl. Fung.

Oidium aceris Flora

Oidium dodonaeae

Pal. J. Bot., Rehovot

Ser., 50

Typus: On *Acer campestre* *Erysiphe*

fl. Fr. 732" [lectotype]

epitype Austria,

Acer campestre,

U. Kruse,

Mycelium

hyphae sinuous, septate, branched, thin-walled, $26.5\text{--}55.0 \times 3.0\text{--}8.0 \mu\text{m}$; hyphal appressoria micro-conidiophores $(23.5\text{--})37.5\text{--}71.5$ (-123.5) $\times 5.0\text{--}9.0 \mu\text{m}$, arising from straight, $16.5\text{--}42.0$ (-61.0) $\times 5.0\text{--}8.5 \mu\text{m}$, basal septum sometimes elevated, up to $13.5 \mu\text{m}$ from the junction with the mother cell,

micro-conidia variable in shape, ellipsoid-ovoid, $18.0\text{--}28.0 \times 13.5\text{--}19.0 \mu\text{m}$, length/width ratio 1.0–1.7 (av. 1.5), with fibrosin bodies; *macro-conidiophores* $52.5\text{--}88.5\text{--}(112.5) \times 5.5\text{--}8.0 \mu\text{m}$, arising from mother cells, foot-cells cylindrical, straight, $18.5\text{--}54.5 \times 5.5\text{--}8.5 \mu\text{m}$, basal septum sometimes elevated, to $17.5 \mu\text{m}$ from the junction with the

macro-conidia

$4.0\text{--}(65.0) \times 10.5\text{--}20.0 \mu\text{m}$, length/width ratio 1.7–4.1 (–5.1) (av. 2.4), with fibrosin bodies; germ tubes terminal, short to moderately

orthotubus *Fibroidium*

scattered to \pm gregarious, $102.0\text{--}170.0 \mu\text{m}$ diam.; *peridium* cells irregularly polygonal, $3.0\text{--}14.0 \mu\text{m}$ diam., arranged \pm radially;

appendages

11.0–12.0

11.0–12.0

11.0–12.0

$31.5\text{--}137.5 \times 4.5\text{--}8.5 \mu\text{m}$, 0.2–1.2 times (av. 0.8) as long as the

asci

8–20, ellipsoid-obovoid, saccate-clavate, $57.0\text{--}98.0\text{--}(108.0) \times 40.0\text{--}65.0 \mu\text{m}$, length/width ratio 1.0–2.0 (av. 1.4), wall up to $4.6 \mu\text{m}$ thick, short-stalked or almost sessile, (6–)8-spored; *ascospores* ellipsoid-ovoid, $19.5\text{--}28.0 \times 11.0\text{--}21.0 \mu\text{m}$, length/width ratio 1.1–

Subspecific classification: *hobbs*

Sawadaea bicornis

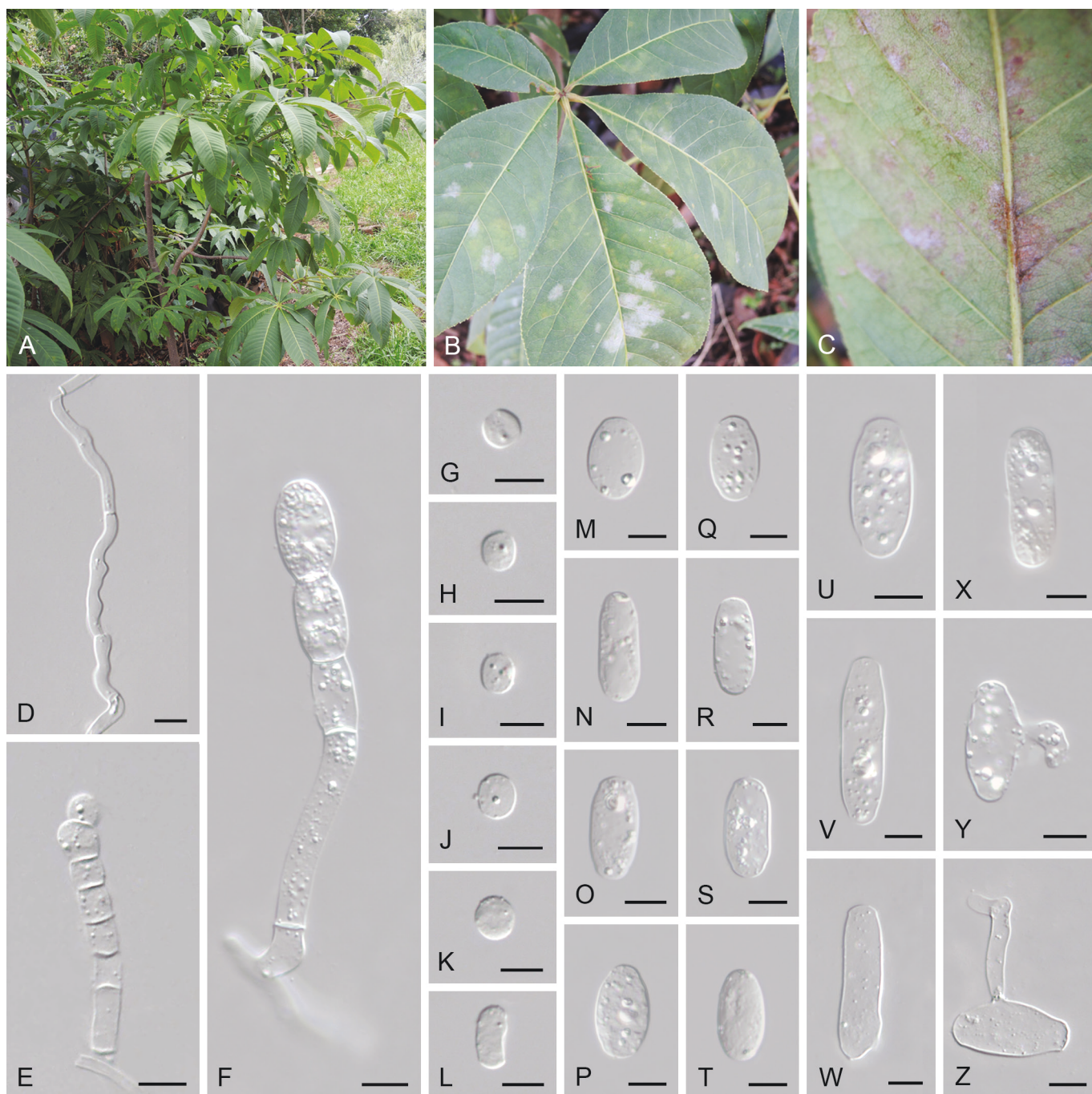


Fig. 6. *Sawadaea aesculi*.
D. germ tubes. Scale bars = $10 \mu\text{m}$.

A. B. C.

F. G. H. I.

M. N. O. P.

subclades that reflect two biological races with different host preferences genetically established and verifiable, they deserve a formal naming *formae speciales*

formae

formae

formae is that it is an official taxonomic

formae

S. bicornis

S. bicornis *bicornis* *polyphaga*

S. bicornis

subclades with specific host preferences (Fig. 1).

Sawadaea bicornis* f. *bicornis, *Acer campestre* *Acer platanoides*

Acer platanoides

Additional materials examined: **Russia**

Acer campestre

T.S. Bulgakov

OO866171); Rostov region, Shakhty, Alexandrovsky Park, on *Acer*

campestre, T.S. Bulgakov

rDNA GenBank No.: OQ866172); Rostov region, Shakhty, Hospital

Acer campestre, 24 Oct. 2018, T.S. Bulgakov

PM92107, ITS+28S rDNA GenBank No.: OQ866160).

Sawadaea bicornis* f. *polyphaga *nov.*

nov.

Etymology

Typus: Germany

Acer pseudoplatanus, 200

holotype

nov.

Diagnosis *bicornis*

Acer

Acer *Sinallaba* *Negundo* *Platanus* *Platanoides*,

subsp.

nov.

J. Kruse

polyphaga

Additional materials examined: **Canada**, Ontario, Niagara Region,

Acer negundo, V. Ilyukhin



Fig. 7. *Sawadaea bicornis*

formae

A, B.

J-L.

M-O.

C.

D.

E, F.

P, Q. Germ tubes. Scale bars = 10 µm.

(HMAU-PM92102, ITS+28S rDNA GenBank No.: OQ866159).

Russia

Acer pseudoplatanus, 492 T.S. Bulgakov

Acer ukurunduense S.Y.

Liu

Acer S.Y. Liu

GenBank No.: OQ866189). USA, *Acer heldreichii*

trautvetteri, 29 M. Bradshaw

ITS rDNA GenBank No.: OR166375); Boston, on *Acer velutina*, 3

M. Bradshaw

No.: OR166383); Boston, on *Acer* M. Bradshaw

(FH00941215, ITS+28S rDNA GenBank No.: OR166382).

Host range and distribution (based on collections confirmed by *bicornis* *Acer* (*campestre*, *platanoides*))

polyphaga *Acer* (*circinatum*, *grandidentatum*, *heldreichii*, *trautvetteri*, *macrophyllum*, *negundo*, *pseudoplatanus*, *saccharinum*, *tsinglingense*, *tataricum*, *tataricum*, *ukurunduense*)

Host range and distribution of *Sawadaea bicornis*

to Braun & Cook 2012): On *Acer amplum*, *catalpifolium* [= *catalpifolium*] *barbinerve*, *campestre*, *cappadocicum*, *cappadocicum* [= *laetum*] *cappadocicum*, *lobelii* [= *lobelii*] *caudatum*, *caudatifolium* [= *kawakamii*] *cissifolium*, *heldreichii*, *trautvetteri*, *hyrcanum*, *ibericum*, *macrophyllum*, *mandshuricum*, *miyabei*, *mono*, *monspessulanum*, *monspessulanum*,

monspessulanum, *turcomanicum* [*turcomanicum*] *negundo*, *obtusatum*, *oliverianum*, *opalus*, *opulifolium* [= *hispanicum*] *pennsylvanicum*, *platanoides*, *pseudoplatanus*, *pseudosieboldianum*, *rubrum*, *rufinerve*, *saccharinum*, *saccharum*, *stevenii*, *tataricum*, *tataricum*, *tataricum*, *ginnala* [= *ginnala*] *tataricum*, *semenovii* [= *semenovii*] *tegmentosum*, *trautvetteri*, *triflorum*, *trilobum*, *truncatum*, *turkestanicum* [= *platanoides*, *turkestanicum*] *ukurunduense*, *velutinum*) Sapindaceae; A

Aesculus hippocastanum, *Alectryon excelsus*, *Dodonaea viscosa*

Acer monspessulanum

et al.

Notes: *Sawadaea bicornis*

et al.

S. bicornis

S. bicornis

S. bicornis

Acer circinatum, *A. macrophyllum*

A. grandidentatum, *A. macrophyllum*

S. bicornis

S. bicornis

subclades. One of them is comprised of sequences obtained from

S. bicornis, *Acer campestre*, *A. platanoides*;

Acer, *Platanoides*

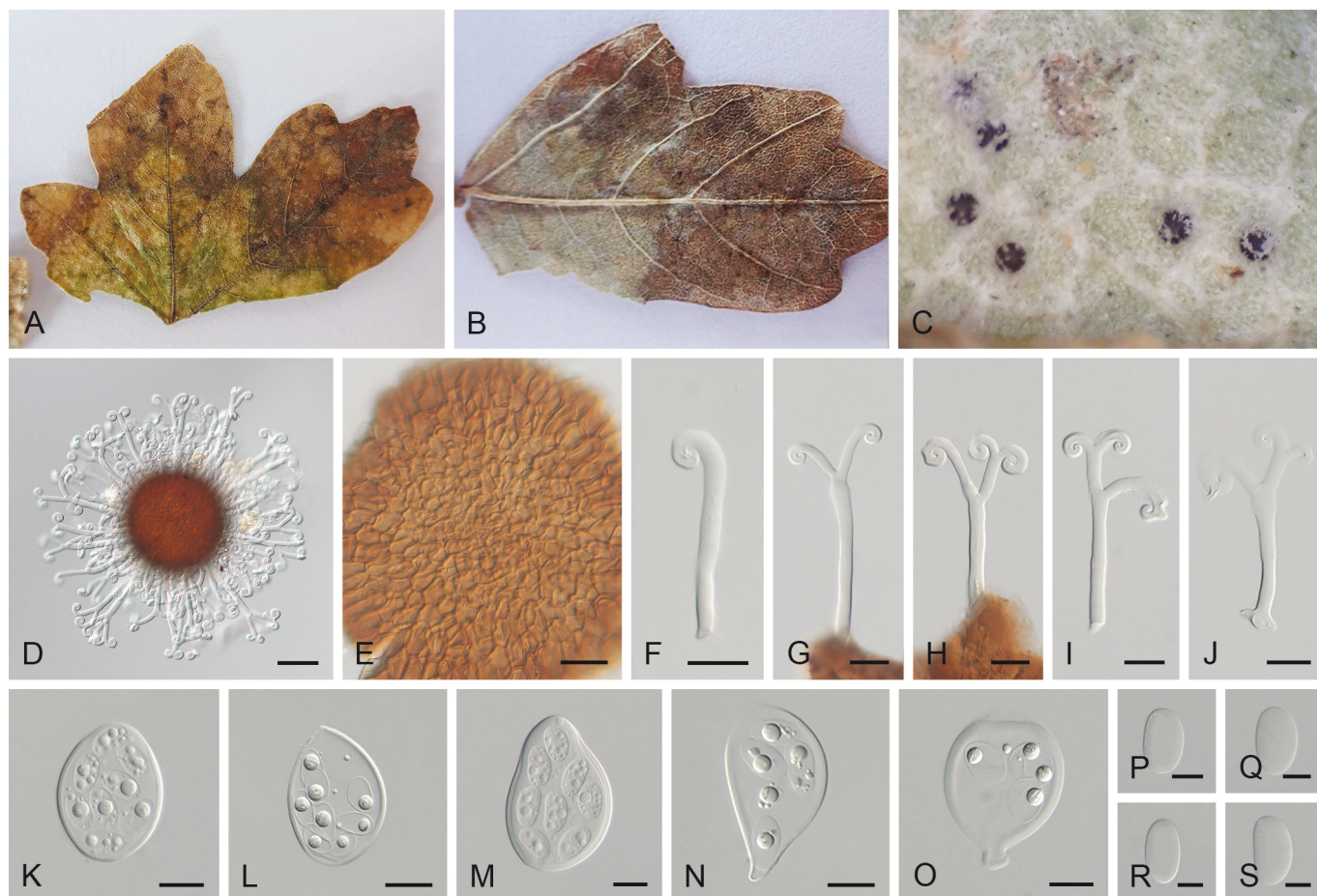


Fig. 8. *Sawadaea bicornis*

Scale bars: D = 50 µm; E–O = 20 µm; P–S = 10 µm.

Acer pseudoplatanus ;
 which might be a misidentification that requires reverification. The
A. pseudoplatanus d A.
A. negundo
A. acer b *Acer* (*A. grandidentata* ,
A. heldreichii b *trautvetteri*, *A. monspessulanum*, *A.*
pseudoplatanus, *A. velutinum*) b *Ginnala* (*A. tataricum*) b
Lithocarpa (*A. tsinglingense*) *Macrophylla* (*A. macrophylla*)
Negundo (*A. negundo*) *Palmata* (*A. circinatum*) b
Platanioidea (*A. cappadocicum*) *Rubra* (*A. saccharinum*)
 These results suggest that the first subclade is confined (or largely
A. Platanioidea
 reflects a wide host range within *Acer*.
 subclades reflect genetically fixed virulence and host preference
formae
Acer
campestre,
Sawadaea *A. pseudoplatanus* *A. negundo*

Sawadaea
 contamination or host misidentifications. Future research should
Sawadaea *Acer*
A. campestre *bicornis*
A. pseudoplatanus *f. polyphaga*
Acer campestre
pseudoplatanus

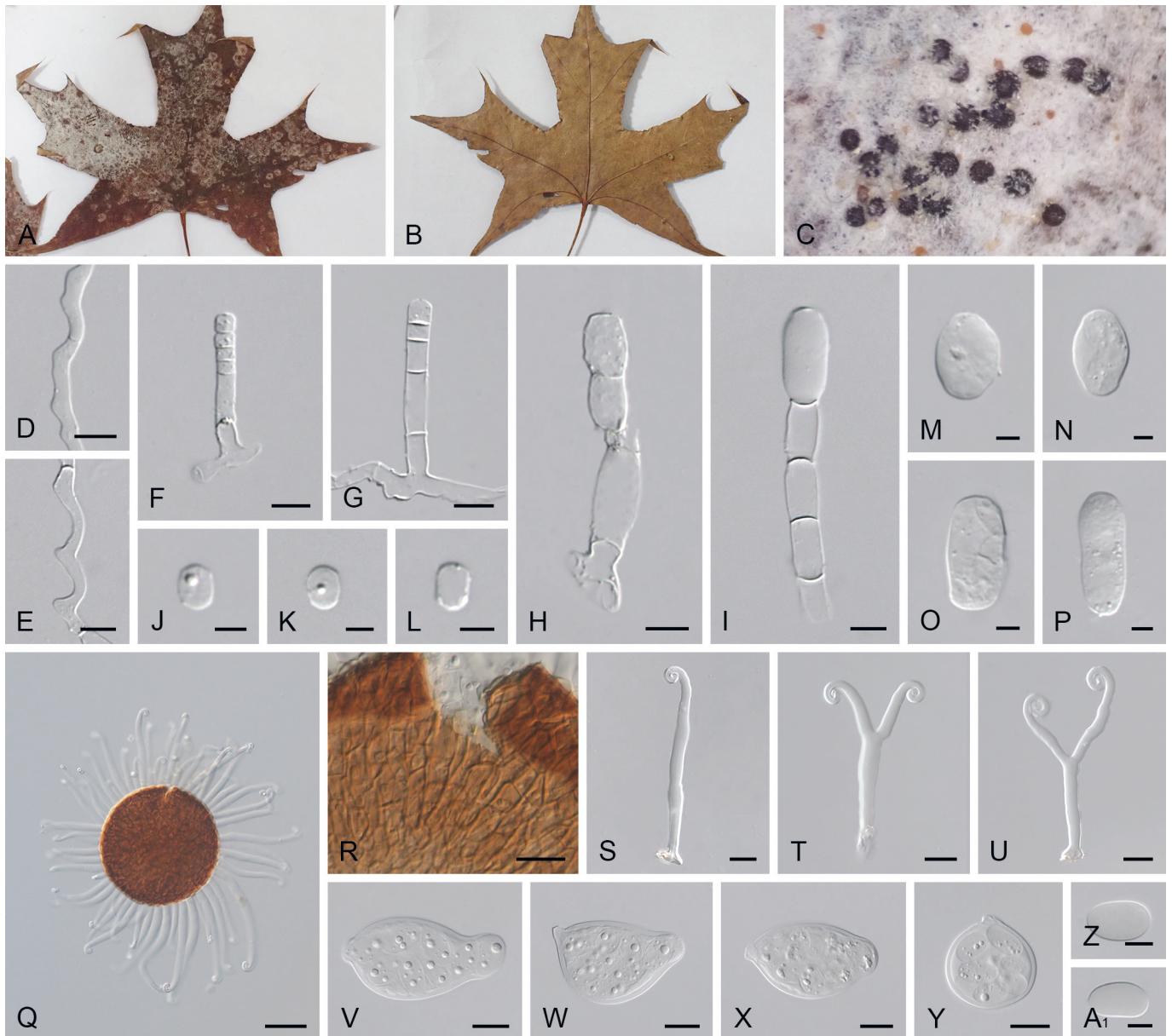


Fig. 9. *Sawadaea bifida*.
 E. G. H. I. L. M. N. O. P. Q. R. S. T. U. V. W. X. Y. Z. A1. Ascospores. Scale bars: Q = 50 µm; H, I, R–Y = 20 µm; D–G, Z, A1 = 10 µm; J–P = 5 µm.

S. bicornis, *Acer campestre*, *d A. platanoides*,
S. bicornis, *d S. negundinis*, *d S. tulasnei*,
f S. bicornis, *d Acer platanoides*, *d A.*
negundo, *d A. platanoideae*, *d A. negundo*

Sawadaea bifida Wkrajins'k. Bot. Zhurn. 47(9)
 9
 Synonyms: *Sawadaea zhengii* 9 Taxonomical study of

Erysiphaceae of Japan: *Acer mono*
holotype
Sawadae Trans. Mycol. Soc. Japan

Typus: Russia
Acer pictum
 mono, 1 Oct. 1989, V.P. Heluta **holotype**
 sequence: OR166385 (ITS+28S rDNA).

Mycelium branched, thin-walled, 12.0–46.5 × 2.5–6.5 µm; *hyphae* branched, thin-walled, 12.0–46.5 × 2.5–6.5 µm; *hyphal appressoria* branched, thin-walled, 12.0–46.5 × 2.5–6.5 µm; *micro-conidiophores* branched, thin-walled, 12.0–46.5 × 2.5–6.5 µm; *micro-conidia* branched, thin-walled, 12.0–46.5 × 2.5–6.5 µm.

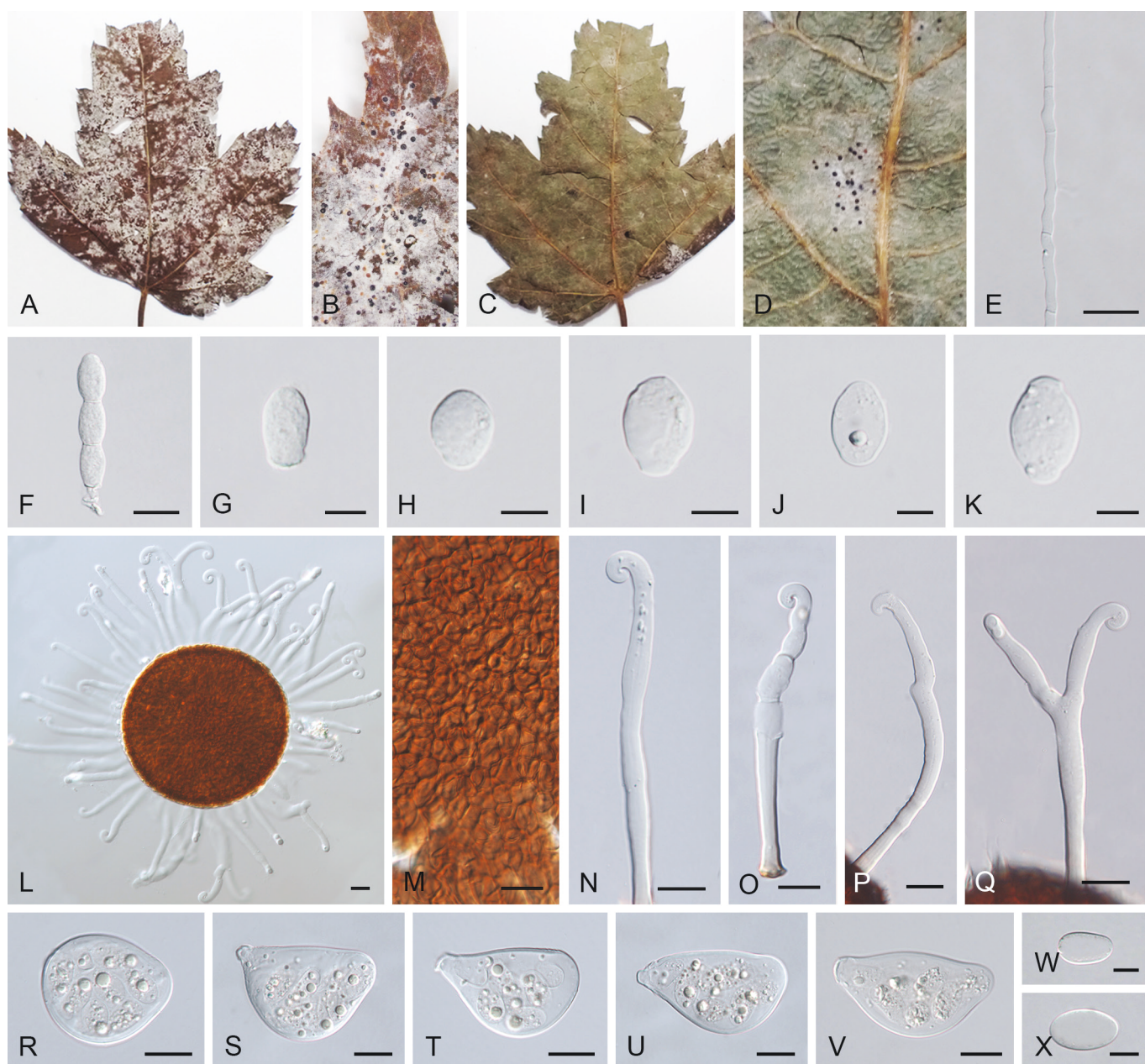


Fig. 10. *Sawadaea bomiensis*. A–C, Asci; D–K, Ascospores; L–V, Asci; W, X, Ascospores. Scale bars: E, F, L–V = 20 µm; G–K, W, X = 10 µm.

ovoid, $5.0\text{--}14.0 \times 4.5\text{--}12.5\ \mu\text{m}$, length/width ratio 1.0–1.6; macroconidiophores $27.0\text{--}52.5 \times 7.5\text{--}10.5\ \mu\text{m}$, arising from the upper surface of mother cells, foot-cells cylindrical, straight, $18.5\text{--}34.5 \times 4.0\text{--}6.5\ \mu\text{m}$, followed by 1–3 shorter cells and catenescant conidia; macroconidia

$17.0\text{--}29.0 \times 9.5\text{--}18.5\ \mu\text{m}$, length/width ratio 1.2–1.9. Chasmothecia

μm diam.; peridium cells

oblong, $4.5\text{--}23.0\ \mu\text{m}$ diam.; appendages

nodulose, deeply cleft, first branching point in the lower half, often

chasmothecial diam., $34.0\text{--}187.5 \times 7.0\text{--}11.5\ \mu\text{m}$, about 0.2–1.2

asci $47.0\text{--}96.5 \times 33.5\text{--}57.5\ \mu\text{m}$, length/width ratio 1.0–2.4, wall relatively thick, up to $4.5\ \mu\text{m}$, almost sessile to short-stalked, (6–)8-spored; ascospores ellipsoid, $18.5\text{--}30.0 \times 10.5\text{--}15.0\ \mu\text{m}$, length/width ratio

Additional materials examined: China,

Acer pictum mono, 5 Oct. 1975,

Y.N. Yu & S.J. Han

Acer pictum

mono, F.Z. Shi

Acer pictum

Y.J. Lu

Acer pictum

H. Huang, Russian

Acer

pictum mono, V.P. Heluta

ITS rDNA GenBank No.: OR166374); Far East, Primorsky Kray,

Acer pictum mono, P. Heluta

(KW-M-71485, ITS rDNA GenBank No.: OR166373).

Host range and distribution: On *Acer pictum* mono, *pictum*

dissectum, *pictum mayrii*) Sapindaceae

Notes: *Sawadaea bifida*

Russian Far East, and its host range is confined to *Acer pictum*

mono (= *A. mono*)

S. negundinis

S. bifida.

S. bifida.

S. bifida.

multilocus approach is needed. The micro-conidiophores ($30\text{--}82 \times$

$6.5\text{--}10\ \mu\text{m}$, vs $21.5\text{--}43 \times 3\text{--}7\ \mu\text{m}$) and micro-conidia ($16\text{--}29 \times 13\text{--}$

$23.5\ \mu\text{m}$, vs $5\text{--}14 \times 4.5\text{--}12.5\ \mu\text{m}$) of *S. negundinis* are significantly

S. bifida.

Acer pictum, *Acer*

Acer

Acer

Sawadaea

on *Acer pictum* mono, *Acer pictum*

Acer pictum mayrii, *nipponicum*

parviflora *Acer* (et al. 2019). The host identification

misidentification with a species of sect. *Platanioidea*

correct identification, to confirm infections of this species on hosts of

Acer

Platanioidea

nipponicum

nipponicum

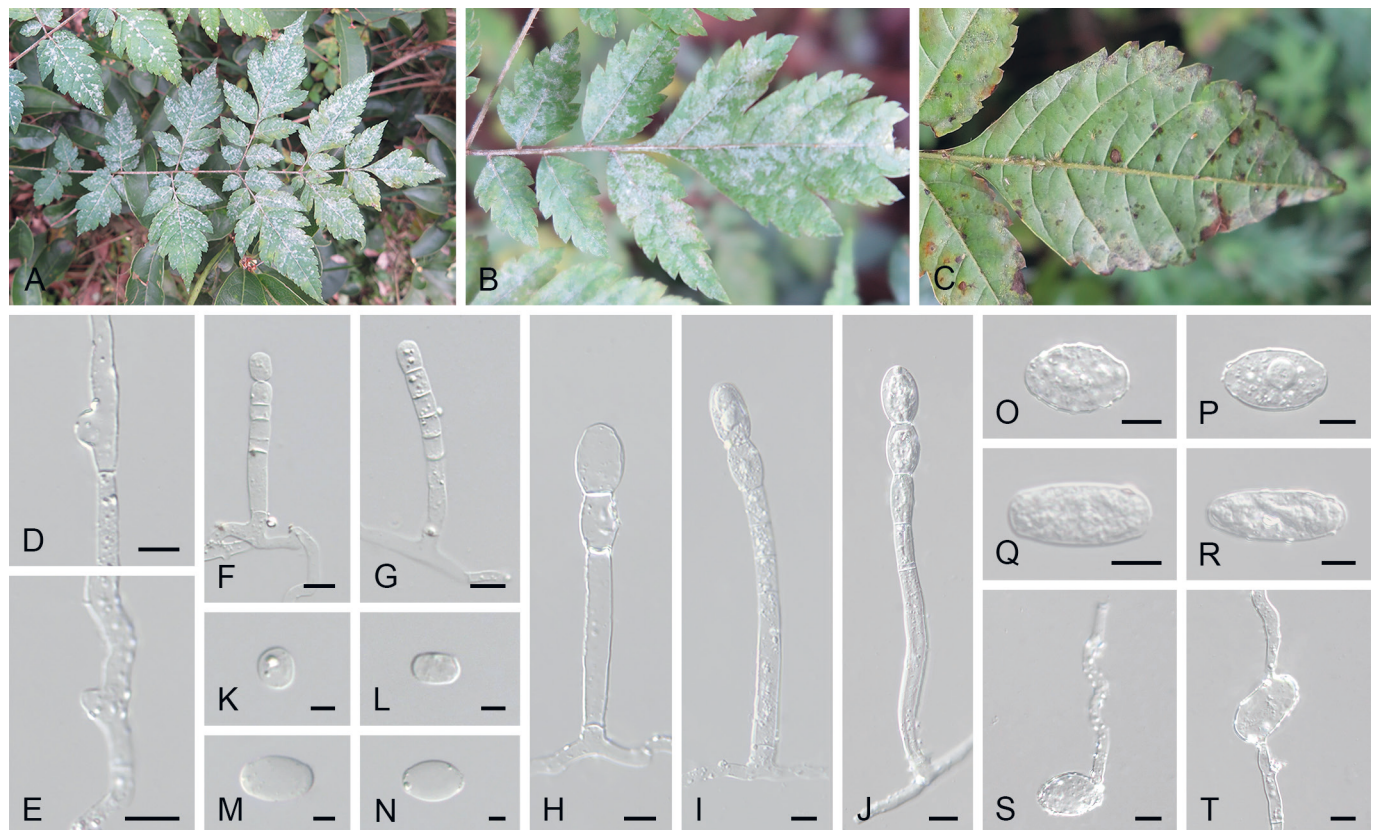


Fig. 11. *Sawadaea koelreuteriae*, B.

F, G, H–J.

K–N.

C.

O–R.

D, E.

D–G, O–T = 10 μm ; K–N = 5 μm .

S, T. Germ tubes. Scale bars: H–J = 20 μm ;

Sawadaea bomiensis *Acta Microbiol.*

Sin. 2011

Typus: China, 66276

Acer caudatum, M.M. Shen (holotype)

Mycelium

hyphae

thin-walled, 2.5–5.5 µm wide; hyphal appressoria

conidia

(18.5–27.0(–33.5) × 11.5–17.5 µm, length/width ratio 1.1–1.7(–2.2)

Chasmothecia

µm diam.; peridium cells irregularly polygonal, 4.5–24.5 µm diam.,

appendages

on

hyphae

0.4–0.8

times as long as the chasmothecial diam., 73.5–174.0 ×

7.5–14.5 µm, mostly somewhat undulate or almost nodulose, thick-

walled

asci

saccate-clavate, 57.0–95.0 × 48.5–60.0 µm, length/width ratio 1.1–

1.7

ascospores ellipsoid-ovoid, (13.0–)17.0–27.0 × 10.5–16.5 µm,

1–1.5

Additional material examined: China, 66276

Acer 66276

on

M. Zang

Host range and distribution: On *Acer caudatum*, Sapindaceae

in

Notes: 66276

*Sawadaea bomiensis*on *Acer caudatum*, 66276

on

on

on

on

on

on

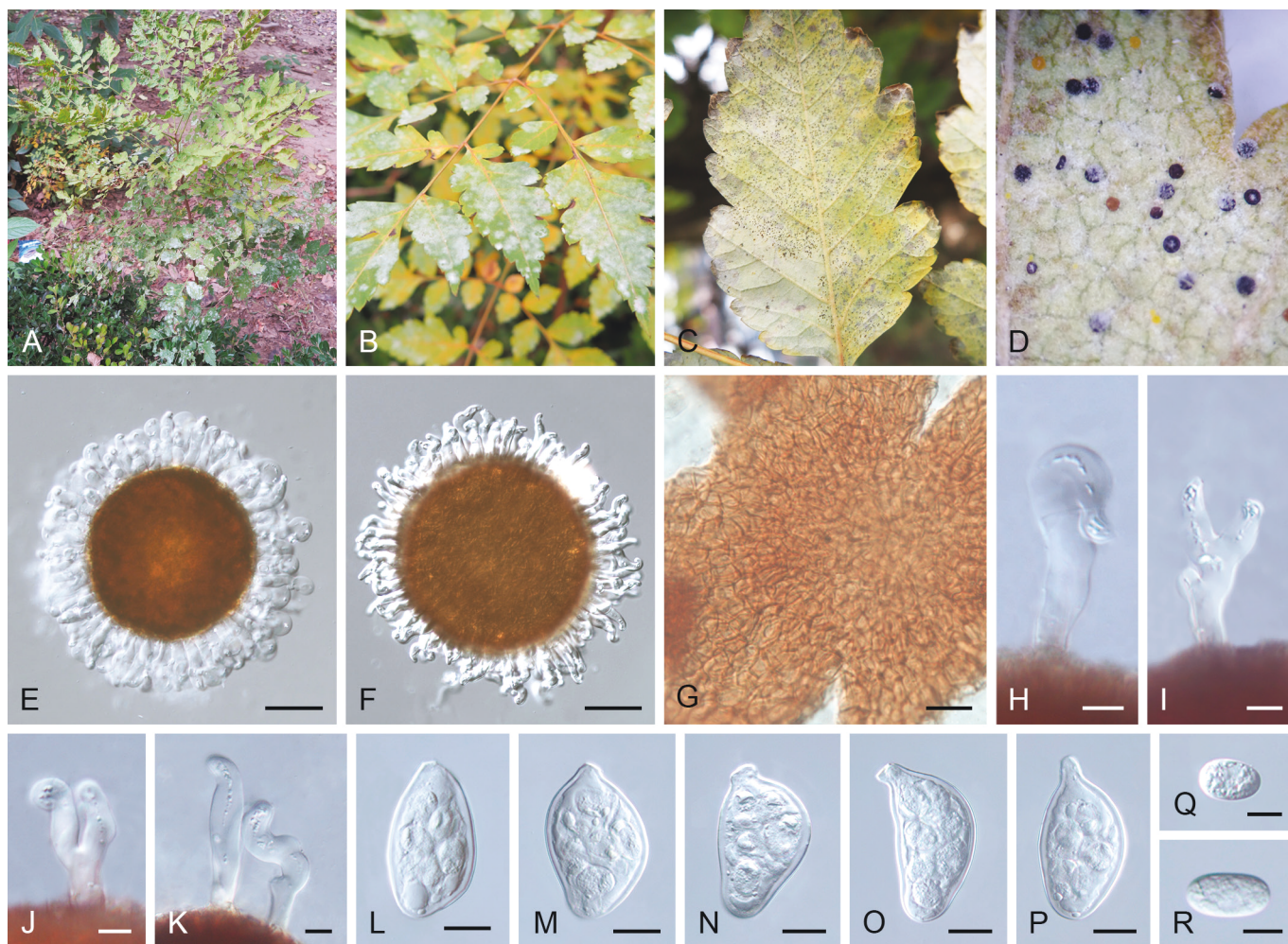
on

on

on

Sawadaea koelreuteriae (M.M. Shen)*Microbiol.* 49Basionym: *Uncinula koelreuteriae* M*Bot. Mag. Tokyo* 27:

9

Synonyms: *Erysiphe koelreuteriae* Bull. Chin.*Bot. Soc.* 29*Typhulochaeta koelreuteriae* Bull. Torrey Bot.*Club.* 73Fig. 12. *Sawadaea koelreuteriae* M.M. Shen

in Figs. 12–K

Scale bars: E, F = 50 µm; G–P = 20 µm; Q, R = 10 µm.

C. 12

L–P. 12

Q, R. 12

Typus: China

S.X. Wei & L. Guo

neotype

Mycelium

sinuous, 5.0–6.0 µm wide; hyphal appressoria

micro-conidiophores

44.0–76.0 × 7.5–9.0 µm, foot-cells 19.5–49.5 × 6.0–8.5 µm; micro-conidia broad ellipsoid-ovoid, 18.5–28.5 × 13.0–18.0 µm, length/

macro-conidiophores 83.0–139.5 × 7.5–11.0 µm, arising from the mother cell, centrally or towards one septum, foot-cells cylindrical, straight, 43.5–97.5 × 7.0–9.0 µm, followed

limoniform, fresh conidia

34.5–49.5 × 10.0–16.0 µm, length/width ratio 2.5–4.1, with fibrosin bodies; germ tubes

orthotubous

Chasmothecia scattered, 135.5–209.0 µm diam.; peridium cells irregularly polygonal, 8.0–18.5 µm diam.; appendages

to seldom forked, often curved, 26.0–54.5 µm, 0.1–0.3 times as

clavate-saccate, 56.0–89.5 × 31.0–53.0 µm, sessile or short-

ascospores 16.5–23.0 × 13.0–16.5 µm, length/width ratio 1.2–1.4, colourless.

Additional materials examined: China

Koelreuteria paniculata, Feng, L. Liu & R.J. Jiang

Koelreuteria paniculata, 21 Oct. 2018, S.R. Tang & L. Liu (HMJAU-PM91891, ITS rDNA GenBank No.: OQ866179); Hubei

Koelreuteria paniculata

S.R. Tang & L. Liu

Koelreuteria

paniculata, S.R. Tang & L. Liu

ITS rDNA GenBank No.: OQ866180; HMJAU-PM91897, ITS rDNA GenBank No.: OQ866182, 28S rDNA GenBank No.: OQ866187;

Koelreuteria paniculata

Oct. 2018, S.R. Tang & L. Liu

Koelreuteria

paniculata, 27 Oct. 2018, S.R. Tang & L. Liu

Koelreuteria

paniculata

J. Feng

Koelreuteria paniculata, S.R. Tang & L. Liu

Koelreuteria

paniculata, S.R. Tang & L. Liu

S.R. Tang & L. Liu

Koelreuteria

paniculata, S.R. Tang & L. Liu

28S rDNA GenBank No.: OQ866184; HMJAU-PM91898, ITS rDNA GenBank No.: OQ866183, 28S rDNA GenBank No.: OQ866188);

Koelreuteria paniculata

S.R. Tang & L. Liu

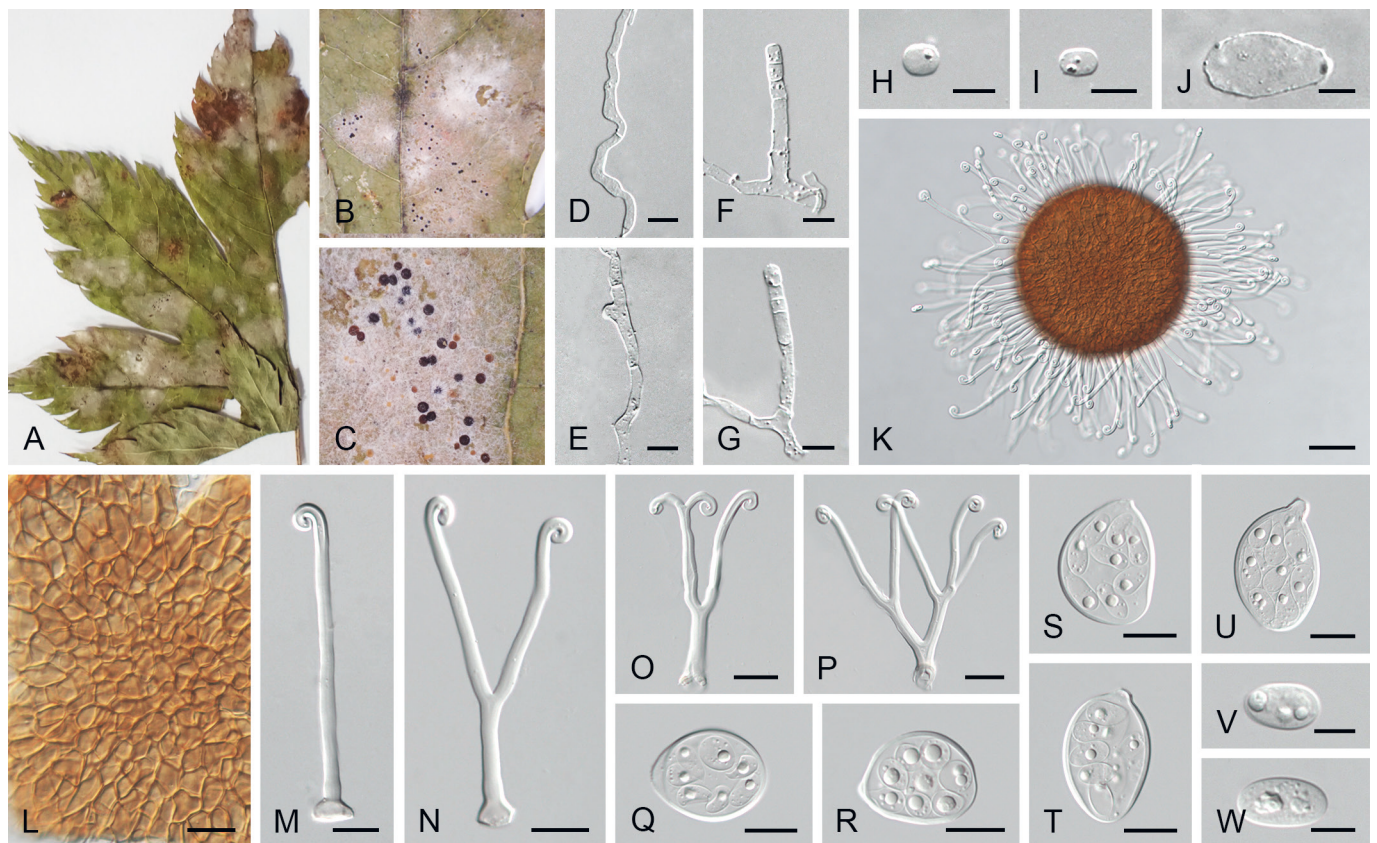


Fig. 13. Sawadaea kovaliana

A, B, C

F, G, H

I, J, K

L, M

N, O, P

Q, R

S, T, U

V, W

X, Y

Z, AA

AB, AC

AD, AE

AF, AG

AH

Scale bars: K = 50 µm; L–U = 20 µm; D–J, V, W = 10 µm.

Koelreuteria paniculata L.R. Tang & L. Liu
PM91894, ITS rDNA GenBank No.: OQ866181); Sichuan Province,
Koelreuteria paniculata L.R. Tang
& L. Liu

Host range and distribution: On *Koelreuteria bipinnata*, *paniculata*,
Koelreuteria Sapindaceae

Notes: This species was first reported by Miyake (1913) and described
as *Uncinula koelreuteriae*

6 *Phyllactinia*

Erysiphe *koelreuteriae*

Phyllochaeta

Phyllochaeta

T. koelreuteriae

Phyllochaeta

Erysiphe

6 *Sawadaea*

Sawadaea which was confirmed by molecular analyses. Liu *et al.*

S. koelreuteriae

sequences were obtained to confirm that this powdery mildew in
Koelreuteria *Sawadaea*.

Sawadaea kovaliana Ukrayins'k. Bot. Zhurn. 47

Typus: Russia

Pseudosieboldianum

2 Oct. 1989, V.P. Heluta **holotype**

sequence: GenBank No. OQ866167 (ITS+28S rDNA)].

Mycelium epiphyllous, forming distinct patches, confluent; **hyphae**
hyphal cell (16.5–)25.5–44.5 ×
3.0–7.5(–9.5) µm; **hyphal appressoria**
micro-conidiophores 36.0–54.5 × 4.5–7.0 µm,
foot-cells straight, cylindrical, 14.0–27.5 × 4.0–6.0 µm, basal
septum sometimes elevated, up to 10.5 µm from the junction
micro-conidia
ellipsoid-ovoid, 7.5–10.5 × 5.0–8.5 µm, length/width ratio 1.1–
macro-conidiophores **macro-conidia**
ellipsoid, 26.0–36.0 × 15.0–18.5 µm, length/width ratio 1.5–2.0.
Chasmothecia gregarious, semiglobose, 146.0–218.0 µm diam.;
peridium cells small, irregularly polygonal, about 4.0–27.5 µm diam.;
appendages

(45.0–)98.5–135.5(–171.5) × 3.5–10.5 µm, 0.2–1.0 times as long

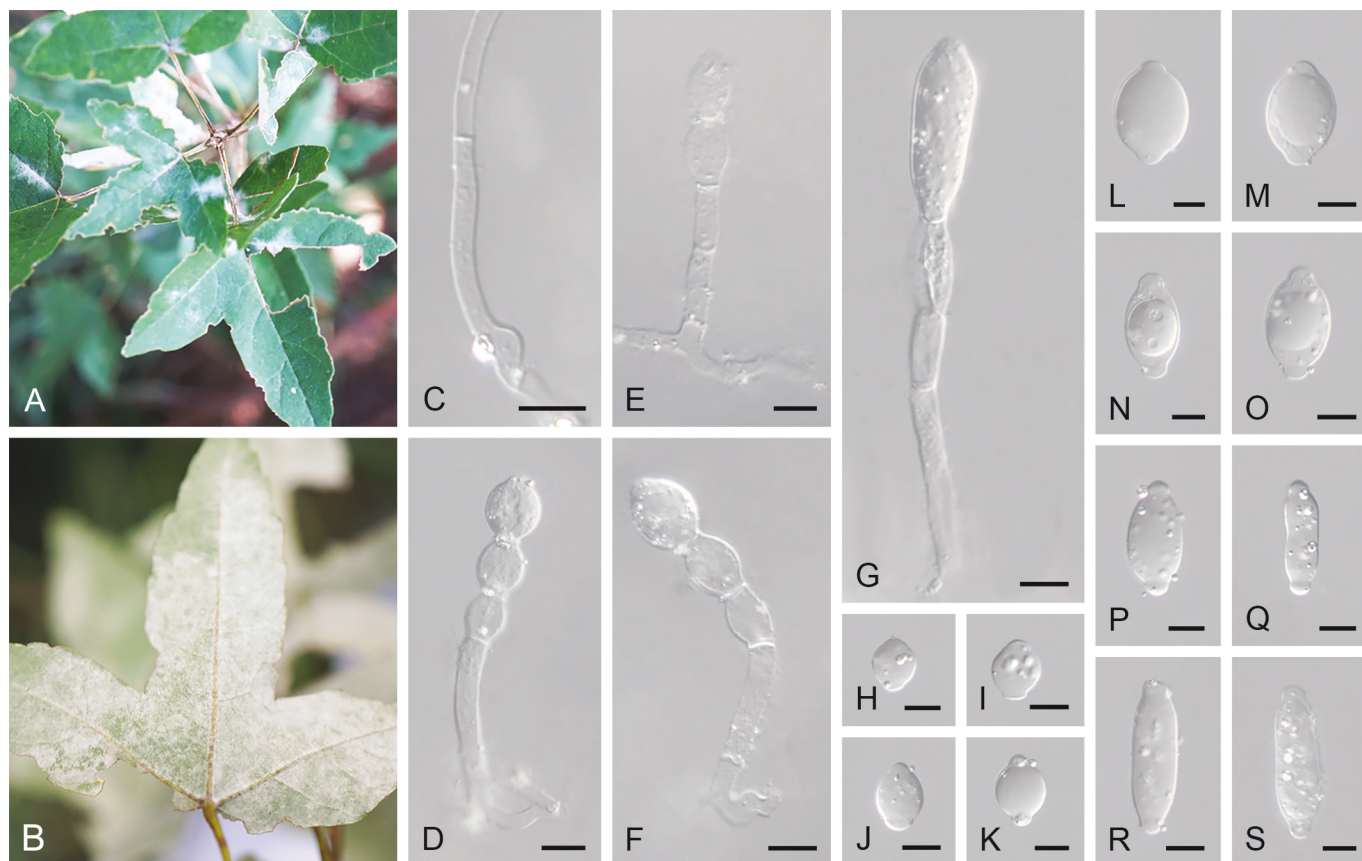


Fig. 14. *Sawadaea nankinensis*

A. B. C. D–F. G. H–K. L–S. Macro-conidia. Scale bars = 10 µm.

asci 24b

about 42.0–83 × 30.0–48.5 µm, length/width ratio 1.3–2.2, short-stalked, 5.0–13.5 × 6.5–11.5 µm, wall up to 3.5 µm thick, 8-spored;

ascospores 16b6b

11.5–24.0 × 8.0–14.0 µm, length/width ratio (1.0–)1.7–2.5(–2.9),

b

Additional material examined: **China**

o *Acer truncatum* Y. Zhao

GenBank No.: OQ866168).

Host range and distribution: On *Acer pseudosieboldianum*, *truncatum*) Sapindaceae

f

Notes: 16b6b

6 *Sawadaea kovaliana* 16b6b

6 *Sawadaea kovaliana* 16b6b

16b6b *Sawadaea*

S. taii *S. kovaliana*

S. taii

S. kovaliana, *Acer palmata*

S. taii *Platanioidea*

S. taii *S. kovaliana*

ascospores

16b6b

16b6b

kovaliana 16b6b

16b6b *A. truncatum* *Acer*

Platanioidea 16b6b

kovaliana 16b6b

be re-amplified to determine its accuracy). Braun & Cook (2012)

16b6b *S. bicornis*

16b6b *Acer pseudosieboldianum*

16b6b *Sawadaea bicornis*

16b6b *S. kovaliana*

16b6b *S. kovaliana*

16b6b *S. kovaliana*

Sawadaea nankinensis 16b6b

Mycoscience 49b

Basionym: *Uncinula nankinensis* 16b6b

China, Bot. Ser., 6b

Synonyms: *Erysiphe nankinensis* 16b6b

Schlechtendalia 40

Typus: **China** *Acer buergerianum*

16b6b F.L. Tai *lectotype* 16b6b

16b6b *lectotype*

Mycelium 16b6b

16b6b *hyphae*

16b6b *hyphae*

16b6b *hyphae* straight or somewhat sinuous, 3.0–7.0 µm;

16b6b *hyphal appressoria* 16b6b

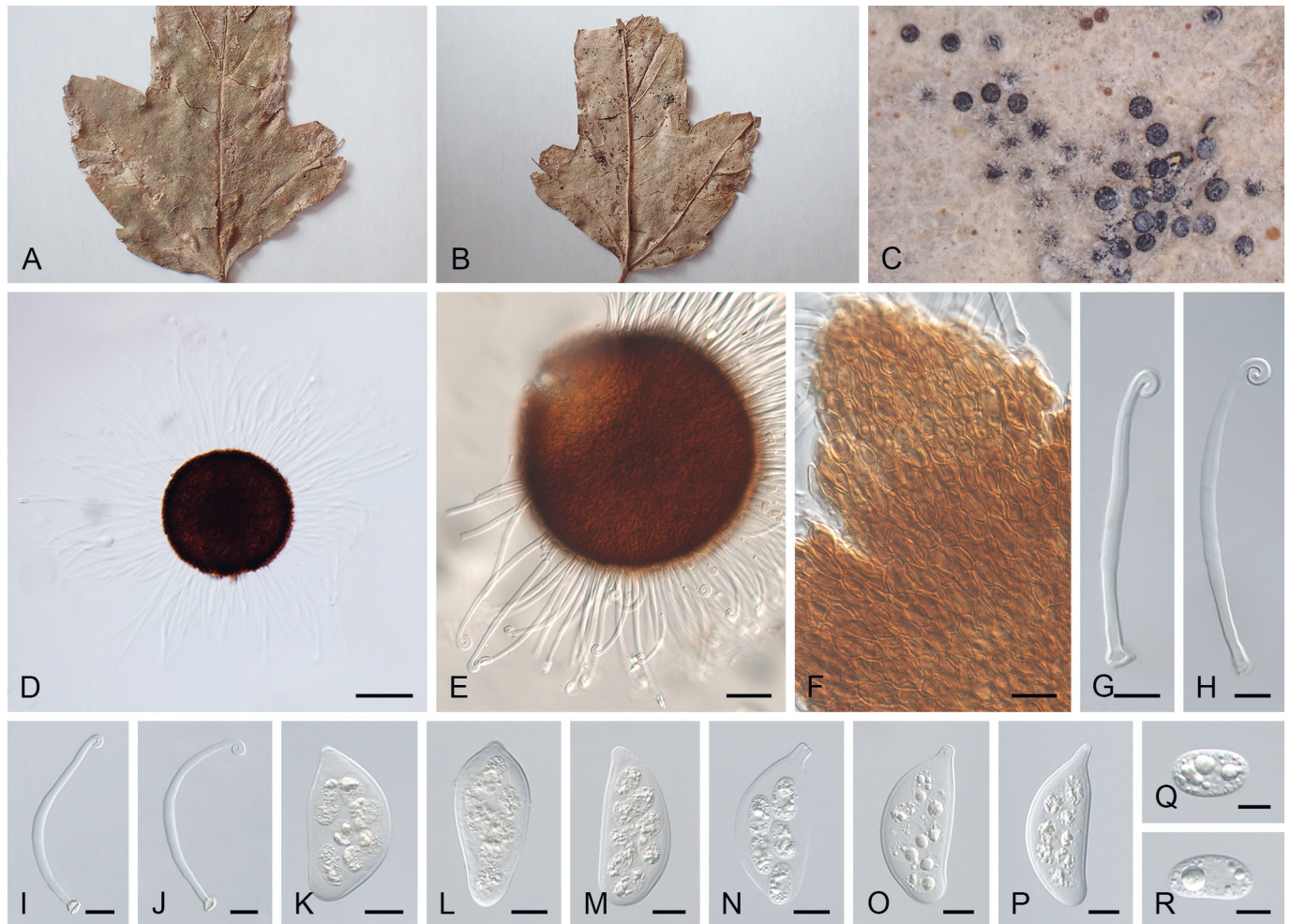


Fig. 15. *Sawadaea nankinensis* 16b6b

16b6b D, E, 16b6b

A, B, C, 16b6b

16b6b

G–J, 16b6b

K–P, 16b6b

Q, R. Ascospores. Scale bars: D = 100 µm; E = 50 µm; F–P

= 20 µm; Q, R = 10 µm.

micro-conidiophores 27.0–59.5 × 5.0–6.5 µm, foot-cells straight, cylindrical, basal septum sometimes elevated, up to 19.5 µm from
 almost globose, 12.0–21.0 × 10.0–20.0 µm, length/width ratio
 macro-conidiophores 90.0–140.0 × 7.0–10.0 µm, followed

macro-conidia

47.0 × 11.0–18.5 µm, length/width ratio 1.3–3.9. *Chasmothecia* hypophyllous, scattered to ± gregarious, large, 233.5–319.5 µm diam. (av. 285.5 µm), globose-lenticular, basal part becoming finally concave; *peridium cells*

17.0(–23.5) µm diam.; *appendages*

145.0–286.0 × 5.0–12.5 µm, *appendages* about 0.5–1.0(–1.2)

asci

100.0(–116.0) × 31.0–52.0 µm, length/width ratio 1.7–3.2 (av. 2.3), mostly short-stalked, stalk about 7.5–17.5(–24.5) × 6.5–10.5(–12.0) µm, rarely sessile, (2–)5–8-spored; *ascospores* ovoid, 18.0–27.0 × 11.0–15.5 µm, length/width ratio 1.2–2.0(–2.4)

Additional materials examined: China,

Acer buergerianum, 21 Oct. 2019, D.N. Jin & X.Y. Wang

Acer buergerianum, 24 Oct.

D.N. Jin & X.Y. Wang

OQ866173); Jiangsu Province, Nanjing City, on *Acer buergerianum*, 29 Oct. 1954, R.Y. Zheng

Acer buergerianum L.

Liu

Host range and distribution: On *Acer buergerianum*, Sapindaceae,

Notes: This species was firstly reported as *Uncinula nankinensis*

Sawadaea,

not mentioned. Only the macro-conidia were described, with germ

orthotubus

Fibroidium

Acer sinopurpurascens

Erysiphe, *Uncinula*

Erysiphe

E. nankinensis et al.

o *Acer buergerianum*

sequences for the first time. Phylogenetic trees were constructed

Sawadaea *Sawadaea*

nankinensis. Moreover, Zheng & Yu (1987) did not find any conidia

in size than previously reported, viz., 233.5–319.5 µm. Zheng

et al.

Acer buergerianum

Sawadaea negundinis Hl. Fac. Agric., Hokkaido Imp.

Univ., Sapporo

Synonyms: *Uncinula negundinis* et al.,

Fungus diseases on cultivated plants of Jilin Province

Sawadaea bicornis

Typus: Japan *Acer negundo*, 21 Oct. 1922,

Homma *holotype*

Mycelium

hyphae

3.5–5.0 µm wide; *hyphal appressoria*

micro-conidiophores 22.5–48.5 × 4.5–7.5 µm, foot-cells 17.0–32.5 × 4.5–7.5 µm, followed by 1–3 shorter cells, basal septum elevated, 4.5–11.0 µm from the junction with the mother

micro-conidia ellipsoid-ovoid, 7.0–12.5(–17.5) × 5.0–10.0(–13.0) µm, length/width ratio 1.1–2.0, with fibrosin bodies; *macro-conidiophores* 30.5–86.0 × 6.0–10.5 µm, arising from the middle or

× 6.0–10.5 µm, followed by 1–3(–4) shorter cells and catenescant

macro-conidia

18.5–31.0 × 11.0–23.5 µm, length/width ratio 1.1–1.8(–2.9), with

fibrosin bodies; *germ tubes*

long, about 10.0–67.5 × 2.5–6.0 µm, *orthotubus*

Fibroidium *Chasmothecia*

247.5 µm diam.; *peridium cells*

5.0–21.0 µm diam.; *appendages*

deeply cleft, first branching point near the middle of the stalk,

60.5–114.5 × 4.0–13.0 µm, *appendages* about 0.4–1 times as long

asci

ovoid, subglobose or irregular, 66.0–82.0 × 30.5–33.0 µm, length/width ratio 2.1–2.6, sessile or short-stalked, 5.5–12.5 × 4.5–9.0 µm

ascospores

almost globose, subcylindrical or irregular, 18.0–20.5 × 10.5–13.5 µm, length/width ratio 1.3–1.8, colourless.

Additional materials examined: China,

Acer negundo Y. Liu & F.Y. Zhao

(HMJAU 02219, ITS rDNA GenBank No.: OR711904, 28S rDNA

Acer negundo Liu

Acer negundo F.Y. Zhao, J.N. Li & J.S. Lu

Acer negundo F.Y. Zhao,

V.N. Nguyen, J.N. Li & J.S. Lu

Acer negundo

G.X. Guan & L. Zhao

Acer triflorum,

L.L. Yang

S.Y. Liu, S. Takamatsu,

OR708548, 28S rDNA GenBank No.: MT462443); Jilin Province,
Acer pictum b mono, 24 Oct. 2010, W.T.
 Jiang (HMJAU 00746, ITS rDNA GenBank No.: OR711903, 28S
 176
 o *Acer negundo*, S.Y. Liu & G.X. Guan
 11
 56 *Acer negundo*, 11 G.X. Guan
 11 11 11
 11 *Acer negundo*, 21
 G.X. Guan & J. Feng
 OQ866174); Jilin Province, Changchun City, on *Acer negundo*,
 11 G.X. Guan & J. Feng
 rDNA GenBank No.: OQ866162); Jilin Province, Changchun City,
 o *Acer negundo*, G.X. Guan & J. Feng
 PM92117, ITS rDNA GenBank No.: OQ866175); Jilin Province,
 11 *Acer negundo*, 21 G.X. Guan &
 J. Feng (HMJAU-PM92118, ITS rDNA GenBank No.: OQ866176);
 11 *Acer triflorum*, 11
 G.X. Guan & J. Feng
 OQ866177); Jilin Province, Gongzhuling City, on *Acer negundo*, P.K.
 Q11 *Acer triflorum*, 4 Oct.
 11 S.Y. Liu, S. Takamatsu, P.L. Qiu & J. Feng

ITS+28S rDNA GenBank No.: OQ866161); Jilin Province, Yanbian
Acer negundo, 13 Oct. 2011, W.T. Jiang
 ITS rDNA GenBank No.: OR708549, 28S rDNA GenBank No.:
 Russia
Acer negundo, 28 Oct. 2018, T.S. Bulgakov
 HMJAU-PM92126, ITS+28S rDNA GenBank No.: OQ866163).

Host range and distribution: On *Acer negundo*, *pictum* & *monotriflorum* Sapindaceae

Notes: 1. Sawadaea Acer negundo
2. Sawadaea bicornis, S. negundinis,
3. S. tulasnei. Acer negundo
6. S. bicornis, S. negundinis
6. S. bicornis, S. negundo
negundinis
Get al. Get al.
Acer negundo
S. bicornis
A. negundo were identified as S. bicornis
S. negundinis

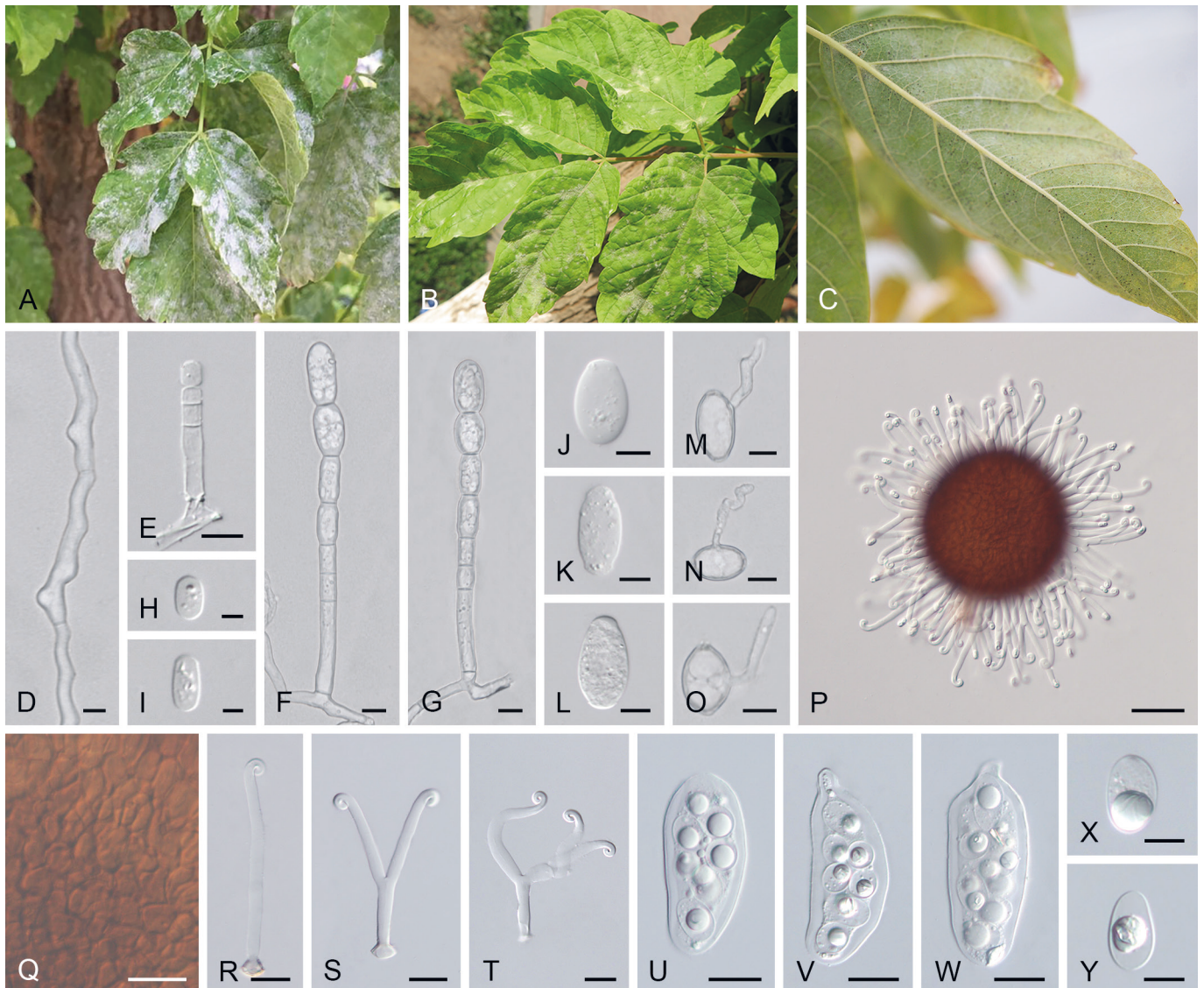


Fig. 16. *Sawadaea negundinis*. B, C. D, E, F, G, H. I, J. K, L, M, N, O, P. Q, R, S, T, U, V, W, X, Y. Ascospores. Scale bars: P = 50 µm; F, G, Q–W = 20 µm; D, E, J–O, X, Y = 10 µm; H, I = 5 µm.

et al.

found significant differences between chasmothecial appendages of *S. negundinis* and *S. bicornis*. *S. negundinis*

S. bicornis

S. bicornis

S. negundinis

A. negundo *S. bicornis*

S. bicornis

S. bicornis

Sawadaea polyfida *S. bicornis*

Microbiol. Sin. 2011

Basionym *Uncinula polyfida* Wankang J.

Synonym: *Sawadaea polyfida* japonica

Mycotaxon 2011

Typus: China, *S. bicornis*

catalpifolium (= *A. catalpifolium*) lectotype

S. bicornis

Mycelium *S. bicornis*

S. bicornis

hyphae straight to sinuous, septate, branched, thin-walled, 3.0–5.0 μm ,

phthal appressoria

S. bicornis

microconidiophores 26.0–39.0 \times 5.0–7.5 μm , arising from the upper

S. bicornis

S. bicornis

\times 5.0–7.0 μm , basal septum sometimes elevated, followed by 1–2

microconidia

shape, ellipsoid-ovoid, 7.5–9.5 \times 6.0–9.0 μm , length/width ratio 1.1–

1.3, (av. 1.2), with fibrosin bodies; macroconidiophores



Fig. 17. *Sawadaea polyfida* *S. bicornis*, C. *S. bicornis*

D, E. *S. bicornis* F, G. *S. bicornis*

R. *S. bicornis* U. *S. bicornis*

= 5 μm .

V–Z. *S. bicornis*

H, I. *S. bicornis*

J–L. *S. bicornis*

M–O. *S. bicornis*

P. *S. bicornis*

A1, B1. Ascospores. Scale bars: Q = 50 μm ; R–Z = 20 μm ; D–I, M–P, A1, B1 = 10 μm ; J–L

× 5.5–9.0 µm, arising from the mother cell, foot-cells cylindrical, straight, 16.0–30.5 × 5.0–8.5 µm, followed by 1–2 shorter cells and macro-conidia

× 11.5–15.5 µm, length/width ratio 1.5–2.0 (av. 1.7), with fibrous germ tubes

Orthotubus fibroidium Chasmothecia scattered to ± gregarious, 129.5–276.5 µm diam.; peridium cells irregularly polygonal, 4.0–24.5 µm diam., arranged ± radially; appendage

apex, apices uncinata to circinate, 28.0–162.5 × 3.5–8.5 (–12.5) µm, 0.2–0.9 times as long as the chasmothecial diam., thick-

asci 8–38, ellipsoid-obovoid, saccate-clavate, (41.0–)70.0–110.0 × (20.0–)46.5–61.5 µm, length/width ratio 1.1–2.3 (av. 1.8), wall thick, 1.8–4.0 µm, almost sessile or with a stalk, 8.5–36.0 × 9.0–17.0 µm, ascospores ellipsoid-ovoid, 12.5–33.0 × 9.5–19.0 µm, length/width ratio 1.1–2.2 (av. 1.7), colourless.

Additional materials examined: China

Acer palmatum, S.R. Tang & L.

Liu

Acer palmatum

dissectum, S.R. Tang & L. Liu

Acer palmatum, Y. Zhang & D.N. Jin

Acer palmatum, S.R. Tang & L. Liu

Acer palmatum, Z.Y. Zhang & D.N. Jin

Acer palmatum, Z.Y. Zhang & D.N. Jin

No.: OR863613); Jilin Province, Wangqing, on *Acer* Y.C. Yang et al.

Acer palmatum

Oct. 2020, Z.Y. Zhang & D.N. Jin

GenBank No.: OR701408); Jiangsu Province, Suzhou City, Qiantang *Acer palmatum*, 9 Oct. 2020, Z.Y. Zhang & D.N. Jin (HMAU-PM92138, ITS+28S rDNA GenBank No.: OR701409);

Acer palmatum, Z.Y. Zhang & X.Y. Wang

Acer palmatum, S.R. Tang & L. Liu

Acer amplum *catalpifolium*, Tao (HMAS13624, 28S rDNA GenBank No.: OQ874975); Yunnan

Acer palmatum, Y. Zhang & D.N. Jin

palmatum, 13 Oct. 2020, Z.Y. Zhang & D.N. Jin (ITS+28S rDNA GenBank No.: OR701411). USA *Acer palmatum*, M. Bradshaw

rDNA GenBank No.: OR166384); Boston, on *Acer palmatum*, M. Bradshaw (OR166379); Boston, on *Acer circinatum*, M. Bradshaw (FH00941966, ITS rDNA GenBank No.: OR166376).

Host range and distribution: On *Acer amplum*, *catalpifolium* [= *catalpifolium*] *japonicum*, *micranthum*, *negundo*, *palmatum* *palmatum*, *palmatum amoenum* [= *amoenum*] *palmatum matsumurae* [= *amoenum matsumurae*] *pseudosieboldianum* [= *takesimensis*] *shirasawanum*, *sieboldianum*, *stachyophyllum* [= *tetramerum*] *tschonoskii* *australe* [= *australe*] *tenuifolium*) Sapindaceae.

Notes: *Uncinula polyfida*

Acer amplum

catalpifolium *Acer*

palmatum *palmatum* *matsumurae*

japonica *palmatum* *matsumurae* (= *A. palmatum* *matsumurae*) (100–)150–200 µm diam. However, owing to the strong variability of

S. polyfida

the Chinese type and Japanese collections is difficult. Hence, Braun

S. polyfida

Sawadaea polyfida

S. polyfida

Acer palmatum *A. circinatum*

A. palmatum *A. pseudosieboldianum*

shirasawanum, *A. sieboldianum*, *A. tenuifolium*) *Acer*

Palmatum

polyfida

Acer circinatum

A. circinatum *Palmatum*

Asia. On the other hand, there are some sequences retrieved from

S. polyfida *Acer*

A. amplum *catalpifolium* *Platanioidea*) *A. micranthum*

A. tschonoskii *australe* *Micrantha*) *A. negundo*

Negundo *stachyophyllum* *Arguta*) *Sawadaea polyfida*

A. pseudosieboldianum (= *A. takesimensis*)

etal. *etal.*

introduction of this species in Europe (Switzerland). Other records

S. polyfida

A. amplum

catalpifolium

S. polyfida *etal.*

would be useful for a further clarification of the phylogeny and

platanoides

polyfida

platanoides

polyfida

are not enough for further attempts. So we tentatively identified it as

S. polyfida. Further study is needed to confirm if *S. polyfida*

platanoides

Sawadaea taii sp. nov.

Etymology:

Typus: China

Acer truncatum, S.Y. Liu, S. Takamatsu,

G.X. Guan & L. Zhao *holotype* *isotype*

Diagnosis *S. bomiensis*

Mycelium

hyphae

septate, branched, thin-walled, $17.0\text{--}34.5 \times 1.5\text{--}6.5 \mu\text{m}$; *hyphal*

apressoria

micro-conidiophores $34.0\text{--}43.0 \times 4.0\text{--}5.5 \mu\text{m}$,

a septum, foot-cells cylindrical, straight, $15.5\text{--}31.5 \times 4.0\text{--}5.5 \mu\text{m}$, basal septum sometimes elevated, to $9.5 \mu\text{m}$ away from the junction

micro-conidia

$5.0\text{--}12.0 \times 4.0\text{--}9.0 \mu\text{m}$, length/width ratio $1.0\text{--}1.9$ (av. > 1), with fibrosin bodies; *macro-conidiophores* $38.0\text{--}50.5 \times 5.5\text{--}8.0 \mu\text{m}$,

a septum, foot-cells cylindrical, straight, $21.5\text{--}39.0 \times 5.5\text{--}8.0 \mu\text{m}$,

macro-

conidia variable in shape, ellipsoid-ovoid, $15.0\text{--}28.0 \times 11.0\text{--}15.5 \mu\text{m}$, length/width ratio $1.1\text{--}1.9$ (av. > 1.5), with fibrosin bodies;

germ tubes

orthotubus

Fibroidium *Chasmothecia*

gregarious, $(115.0\text{--})142.5\text{--}185.0\text{--}(226.0) \mu\text{m}$ diam.; *peridium* cells irregularly polygonal, $4.5\text{--}17.5 \mu\text{m}$ diam., arranged \pm

appendages

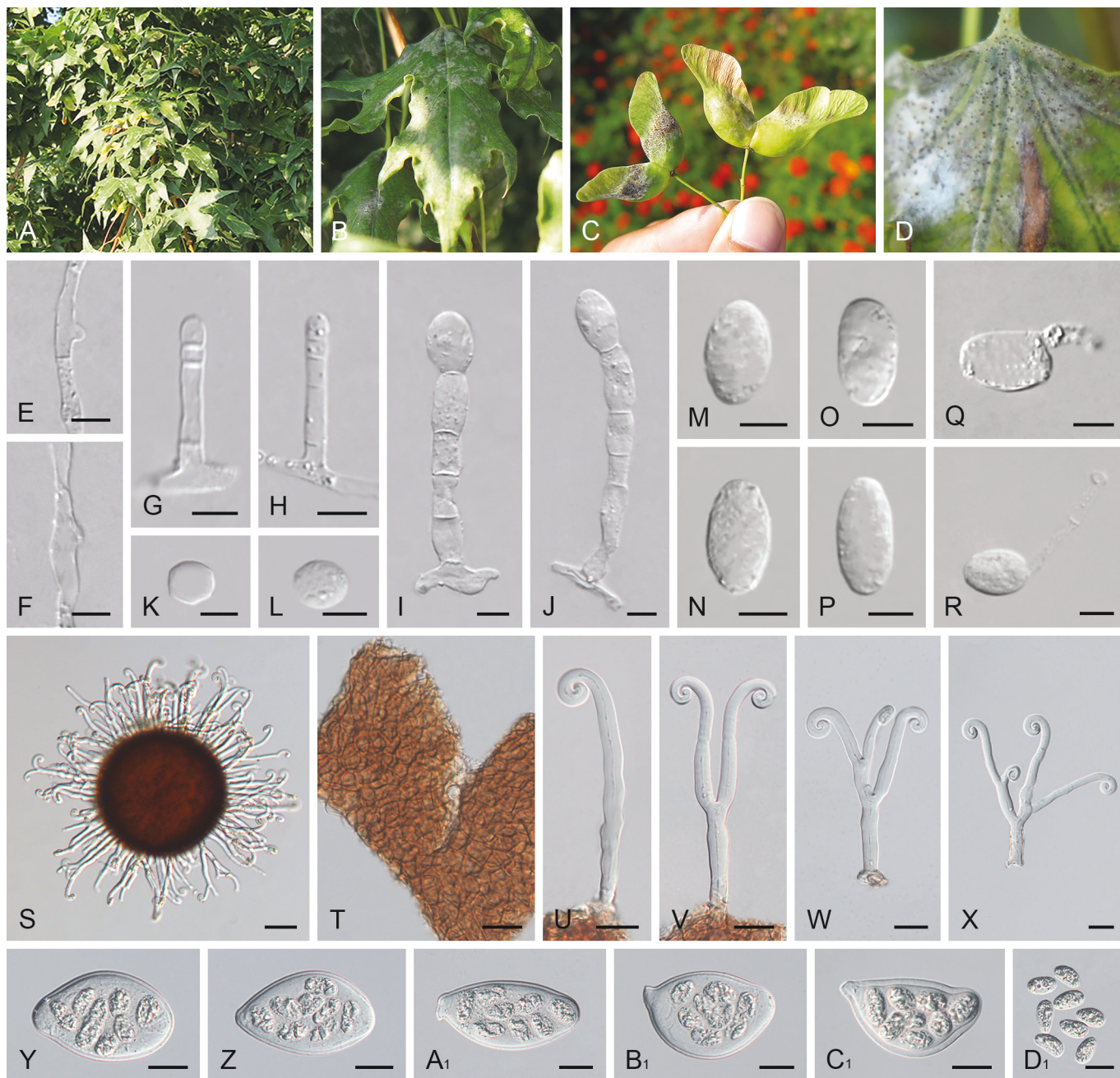


Fig. 18. *Sawadaea taii* sp. nov.

A, B, C, D.

E, F, G, H.

T.

G, H.

U-X.

Y-C1.

I, J.

D1.

K, L.

M-P.

μm ; E-R, D1 = $10 \mu\text{m}$.

70.0–167.5(–225.0) × 4.0–14.0 µm, 0.4–1.3 times (av. 0.8) as long as wide; asci 8-spored, 47.5–68.0 µm, length/width ratio 1.2–1.7 (av. 1.4), wall thick, 1.5–7.0 µm, short-stalked or almost sessile, stalk length up to 7.0 µm; ascospores ellipsoid-ovoid, 16.0–23.5 × 12.5–15.5 µm, length/width ratio 1.1–1.8 (av. 1.4), colourless.

Additional materials examined: China

Liquidambar formosana, 21 Oct. 2018, S.R. Tang & L. Liu

Acer truncatum, Z. Liu rDNA GenBank No.: OQ866158; Jilin Province, Changchun City, *Acer truncatum* L.

Acer truncatum, 2 Oct. 2018, S. Takamatsu, S.Y. Liu, P.L. Qiu & J. Feng

Q.M. Wang OR841072; Yunnan Province, Weixi Lisu Autonomous County, on *Acer barbinerve*, J. Han GenBank No.: OQ866169).

Host range and distribution: On *Acer barbinerve*, *truncatum* Sapindaceae, *Liquidambar formosana*, Hamamelidaceae.

Notes: *Sawadaea* Hamamelidaceae is reported in this study for the first time. *Sawadaea taii*

S. bomiensis, *S. bomiensis*, *S. acerina*, *ginnala*, *Sawadaea taii*, *Acer tataricum*

further attempts. So we tentatively identified them as *S. taii*. A study is needed to confirm if *S. taii* is *tataricum*.

Sawadaea tulasnei Fac. Agric. Hokkaido Univ.

Basionym *Uncinula tulasnei* Fungi Rhen. Exs. Mem. Synonyms: *U. aceris tulasnei* Torrey Bot. Club 90 *U. bicomis tulasnei* Mycologia 44 *Erysiphe varium* Scleromyc. Suec.

Typus: Germany, *Acer platanoides*, Fungi Rhen. Exs., Suppl., Fasc. III, Lectotype

isolectotypes Fungi Rhen. Exs., Suppl.

Germany, BHM

J. Kruse

Mycelium hyphae straight to sinuous, septate, branched, thin-walled, 12.5–45.5 × 2.0–6.5 µm; hyphal appressoria

micro-conidiophores 24.0–53.0 × 4.5–7.0 µm, arising from the mother cell, centrally or × 4.0–7.5 µm, basal septum sometimes elevated, to 9 µm from

micro-conidia ovoid, 6.5–14.0 × 4.0–9.0 µm, length/width ratio 1.0–2.3 (av. 1.4), with fibrosin bodies; macro-conidiophores 27.0–96.0(–141.0) × 5.0–9.5(–11.0) µm, arising from the mother cell centrally or not centrally, foot-cells cylindrical, straight, 13.0–51.0 × 5.0–10.0 µm, basal septum sometimes elevated, to 15 µm from the junction

macro-conidia ovoid, 14.0–49.5(–59.5) × 10.5–17.5(–26.0) µm, length/width ratio 1.0–4.1 (av. 1.7), with fibrosin bodies; germ tubes terminal, orthotubous Fibroidium

Chasmothecia scattered to ± gregarious, 170.0–234.5 µm diam. (av. > 200 µm); peridium cells 18.5(–25.5) µm diam., arranged ± radially; appendages

× 5.5–11.0 µm, 0.3–1.5 times (av. 0.7) as long as the chasmothecial

asci 105.5 × (27.5–)40.0–69.0 µm, length/width ratio 1.3–3.0 (av. 1.7), wall thick, to 5.6 µm, short-stalked or almost sessile, stalked length up to 14.6 µm, 8-spored; ascospores ellipsoid-ovoid, 18.5–30.0 × 14.0–18.5 µm, length/width ratio 1.2–2.0 (av. 1.6), colourless.

Additional materials examined: Canada, Ontario, Niagara Region,

Acer platanoides, V. Ilyukhin PM92145, ITS+28S rDNA GenBank No.: OQ866165; Ontario, *Acer platanoides*, V. Ilyukhin (HMJAU-PM92146); Ontario, Niagara *Acer platanoides*, V. Ilyukhin (HMJAU-PM92147, 28S rDNA GenBank No.: OQ866193). Russia

Acer platanoides, T.S. Bulgakov PM92148, ITS rDNA GenBank No.: OQ866178; Rostov region, *Acer platanoides*, T.S. Bulgakov

Acer platanoides, 24 Oct. 2018, T.S. Bulgakov

Acer platanoides, T.S. Bulgakov ITS+28S rDNA GenBank No.: OQ866166). USA *Acer platanoides*, Pfister GenBank No.: OR166381; Boston, on *Acer platanoides*, M. Bradshaw OR166380).

Host range and distribution (phylogenetically proven hosts): On *Acer platanoides*, *macrophyllum*, *tataricum*

Notes: *Sawadaea tulasnei* on *Acer platanoides*

tulasnei is largely confined

et al. et al.

S. tulasnei on *macrophyllum* (*Acer macrophyllum*)

A. tataricum (*Ginnala*)

et al.

S. tulasnei is not strictly confined

A. platanoides *Platanoides*

S. tulasnei

Acer (*crataegifolium*, *japonicum*, *macrophyllum*, *miyabei*, *monspessulanum*, *cinerascens* [= *cinerascens*], *palmatum*, *A. pictum*, *mayrii* [= *mayrii*], *A. pictum*, *mono* [= *mono*], *platanoides*, *stevenii*, *tataricum*, *tataricum*, *tataricum*, *ginnala* [= *ginnala*], *truncatum*, *ukurunduense*)

A. cappadocicum, *A. monspessulanum*, *turcomanicum*

(= *A. turcomanicum*) et al.

S.

tulasnei

S. tulasnei

S. tulasnei

S. tulasnei

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Takamatsuella taxonomic

manual of the *Erysiphales* (powdery mildews)

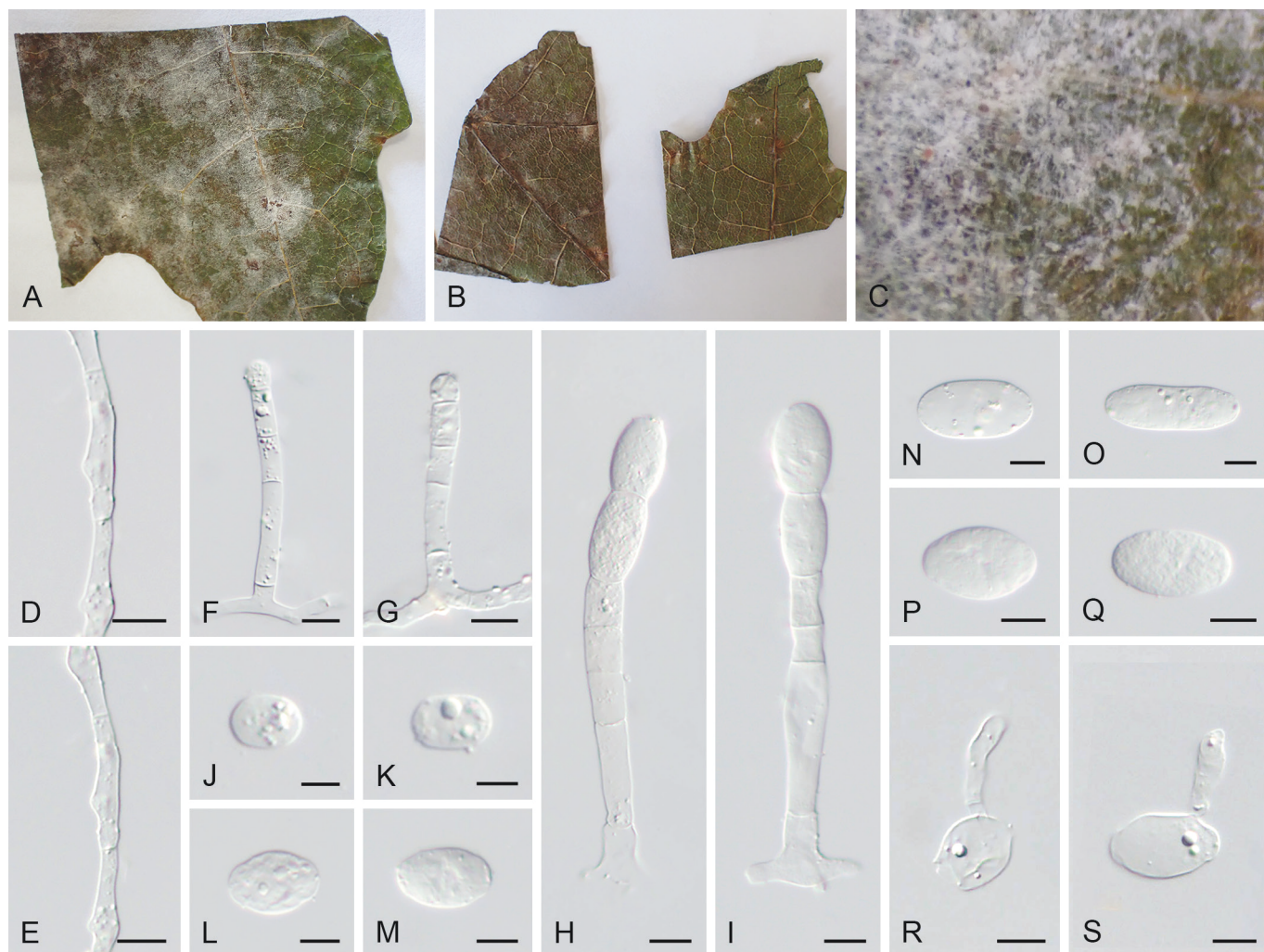


Fig. 19. *Sawadaea tulasnei*. A, B, C, D, E, F, G, H, I, J, K, L, M, N, O, P, Q, R, S. bars: D-I, N-S = 10 µm; J-M = 5 µm.

Type species: *Takamatsuella circinata*

A. Shi (= *Uncinula circinata*)

Takamatsuella circinata

Taxonomic manual of the Erysiphales (powdery mildews)

Basionym: *Uncinula circinata* Bot. 102

Synonym: *Erysiphe circinata*

Schlechtendalia

Typus: USA, *Acer spicatum*

Peck lectotype

isoelectotype

Mycelium

hyphae

sparingly branched, hyaline, thin-walled, smooth, 2.5–8.0 µm

phythal appressoria

conidiophores 27.0–107.5 × 5.0–8.0 µm, arising from mother cells,

conidia

20.0–41.0 (–56.5) × 5.0–9.5 µm, basal septum sometimes elevated, to 10.5 µm from the junction with the mother cell, followed by 1–3

conidia

ellipsoid-ovoid, 17.0–29.0 × 9.5–13.5 µm, length/width ratio

Chasmothecia

(148.5–)236.5–298.5 (–318.0) µm diam., hemispherical to lenticular, flattened or slightly concave in the lower half with age; peridium cells inconspicuous, irregularly polygonal, about 3.5–27.5 µm diam.;

appendages

below the equator, stiff to flexuous, simple, rarely forked, apices tightly uncinately to circinate, 31.0–245.0 × 3.0–8.5 µm, about 0.2–1.0

asci

clavate-saccate, 48.5–100.0 × 20.0–48.0 µm, length/width ratio (1.6–)2.0–3.4 (av. 2.3), sessile or stalked, stalk 5.0–14.5 × 6.0–11.5 µm, (4–)6–8-spored; ascospores ellipsoid-ovoid, 13.0–26.0 × 11.0–15.0 µm, length/width ratio 1.1–2.5 (av. 1.6), colourless.

Additional materials examined: China

Acer sp., 29 Oct. 1954,

B.N. Jiang

Acer wilsonii

, 15 Oct. 1978, R.Y. Zheng

& Y.Q. Lai

Acer pycnanthum

2

M. Bradshaw

OR166378); New Hampshire, on *Acer rubrum*, W.G.

Farlow *Acer spicatum*

, 12 Oct. 1968,

C.T. Rogerson & S.J. Smith

OR166377); Washington, on *Acer*

J.B. Norton

Host range and distribution (Braun & Cook 2012): On *Acer* (*lasycarpum*, *glabrum*, *nigrum*, *pennsylvanicum*, *pycnanthum*, *rubrum*, *saccharinum*, *saccharum*, *spicatum*, *wilsonii*) Sapindaceae; Asia (China), North America (Canada [Manitoba, Ontario, Quebec],

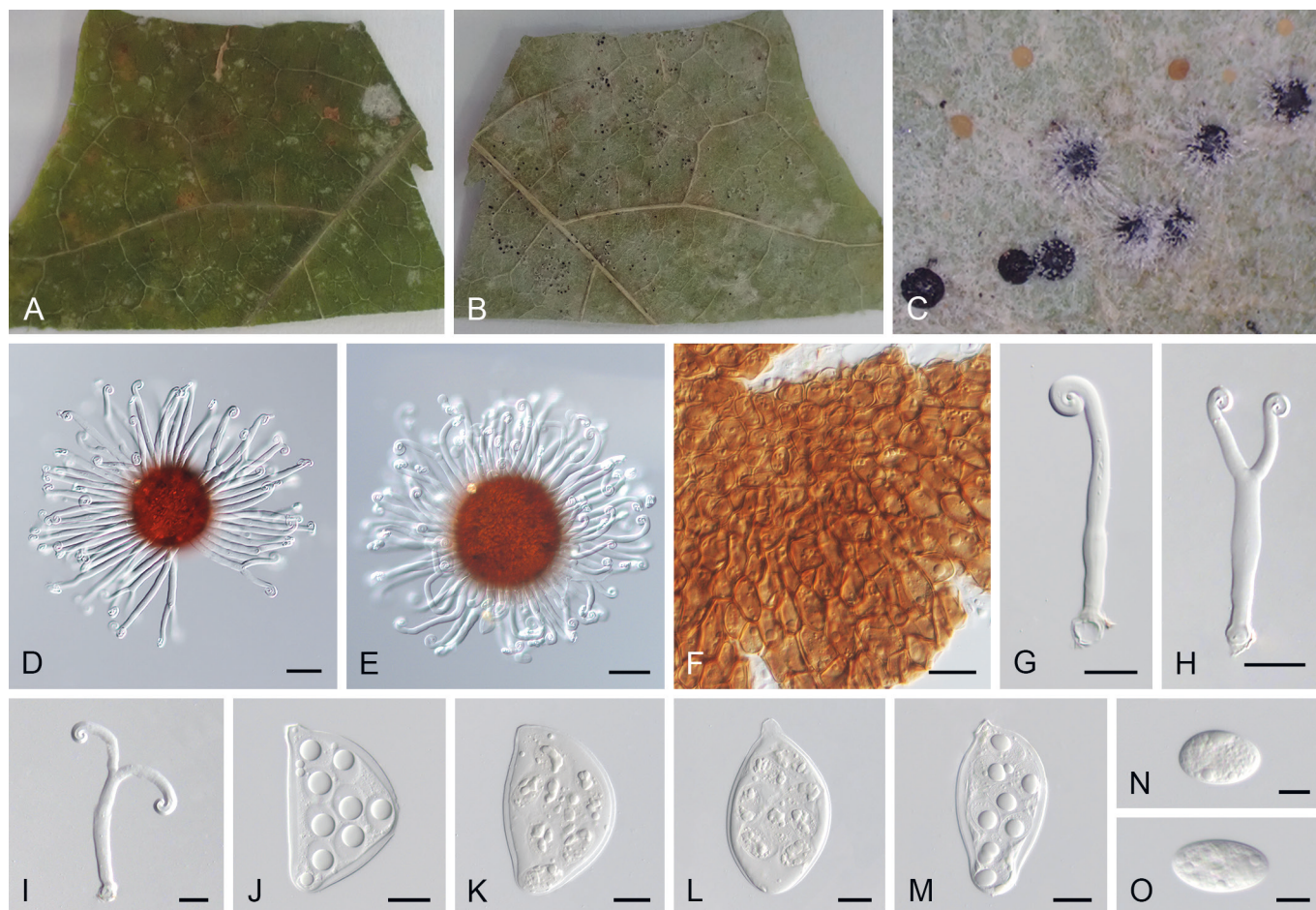


Fig. 20. *Sawadaea tulasnei*

A, E

A

D–I

B

J–M

C

N, O. Ascospores. Scale bars: D, E = 50 µm; F–M = 20 µm;

N, O = 10 µm.

et al. *Uncinula nankinensis* b
 b Sawadaea, *Uncinula koelreuteriae* b Sawadaea
 Sawadaea
 S. bifida d S. kovaliana clarified the two names and allowed us
 S. acerina, S. aceris-arguti, S. taii.
 an important role in the reflection of the phylogenetic relationships
 Sawadaea d Takamatsuella
 Takamatsuella
 Sawadaea. Takamatsuella circinata is
 only species in the genus, and confirmed from China based on
 morphological examinations, although phylogenetic confirmation is
 Sawadaea
 Acer S. koelreuteriae
 Koelreuteria S. aesculi Aesculus
 S. taii Acer Sapindaceae
 d Liquidambar Hamamelidaceae
 Erysiphe necator,
 Vitaceae, Carica
 papaya Caricaceae et al.
 Sawadaea
 acer
 acer
 b Koelreuteria, Aesculus Liquidambar
 Acer buergerianum
 S. nankinensis
 Koelreuteria puniculata S. koelreuteriae
 Aesculus
 chinensis S. aesculi
 acer
 S. polyfida, S. negundinis, S. bifida
 Sawadaea
 b S. bicornis, S. polyfida S. tulasnei S. tulasnei
 Sawadaea
 S. acerina,
 S. bicornis, S. koelreuteriae S. nankinensis
 clades are not yet sufficiently supported, e.g., the two species S.
 bifida d S. negundinis
 et al. S. bifida, Acer pictum Platanoidea
 S. bifida + S. negundinis
 cluster (Clade 6) that possibly reflects an additional cryptic species,

S. tulasnei
 Sawadaea Acer crataegifolium d
 A. rufinerve, Acer Macrantha
 Sawadaea
 o A. ginnala Ginnala
 cases, the identities of the host plants should be checked to confirm
 identifications.
 Sawadaea tulasnei
 S. bifida
 acerina d S. taii Sawadaea bicornis
 et al.
 subclades, one of these is largely confined to Acer campestre
 S. bicornis A. platanoides
 b Acer Platanioidea.
 A. negundo A. pseudoplatanus
 forma speciales
 forma specialis
 as such they do not have types that make such names verifiable.
 Forma speciales
 difficult. As such it is not surprising that results by different authors
 have often been contradictory, difficult to understand, and hard to
 Erysiphe neolycopersici
 et al. Pseudoidium neolycopersici
 render these races objectively verifiable. In addition, the genetic
 such cases warrants the application of an official rank covered by the
 century and the first half of the 20th
 1910). Jaczewski (1927) was the first author in powdery mildews
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- Phytopathogenic Fungi from South Africa. *Australasian Mycologist* 23: 109-114. https://www.australasianmycologicalsociety.com/_files/ugd/86d4e0_14
- More française 6. *The Proceedings of The International Maple Symposium*: 27-28.
- Ascomycetes. *Bothalia* 48: 1-10.
- Methods in Ecology and Evolution 12: 1-10.
- Berichte der Deutschen Botanischen Gesellschaft 12: 1-10.
- Fungal Databases. U.S. National Fungus Collections, ARS, USDA.
- Felsenstein J (1985). Confidence limits on phylogenetics: an approach using the bootstrap. *Evolution* 39: 783-791.
- Systema mycologicum. 3(1). 1-10.
- et al. *Acer* (Sapindaceae): genetic evidence for Out-of-Asia hypothesis with *Scientific Reports* 10: 1-10.
- Gorter G (1988). Identification of South African *Erysiphaceae*. *Phytophylactica* 20: 1-10.
- Acer* - *Acer*. *Plant Systematics and Evolution* 267: 1-10.
- Pyttariales, Helotiales. *Mycological Progress* 20: 1-10.
- Flora of fungi of Ukraine. Powdery mildew fungi. *Ukrainian Botanical Journal* 47: 1-10.
- Heluta VP, Dzhanan VV, Senchylo OO (2016). A first record of a powdery mildew *Sawadaea bicornis* *Acer velutinum*. *Plant Introduction* 9: 1-10.
- Incinula bicornis* *Aesculus hippocastanum*. *Ukrainian Botanical Journal* 41: 1-10.
- Sawadaea tulasnei*. *Mycoscience* 37: 1-10.
- et al. *Sawadaea*, *Erysiphaceae*. *Mycological Research* 109: 1-10.
- Erysiphaceae*. *Journal of the Faculty of Agriculture, Hokkaido Imperial University* 38: 1-10.
- Sawadaea tulasnei* *Acer platanoides*. *Plant Disease* 92: 1-10.
- Erysiphaceae*. *Transactions of the Sapporo Natural History Society* 5: 1-10.
- Karmannyi opredelitel' gribov II. Muchnistorosyanye gribov*. Gosudarstvennogo Instituta Opytnoy Agronomii, Leningrad, 1966.
- et al. *Sawadaea aesculio* *Aesculus chinensis*. *Plant Disease* 103: 1-10.
- et al. toward a new phylogenetic classification of *Leotiomyces*. *IMA Fungus* 10: 1-10.
- Aladağlar and Bolkar Mountains in Turkey. *The Journal of Fungus (Mantar Dergisi)* 11: 1-10.
- Klika J (1923). Monografie českých padlí: příspěvek k mykologickému a fytopatologickému výzkumu Čech. *Masarykova Akademie Práce, Praha* 23: 1-10.
- Kochman J (1960). O nowych dla Polski macziakach prawdziwach. *Acta Agrobotanica* 9: 1-10.
- et al. *Molecular Biology and Evolution* 35: 1-10.
- Acer takesimense* *Sawadaea polyfida*. *The Plant Pathology Journal* 27: 9: 1-10.
- Léveillé JH (1851). Organisation et disposition méthodique des espèces. *Annales des Sciences Naturelles, Botanique, Ser. 3*, 15: 1-10.
- et al. *Journal of Systematics and Evolution* 57: 1-10.
- Species plantarum*, 6(1). *Ukrainian Botanical Journal* 47: 1-10.
- Acer* (Aceroidae, Sapindaceae). *Harvard Papers in Botany* 11: 1-10.
- The Erysiphaceae of Inner Mongolia*. *Journal of Fungal Research* 15: 1-10.
- The Botanical Magazine (Tokyo)* 27: 1-10.
- The Botanical Magazine (Tokyo)* 28: 1-10.
- Mycologia* 92: 1-10.

- Transactions of the Mycological Society of Japan 178
- Transactions of the Mycological Society of Japan 20:
- Taxonomical study of Erysiphaceae of Japan
- Otani (1988). Seiya Ito's Mycological Flora of Japan. Vol. 3. Ascomycotina, No. 2, Onygenales, Eurotiales, Ascosphaeriales, Microascales, Ophiostomales, Elaphomycetales, Erysiphales. 85
- Cultivated plant fungal diseases in Jilin Province
- Deutschlands Kryptogamen-Flora, 1 (Pilze)
- bio.ed.ac.uk/software/figtree>
- et al. *Aesculus assamica* Journal of Plant Pathology 10
- Bioinformatics 19
- Sylloge Fungorum Omnium Hucusque Cognitorum. Vol. I. Patavii: Sumptibus auctoris
- Monograph of the Erysiphaceae
- Sawada K (1914). Classification of the Erysiphaceae Agricultural Experiment Station Formosa Bulletin 9
- Scholin CA, Herzog M, Sogin M, Anderson DM (1994). Identification of group and strain specific genetic markers for globally distributed *Alexandrium* Dinophyceae 30
- Journal of Phycology 30
- Shi A, Mmbaga M (2006). Identification of a powdery mildew pathogen in SNA Research Conference 51:2
- Erysiphaceae of Korea
- Awadadea koelreuteriae* Koelreuteria paniculata. Journal of Microbiology 49
- al. *Pseudoidium pedaliacearum*. Mycological Progress 18
- Awadadea tulasnei* *Acer platanoides* in Oregon. Plant Disease 98:2
- Acer* L. Aceraceae Journal of Plant Research 113
- PAUP*. Phylogenetic analysis using parsimony (*and other methods) version 4.0b10.
- Uncinula* *Acer trifidum* Contributions from the Biological Laboratory of the Science Society of China 64
- Erysiphaceae Bulletin of the Chinese Botanical Society 15
- Erysiphaceae Bulletin of the Chinese Botanical Society 2
- Erysiphaceae Club Bulletin of the Torrey 73
- Sylloge Fungorum Sinicorum Sinensia 30
- Mycoscience 42:9
- Awadadea nankinensis* *Acer buergerianum*. Mycoscience 4
- Erysiphales Journal of Fungal Research 16
- BioTechniques 10
- Awadadea polyfida* *Acer palmatum* Plant Disease 106
- Erysiphaceae Banking Journal 11
- White TJ, Bruns T, Lee S, Taylor J (1990). Amplification and direct PCR protocols: a guide to methods and application
- Acta Microbiologica Sinica: 2
- Awadadea* Acta Microbiologica Sinica 20
- Flora Fungorum Sinicorum. Vol. 1, Erysiphales.