

**Insect conservation, technological traps,
and the fading arts of natural history and field ecology**

Lee A. Dyer¹, Angela M. Smilanich¹, Zachariah Gompert², Matthew L. Forister¹

¹Program in Ecology, Evolution and Conservation Biology, Biology Department, University of
Nevada, Reno, NV 89557

²Department of Biology, Utah State University, 5305 Old Main Hill, Logan, UT 84322

Introduction

The union of modern technology and computational intelligence with conservation could be a Promethean gift to help humans avert impending catastrophes due to climate change, loss of biodiversity, and reductions in the density of key functional groups, including insect pollinators. As insect ecologists continue their struggle to document and protect global biodiversity (Slade and Ong 2023, Diamond et al. 2023, Leandro 2023), we seem to have a cornucopia of powerful methods, including artificial intelligence (AI) species identifications, access to big data, rapid advances in genomics, satellite data, and steadily improving drone technology, yet perhaps we are throwing out valuable tradition while rushing to embrace these tools. Wherefore and wither the naturalist (Futuyma 1998)? Indeed, what is the role of the naturalist or field ecologist in insect science today? As teachers and mentors, are we doing enough to maintain a focus on complex ecological interactions and natural history, especially in the tropics where there is so much work to do (Powers 2024)? Many ecologists are embracing large, synthetic databases and

automated identification methods (van Klink et al. 2022), or even contests to remotely identify as many rainforest species as possible in 24 hours (<https://www.xprize.org/prizes/rainforest>) in areas where most insect species have not been described. At the same time, we may be forgetting the insight garnered by the slow contemplation of an entangled bank (Darwin 1859) or the awe-inspiring observations on a long Malay Archipelago voyage (Wallace 1869). The original methods of natural history and the joys of observing the natural world, collecting insects by hand, and using suites of morphological characters to assign a morphospecies categorization to an observed arthropod are central to taxonomic discovery and ecology. It is our contention that these practices need to remain at the center of any serious conservation effort to document and preserve biological diversity. What use is a list of species obtained by instruments on drones and identified to some taxonomic level with molecular barcodes or machine learning algorithms? Do we actually want a future in which a majority of diversity data are collected and curated with minimal human oversight? Now is the time to consider what would be gained and what would be lost in such a world. These concerns are especially relevant to understanding interaction diversity, which has been the focus of more attention as network science has been better integrated into ecology (Dyer et al. 2010). Here, we present current opinions on how to efficiently quantify biodiversity without abandoning careful natural history and ecology studies on the ground.

Recent reviews and editorials have summarized advances in technology that have putatively advanced or are on the verge of revolutionizing the study of insect biodiversity (e.g., van Klink et al. 2022, Powers 2024), highlighting new methods to collect, analyze, and interpret ecological data. A review by van Klink et al. (2022) focuses on computer vision, acoustic sensors, radar, and molecular methods, and these modern and not-so-modern methods are often

touted as answers to the biodiversity crisis. With these new methods, subdisciplines such as barcoding taxonomy, conservation-omics, and even aeroecology are gaining traction as the focus for diversity research and funding (e.g., Gostel et al. 2022, De León et al. 2023). Combined with recent developments of more powerful drones, new methods do indeed have the potential to advance diversity surveys and inventories by improving speed and capacity (Madden et al. 2022). In fact, drones are an increasingly employed method in ecology used to capture high-resolution images and collect samples from hard-to-reach areas, potentially providing terabytes of data for insects in all terrestrial ecosystems. There is no doubt that these images have provided insights not possible without them. Furthermore, many of the newest tools are seen as almost a requirement for biodiversity research, including AI, environmental DNA (eDNA), DNA barcoding, and big data platforms. Statistical approaches that fall under the umbrella of machine learning are now widely used for analyzing images, identifying species, and automating the initial stages of biodiversity research. Although not novel for insect science, molecular methods, including the use of eDNA and DNA barcoding, are still rapidly evolving and have enhanced species detection and identification efforts considerably. Finally, big data facilitates inferences related to ecological interactions by integrating diverse sources of data, while AI-powered systems are used for classification and species identifications.

A defense of natural history and critique of selected modern methods

Traditional natural history

The observational methods employed by Pliny the Elder (*Naturalis Historia*, 79 CE) two thousand years ago, particularly for Hymenoptera and Lepidoptera, might appear inconsequential or naive when compared to the scientific tools available today. However, despite the promise of

current approaches, traditional methods of studying biodiversity via focused observation, physical collection, and morphological descriptions remain foundational, providing important insights that continue to complement modern techniques. These pillars of insect science are part of basic natural history, and although they are time-intensive, they yield rich data about insect behavior, systematic relationships, and ecological roles. Direct observations of insects and their interactions within ecosystems offer nuanced understandings that technology alone cannot currently capture. In most cases, manually trapping or directly collecting insects should be the primary methods for biodiversity surveys and inventories that are needed for both research and conservation efforts. Morphological analysis remains important for species identification and the description of new species, despite being time-consuming and requiring specialized expertise. Although the issue is of course not unique to insect science, researchers face challenges with complex analyses and the task of understanding and synthesizing the scientific literature that continues to grow at an unmanageable rate. AI, which includes increasingly popular machine learning methods, has promised solutions to these challenges. While the benefits might be obvious, for example for rapid insect identification and the speed with which code can be written in popular statistical languages, the costs are not yet apparent and might be accruing, where depth of consideration is sacrificed for speed of publication (e.g., London, and Kimmelman 2020, Ioannidis 2005, Smaldino and McElreath 2016).

Barcoding

A transformative addition to the insect natural historian's toolbox is the increasing use of molecular methods that have rapidly advanced over the past 50 years, including DNA "barcoding." For insects, barcoding typically involves sequencing a short stretch of the

93 mitochondrial gene for the enzyme Cytochrome Oxidase I (COI), which is then compared to
94 available databases for the purpose of generating species lists for a sample or region, and for
95 contributing to taxonomic revisions and species descriptions (Wilson 2012). However, the
96 reliance on such barcodes has in many areas of study eclipsed traditional methods and
97 surprisingly has also made it difficult to embrace more advanced genetic tools. For example,
98 projects focused on rearing immature insects to get estimates of trophic interaction diversity
99 employ inexpensive methods for estimating this dimension of biodiversity and yield museum
100 specimens that can be used for phylogenomics or conservation research (e.g., reviewed by
101 Salcido et al. 2022). Yet, some have argued forcefully that barcoding is needed for these projects
102 for immediate identification of immature insects or their parasitoids and have even argued that
103 barcoding makes the public more bio-literate (Janzen and Hallwachs 2021). Given the worsening
104 taxonomic impediment (Meier et al. 2024) and the alarming rates of insect species losses,
105 especially in the tropics (Wagner et al. 2021), it is not productive to insist that all studies in insect
106 biodiversity and taxonomy should rely on this or any other one approach that is tied to a
107 particular technology (e.g., Meier et al. 2024). Incomplete taxon sampling is also a serious
108 problem with the barcoding approach to characterizing both communities and building
109 phylogenies (Meyer & Paulay 2005; also see Virgilio et al. 2010), and this is especially relevant
110 to the hyper-diverse insect taxa found in the tropics. It has not been difficult to find examples
111 where the barcode fails to separate recognized taxa of Lepidoptera (Gompert et al. 2006, Forister
112 et al. 2008, Wilson et al. 2012), and issues with barcoding have been widely discussed in the
113 literature (e.g. Rubinoff et al. 2006, Taylor and Harris 2012, Mallo and Posada 2016). Finally, in
114 the areas of the world where biodiversity research is needed the most – tropical rainforests –
115 there is a dearth of sequence data in the BOLD database for most taxa of insects – in fact, BOLD

covers about 4% of the conservative estimate of 5.5 million existing species of insects (van Klink et al. 2022). In these areas that are rich in undocumented biodiversity it makes more sense to prioritize collecting natural history data, getting collections into museums, and pairing molecular with morphological approaches for estimating species and interaction diversities.

Modern population genetics and phylogenetic analyses utilize high-throughput next-generation sequencing (NGS) often paired with reduced representation approaches such as double digest restriction-site (ddRADseq) or genotype-by-sequencing (GBS) to quickly generate information on thousands or tens of thousands of loci. Sequencing approaches are advancing rapidly, and whole genome sequences are now being generated in highly replicated population or systematic studies (Ribeiro and Espíndola 2023, Webster et al. 2023). GBS, for example, offers far greater resolution and scalability compared to traditional barcoding, making it a superior tool for fine-scale population studies and similar applications (Andrews et al. 2016). Seen in that light, the insistence on traditional barcoding does not necessarily offer much above and beyond traditional morphological approaches to species identifications and descriptions (e.g., Chapple & Ritchie 2013), and may in fact be a hindrance to biodiversity research in the extent to which a field becomes anchored to a single, ossified technology. We recognize of course that the picture might be different if an important fraction of insects had actually been "barcoded," but see the estimate of 4% above. Nevertheless, one often hears the informal argument that having molecular data is still better than morphology since it is at least possible to generate a phylogenetic hypothesis using COI data. Among other issues, this argument ignores the problem that the utility of one small fragment of DNA is expected to drop in proportion to the speed and breadth of adaptive radiations, which of course characterize much of insect diversity. This is because ancestral polymorphism is retained across species boundaries in rapidly radiating

groups, which interacts with the problem of mitochondrial introgression through hybridization. In either case, for species lists and rapid interaction diversity assessments, a morphological focus still generates specimens that can be preserved in museums and later used in population genomic, phylogenomics, or genetic diversity studies. There is an appealing irony here that the older, museum-based approach is more flexible and facilitates evolving approaches and methodologies which we of course support, as long as the foundations of our field are not abandoned.

Satellite imagery and modern drone technology

Remote sensing technology includes satellite and airborne sensors, spanning a gradient in methodologies, including diverse types of satellite data, airborne drones, tractor drones, and ground lidar. These approaches offer powerful tools for estimating insect diversity or associated ecological variables, with satellite data providing the highest spatial and temporal resolutions. Enhancements in these technologies have yielded observations of insect habitats and behavior, detailed data on habitat structure, estimates of insect herbivory, measures of light pollution, long-term climate data, microhabitat weather parameters, and detection of insects (Rhodes et al. 2022). Guided by causal hypotheses about how these observed variables are related, these technologies will help us understand how global change parameters are affecting insect populations and diversity and could help with management or conservation decisions.

Similarly, drones are already enhancing insect diversity surveys by enabling the collection of more data from previously inaccessible or challenging environments, especially canopies and other upper layers of forests, which are often rich in insect diversity (de Souza Amorim et al. 2022) but difficult to reach. Drones can also be used to collect foliar canopy samples, spectral data (e.g., Raman spectroscopy, Sharma et al. 2023), and insects via drone-

delivered insect traps (lost Filho et al. 2020). Specialized drone sensors provide a method to collect audio and video data, high resolution images, and eDNA from canopies, phytotelmata, and other hard to reach communities, allowing for indirect detection of diverse insect communities that are otherwise difficult to census. When combined with technological advances summarized here, drones have the potential to significantly enhance biodiversity surveys and inventories.

Clearly, advanced remote sensing technologies delivered via drones and satellite data offer significant advantages for insect diversity surveys and assessments, but traditional trapping and observational methods offer rich information that these technologies alone cannot provide (e.g., de Souza Amorim et al. 2022). Malaise, light, pitfall, and other trapping methods combined with physical collecting and searching allow for the direct observation of seasonal changes and species behaviors, vertical canopy stratification, life history traits, and interactions with other species, all of which are components of functional and interaction diversities – which likely provide more insight into stability and ecosystem function (Dyer et al. 2018). And, in an increasingly urbanized world, the urban bioark is best sampled using these traditional methods (Diamond et al. 2023). Furthermore, questions about insect persistence in the face of global change need to utilize traditional approaches like systematics, assessments of species abundance, and insect disease ecology (Mason and Shikano 2023), all of which require physical sampling for morphological and genetic analyses that remote sensing alone cannot achieve.

Big data

Large, aggregated databases from noninteractive citizen science projects (e.g., Prudic et al. 2023, Plummer et al. 2024), data repositories like the Encyclopedia of Life (EOL), and pooled data

185 from labs worldwide (Forister et al. 2015) represent a unique feature of modern ecology, in
186 which anyone with a smart phone and no training can contribute potentially valuable data
187 through free platforms like iNaturalist. However, there are shortcomings and challenges, even if
188 the methods are relatively standardized (Robinson et al. 2023). The main issues with these
189 approaches are: 1) the lack of direct observations for many aggregated databases; 2)
190 unstandardized methodologies including, in many cases, a lack of absence-data or negative
191 observations; 3) poor or variable quality control; and 4) less time in nature for individual
192 investigators. Direct observations by researchers working with organisms in the field remain the
193 gold standard for understanding ecological interactions (Powers 2024, Dyer et al. 2010),
194 including insect-plant, insect-predator, and insect-soil interactions, and without such natural
195 history, we are more likely to get inaccurate assessments of insect roles and interactions within a
196 focal ecosystem (Dickinson et al. 2010). Lack of standardization for data collected by different
197 citizen scientists or research groups used to create big data can reduce data reliability for
198 meaningful inferences (Bird et al. 2014). Lack of quality control is perhaps less important for
199 combined citizen science datasets that follow a specific method, but many big datasets combined
200 from diverse sources may not include sufficient validations or verification processes,
201 undermining their utility (Kosmala et al., 2016, but see Dyer et al. 2016).

202 Another manifestation of big data involves the aggregation of -omics databases. In
203 contrast to databases generated from dispersed or publicly-sourced observations, the -omics
204 perspective involves the combination of many different types of data, often sourced from
205 different labs with different technologies. For example, this could involve the combination of
206 genomics with metabolomics, proteomics, and even phenomics as the compilation of phenotypic
207 (including morphological) data (Houle et al. 2010). On the one hand, it is impossible to dispute

208 the value to organismal biology that potentially derives from datasets that link such disparate
209 types of information. On the other hand, the value of such aggregation can only be as good as the
210 information that binds different datasets together, which of course brings us back to the need for
211 the most rigorous taxonomic information, which should not be based solely on COI barcoding.
212 Moreover, -omics approaches in ecology and evolutionary biology will often happen at the scale
213 of species, which presents a number of challenges in light of all the issues discussed above. For
214 example, automated AI-based identification will often produce identifications above the species
215 level (e.g., to taxonomic order), which limits the extent to which they can be merged with -omics
216 datasets. Even when species-level identifications are possible, we should be cautious of the
217 extent to which relevant mechanisms can be understood at that level. A contemporary example is
218 the goal of assembling global or regional trait databases for insects which can, in theory, be
219 merged with monitoring databases to investigate relationships between population trajectories (or
220 responses to climate) and ecological traits. We have ourselves contributed to such analyses, but
221 we also acknowledge that variation below the species level is almost certainly required to
222 achieve a meaningful understanding of population biology and adaptation.

223 But the most important argument that should temper our enthusiasm for science based
224 only on big, aggregated or -omics datasets is that they yield less time in nature for the authors of
225 papers using these data. Direct engagement with the natural environment is essential for
226 developing a deep understanding of insect ecology (Powers 2024, Bonney et al., 2014) and for
227 advancing theory, and it is not hard to find examples in which conceptual advances stagnate
228 without fresh insights from the field. For example, the match-mismatch hypothesis was an early
229 expectation from the area of global change biology that warming temperatures would lead to a
230 mismatch in the phenology of consumers and resources. While this might be true for extremely

specialized and obligate interactions, many field biologists would expect spatial dynamics to dilute mismatch effects since most consumers already deal with temporal heterogeneity in the availability of resources across the landscape. And, indeed, based on meta-analysis of careful studies, evidence for the match-mismatch hypothesis is weak (Kharouba and Wolkovich 2023). In other cases, predictions based on theory might be borne out by work with large, aggregated datasets, but explanations cannot be generated without work in the field. For example, another early expectation from global change biology is the idea that geographic ranges will shift with warming temperatures as organisms move to track climatic niches along latitudinal and elevational gradients. Those patterns can indeed be detected based on large-scale datasets, but there is also consensus that direct observations and experiments with wild organisms are essential to understanding mechanisms (see Hsiung et al. 2018 for a discussion in the context of elevational movement). All of this reinforces the value of field experiences and observations, which are not simply complementary to other approaches, but will remain the fount of both the highest quality data and insights leading to new theory.

Conclusion

AI, drones, DNA barcoding, -omics, remotely collected big citizen science databases, and development of rapid diversity assessments are certainly helping efforts to characterize the insect communities found in the most diverse ecosystems such as lowland tropical wet forests. However, technology and methods evolve rapidly, so there should be flexibility that prevents a single methodological approach, such as barcoding, from becoming entrenched. In contrast, the relatively unmodified observational methods of traditional naturalists and field ecologists are still the most important part of modern biodiversity studies. Even when they are overshadowed by the

254 glitter of new approaches, it is our contention that insect biology and biodiversity studies still
255 rely fundamentally on field- and organismal-based knowledge. When combined with less
256 technologically alluring methods or ways of knowing that are centered on traditional natural
257 history, insect ecologists will make substantial progress towards characterizing diversity, setting
258 conservation priorities, and protecting insects (Leandro 2023). These methods can be improved,
259 but not replaced, by modern technologies. Declines in insect diversity will continue despite our
260 best efforts, and they are yet another emerging feature of global change. Battling these declines,
261 mitigating their impacts, and attempting to document multiple dimensions of insect diversity as
262 these dimensions are rapidly degrading will all require multiple concerted approaches (Forister et
263 al. 2024). Certainly, the methods we critique here will be a key part of these battles, but not at the
264 expense of basic natural history.

265 Finally, we ask that we, as a field, seriously consider our long-term goals and always ask
266 if particular technological advances impede or enable progress. As a thought experiment,
267 imagine a world in which remaining natural areas are outfitted with a high density of remotely-
268 operated visual, chemosensory and auditory detectors that report on the identity and presence of
269 all insects that pass near the sensors. This kind of monitoring will by definition focus on adults,
270 which are not always the life history stage with the greatest ecological impact, but we can ignore
271 that for the moment. There is also a massive issue of electronic waste that should be considered,
272 especially when the recycling of such waste is a burden that wealthy countries typically put on
273 the most vulnerable individuals in less developed nations. But, let us assume for the moment that
274 the waste issue can be meaningfully addressed; and let us also assume that the sensors can
275 themselves be made inconspicuous. Then one could argue that the impact on natural ecosystems
276 would only be positive, as it would provide researchers and the public with real-time feedback

on, for example, climate change impacts on wild populations. Or, one can ask if anyone in that future would still care about insects. Human beings have a well-recognized issue with entomophobia that can get worse as people reduce their exposure to the natural world (Gardiner and Roy 2022; Soga and Gaston 2022). Thus an army of drones might inventory a rainforest, or at least generate some kind of tentative catalog (albeit without data on ecological interactions), but does that inventory increase our political or societal motivation to protect the forest? If the answer is no, then we should not fool ourselves that technological advances will solve the human problem that is of course at the root of the biodiversity crisis. In any and all cases, we know that vast hosts of species will be gone before they are known to modern science. It is our contention that knowing fewer of them, by direct observations and with well-curated specimens, will be preferable to knowing more of them with less comprehensive information about each.

In summary, we of course know that insect biology and ecology must take advantage of novel technologies and approaches, some of which are summarized in the other articles collected in this issue. It has not been our goal to belittle the potential value of the newest approaches, rather we want to encourage researchers to critically evaluate all technologies and not assume that new is better especially when so much is at stake.

Literature Cited

Anderson, K., & Gaston, K. J. 2013. Lightweight unmanned aerial vehicles will revolutionize spatial ecology. *Frontiers in Ecology and the Environment* 11:138-146.
<https://doi.org/10.1890/120150>.

298 Andrews, K.R., Good, J.M., Miller, M.R., Luikart, G. and Hohenlohe, P.A., 2016. Harnessing the
 299 power of RADseq for ecological and evolutionary genomics. *Nature Reviews Genetics*,
 300 17(2), pp.81-92.

301 Aucone, E., Kirchgeorg, E., Valentini, A., Pellissier, L., Deiner, K., & Mintchev, S. 2023. Drone-
 302 assisted collection of environmental DNA from tree branches for biodiversity monitoring.
 303 *Science Robotics*. <https://doi.org/10.1126/scirobotics.add5762>

304 Bird, T. J., Moffett, A., & Fiedler, P. L. (2014). Making data count: integrating citizen
 305 contributions. *Nature* 515: 320-321.

306 Bonney, R., Shirk, J. L., Phillips, T. B., Wiggins, A., Ballard, H. L., Miller-Rushing, A. J., &
 307 Parrish, J. K. (2014). Next steps for citizen science. *Science* 343:1436-1437.

308 Cunha, D. G. F., De Moraes, J. S., da Costa, G. M., & Duarte, M. E. R. 2019. Unmanned Aerial
 309 Vehicles (UAVs) in ecological monitoring: A review and recommendations. *Remote*
 310 *Sensing in Ecology and Conservation* 5:205-213. <https://doi.org/10.1002/rse2.100>.

311 De León, L.F., Silva, B., Avilés-Rodríguez, K.J. and Buitrago-Rosas, D., 2023. Harnessing the
 312 omics revolution to address the global biodiversity crisis. *Current Opinion in*
 313 *Biotechnology*, 80, p.102901.

314 de Souza Amorim, D., Brown, B.V., Boscolo, D., Ale-Rocha, R., Alvarez-Garcia, D.M., Balbi,
 315 M.I.P., de Marco Barbosa, A., Capellari, R.S., de Carvalho, C.J.B., Couri, M.S. and de
 316 Vilhena Perez Dios, R., 2022. Vertical stratification of insect abundance and species
 317 richness in an Amazonian tropical forest. *Scientific Reports*, 12:1734.

318 Diamond, S.E., Bellino, G. and Deme, G.G., 2023. Urban insect bioarks of the 21st century.
 319 Current Opinion in Insect Science, 57, p.101028.

320 Dickinson, J. L., Zuckerberg, B., & Bonter, D. N. (2010). Citizen science as an ecological
 321 research tool: challenges and benefits. Annual Review of Ecology, Evolution, and
 322 Systematics, 41 149-172.

323 Forister, M.L., Dyer, L.A., Gompert, Z. and Smilanich, A.M., 2024. Editorial overview: Global
 324 change biology (2023)-Novel perspectives on futures, mechanisms, and the human
 325 element of insect conservation in the Anthropocene. Current Opinion in Insect Science,
 326 62, p.101175.

327 Forister, M.L., Novotny, V., Panorska, A.K., Baje, L., Basset, Y., Butterill, P.T., Cizek, L., Coley,
 328 P.D., Dem, F., Diniz, I.R. and Drozd, P., 2015. The global distribution of diet breadth in
 329 insect herbivores. Proceedings of the National Academy of Sciences, 112(2), pp.442-447.

330 Futuyma, D.J. 1998. Wherefore and whither the naturalist? The American Naturalist, 151:1-6.

331 Gardiner, M.M. and Roy, H.E., 2022. The role of community science in entomology. *Annual*
 332 *review of entomology*, 67(1), pp.437-456.

333 Gompert, Z., Nice, C.C., Fordyce, J.A., Forister, M.L. and Shapiro, A.M., 2006. Identifying units
 334 for conservation using molecular systematics: the cautionary tale of the Karner blue
 335 butterfly. *Molecular ecology*, 15(7), pp.1759-1768.

336 Gostel, M.R. and Kress, W.J., 2022. The expanding role of DNA barcodes: Indispensable tools
 337 for ecology, evolution, and conservation. Diversity, 14(3), p.213.

338 Houle, D., Govindaraju, D.R. and Omholt, S., 2010. Phenomics: the next challenge. *Nature*
339 reviews genetics, 11(12), pp.855-866.

340 Hsiung, A.C., Boyle, W.A., Cooper, R.J. and Chandler, R.B., 2018. Altitudinal migration:
341 ecological drivers, knowledge gaps, and conservation implications. *Biological*
342 *Reviews*, 93(4), pp.2049-2070.

343 Ioannidis, J. P. A. 2005. Why Most Published Research Findings Are False. *PLOS Medicine*,
344 2(8), e124. DOI: 10.1371/journal.pmed.0020124

345 Janzen, D.H. and Hallwachs, W., 2021. To us insectometers, it is clear that insect decline in our
346 Costa Rican tropics is real, so let's be kind to the survivors. *Proceedings of the National*
347 *Academy of Sciences*, 118(2), p.e2002546117.

348 Kharouba, H.M. and Wolkovich, E.M., 2023. Lack of evidence for the match-mismatch
349 hypothesis across terrestrial trophic interactions. *Ecology Letters*, 26(6), pp.955-964.

350 Koh, L. P., & Wich, S. A. 2012. Dawn of drone ecology: low-cost autonomous aerial vehicles for
351 conservation. *Tropical Conservation Science* 5: 121-132.
352 <https://doi.org/10.1177/194008291200500202>.

353 Kosmala, M., Wiggins, A., Swanson, A., & Simmons, B. 2016. Assessing data quality in citizen
354 science. *Frontiers in Ecology and the Environment*, 14:551-560.

355 Leandro, C., 2023. Insect and arthropod conservation policies: the need for a paradigm shift.
356 *Current Opinion in Insect Science*, 58, p.101075.

357 London, A. J., & Kimmelman, J. 2020. Against pandemic research exceptionalism. *Science*,
358 368(6490), 476-477. DOI: 10.1126/science.abc1731

lost Filho, F.H., Heldens, W.B., Kong, Z. and de Lange, E.S., 2020. Drones: innovative technology for use in precision pest management. *Journal of economic entomology*, 113(1), pp.1-25.

Madden, J.C., Brisson-Curadeau, É., Gillung, J.P., Bird, D.M. and Elliott, K.H., 2022. Optimal settings and advantages of drones as a tool for canopy arthropod collection. *Scientific Reports*, 12(1), p.18008.

Mason, C.J. and Shikano, I., 2023. Hotter days, stronger immunity? Exploring the impact of rising temperatures on insect gut health and microbial relationships. *Current opinion in insect science*, 59, p.101096.

Meier, R., Hartop, E., Pylatiuk, C. and Srivathsan, A., 2024. Towards holistic insect monitoring: species discovery, description, identification and traits for all insects. *Philosophical Transactions of the Royal Society B*, 379(1904), p.20230120.

Prudic, K.L., Zylstra, E.R., Melkonoff, N.A., Laura, R.E. and Hutchinson, R.A., 2023. Community scientists produce open data for understanding insects and climate change. *Current Opinion in Insect Science*, p.101081.

Rhodes, M.W., Bennie, J.J., Spalding, A., French-Constant, R.H. and Maclean, I.M., 2022. Recent advances in the remote sensing of insects. *Biological Reviews*, 97(1), pp.343-360.

Ribeiro, T.M. and Espíndola, A., 2023. Integrated phylogenomic approaches in insect systematics. *Current Opinion in Insect Science*, p.101150.

Robinson, M.L., Hahn, P.G., Inouye, B.D., Underwood, N., Whitehead, S.R., Abbott, K.C., Bruna, E.M., Cacho, N.I., Dyer, L.A. Abdala-Roberts, L., and Herbivory Variability

380 Network. 2023. Plant size, latitude, and phylogeny explain within-population variability
 381 in herbivory. *Science*, 382(6671), pp.679-683.

382 Salcido, D.M., Sudta, C. and Dyer, L.A., 2022. Plant-caterpillar-parasitoid natural history studies
 383 over decades and across large geographic gradients provide insight into specialization,
 384 interaction diversity, and global change. In *Caterpillars in the middle: Tritrophic*
 385 *interactions in a changing world* (pp. 583-606). Cham: Springer International Publishing.

386 Sharma, M., Sharma, B., Gupta, A.K. and Pandey, D., 2023. Recent developments of image
 387 processing to improve explosive detection methodologies and spectroscopic imaging
 388 techniques for explosive and drug detection. *Multimedia Tools and Applications*, 82(5),
 389 pp.6849-6865.

390 Slade, E.M. and Ong, X.R., 2023. The future of tropical insect diversity: strategies to fill data
 391 and knowledge gaps. *Current Opinion in Insect Science*, 58, p.101063.

392 Smaldino, P. E., & McElreath, R. (2016). The Natural Selection of Bad Science. *Royal Society*
 393 *Open Science*, 3(9), 160384. DOI: 10.1098/rsos.160384

394 Soga, M. and Gaston, K.J., 2022. Towards a unified understanding of human–nature
 395 interactions. *Nature Sustainability*, 5(5), pp.374-383.

396 Truelove, N. K., Patin, N. V., Min, M., Pitz, K. J., Preston, C. M., Yamahara, K. M., & Chavez, F.
 397 P. 2022. Expanding the temporal and spatial scales of environmental DNA research with
 398 autonomous sampling. *Environmental DNA*, 4:972-984.
 399 <https://doi.org/10.1002/edn3.299>.

400 van Klink, R., August, T., Bas, Y., Bodesheim, P., Bonn, A., Fossøy, F., Høye, T.T., Jongejans, E.,
401 Menz, M.H., Miraldo, A. and Roslin, T., 2022. Emerging technologies revolutionise
402 insect ecology and monitoring. *Trends in ecology & evolution*, 37(10), pp.872-885.

403 Wagner, D.L., Grames, E.M., Forister, M.L., Berenbaum, M.R. and Stopak, D., 2021. Insect
404 decline in the Anthropocene: Death by a thousand cuts. *Proceedings of the National*
405 *Academy of Sciences*, 118(2), p.e2023989118.

406 Webster, M.T., Beaurepaire, A., Neumann, P. and Stolle, E., 2023. Population genomics for
407 insect conservation. *Annual Review of Animal Biosciences*, 11(1), pp.115-140.

408 Wilson, J.J., 2012. DNA barcodes for insects. *DNA barcodes: Methods and protocols*, pp.17-46.

409

410