



OPEN ACCESS

EDITED BY

Brian Gene Coon,
Oklahoma Medical Research Foundation,
United States

REVIEWED BY

Audrey Cleuren,
Oklahoma Medical Research Foundation,
United States
Ran You,
Nanjing Children's Hospital, China

*CORRESPONDENCE

Eno E. Ebong,
✉ e.ebong@northeastern.edu

RECEIVED 02 March 2024

ACCEPTED 27 May 2024

PUBLISHED 04 July 2024

CITATION

O'Hare N, Millican K and Ebong EE (2024).
Unraveling neurovascular mysteries: the role of
endothelial glycocalyx dysfunction in
Alzheimer's disease pathogenesis.
Front. Physiol. 15:1394725.
doi: 10.3389/fphys.2024.1394725

COPYRIGHT

© 2024 O'Hare, Millican and Ebong. This is an
open-access article distributed under the terms
of the [Creative Commons Attribution License
\(CC BY\)](#). The use, distribution or reproduction in
other forums is permitted, provided the original
author(s) and the copyright owner(s) are
credited and that the original publication in this
journal is cited, in accordance with accepted
academic practice. No use, distribution or
reproduction is permitted which does not
comply with these terms.

Unraveling neurovascular mysteries: the role of endothelial glycocalyx dysfunction in Alzheimer's disease pathogenesis

Nicholas O'Hare¹, Karina Millican² and Eno E. Ebong^{1,2,3*}

¹Department of Chemical Engineering, Northeastern University, Boston, MA, United States, ²Department of Bioengineering, Northeastern University, Boston, MA, United States, ³Department of Neuroscience, Albert Einstein College of Medicine, New York, NY, United States

While cardiovascular disease, cancer, and human immunodeficiency virus (HIV) mortality rates have decreased over the past 20 years, Alzheimer's Disease (AD) deaths have risen by 145% since 2010. Despite significant research efforts, effective AD treatments remain elusive due to a poorly defined etiology and difficulty in targeting events that occur too downstream of disease onset. In hopes of elucidating alternative treatment pathways, now, AD is commonly being more broadly defined not only as a neurological disorder but also as a progression of a variety of cerebrovascular pathologies highlighted by the breakdown of the blood-brain barrier. The endothelial glycocalyx (GCX), which is an essential regulator of vascular physiology, plays a crucial role in the function of the neurovascular system, acting as an essential vascular mechanotransducer to facilitate ultimate blood-brain homeostasis. Shedding of the cerebrovascular GCX could be an early indication of neurovascular dysfunction and may subsequently progress neurodegenerative diseases like AD. Recent advances in in vitro modeling, gene/protein silencing, and imaging techniques offer new avenues of scrutinizing the GCX's effects on AD-related neurovascular pathology. Initial studies indicate GCX degradation in AD and other neurodegenerative diseases and have begun to demonstrate a possible link to GCX loss and cerebrovascular dysfunction. This review will scrutinize the GCX's contribution to known vascular etiologies of AD and propose future work aimed at continuing to uncover the relationship between GCX dysfunction and eventual AD-associated neurological deterioration.

KEYWORDS

endothelial glycocalyx, Alzheimer's disease, blood-brain barrier, neurovascular dysfunction, vascular mechanobiology, vascular etiology

Abbreviations: AD, Alzheimer's Disease; A β , Amyloid Beta; BBB, Blood Brain Barrier; CLSM, Confocal Laser Scanning Microscopy; CNS, Central Nervous System; DCE MRI, Dynamic Contrast-Enhanced Magnetic Resonance Imaging; eNOS, Endothelial Nitric Oxide Synthase; GAG, Glycosaminoglycan; GCX, Endothelial Glycocalyx; HA, Hyaluronic Acid; HS, Heparan Sulfate; ICAM-1, Intercellular adhesion molecule-1; IL, Interleukin; MMP, Matrix Metalloproteinase; NFT, Neurofibrillary Tangle; NO, Nitric Oxide; NVU, Neurovascular Unit; RF/FS, Rapid Freezing/Freeze Substitution; RNS, Reactive Nitrogen Species; ROS, Reactive Oxygen Species; SDF, Side-Stream Darkfield Microscopy; TEM, Transmission Electron Microscopy; TPLSM, Two-Photon Light Scanning Microscopy; QUTE-CE MRI, Quantitative Ultra-Short Time-To-Echo Contrast-Enhanced Magnetic Resonance Imaging.

1 Introduction: the role of vascular dysfunction in Alzheimer's disease

Alzheimer's disease (AD) is a neurodegenerative disorder that causes a gradual and irreversible loss of neuronal function in the brain, resulting in progressive impairment of cognitive abilities, such as memory, decision-making, and communication skills (Sheppard and Coleman, 2020). As the disease advances, it leads to increasingly severe symptoms and eventual loss of independence, leading to devastating effects not only for the individual but also their loved ones, caregivers, and society as a whole. In the United States alone, an estimated 6.2 million people over the age of 65 suffer from the condition as of 2021 (Alzheimer, 2021). Unfortunately, the number of cases of AD is projected to increase two to four times by 2050 without significant medical breakthroughs (Alzheimer's disease facts and figures, 2021; Kumar et al., 2022). There are currently no cures for AD, and the only clinical successes have been related to delaying the inevitable cognitive and behavioral decline associated with the disease.

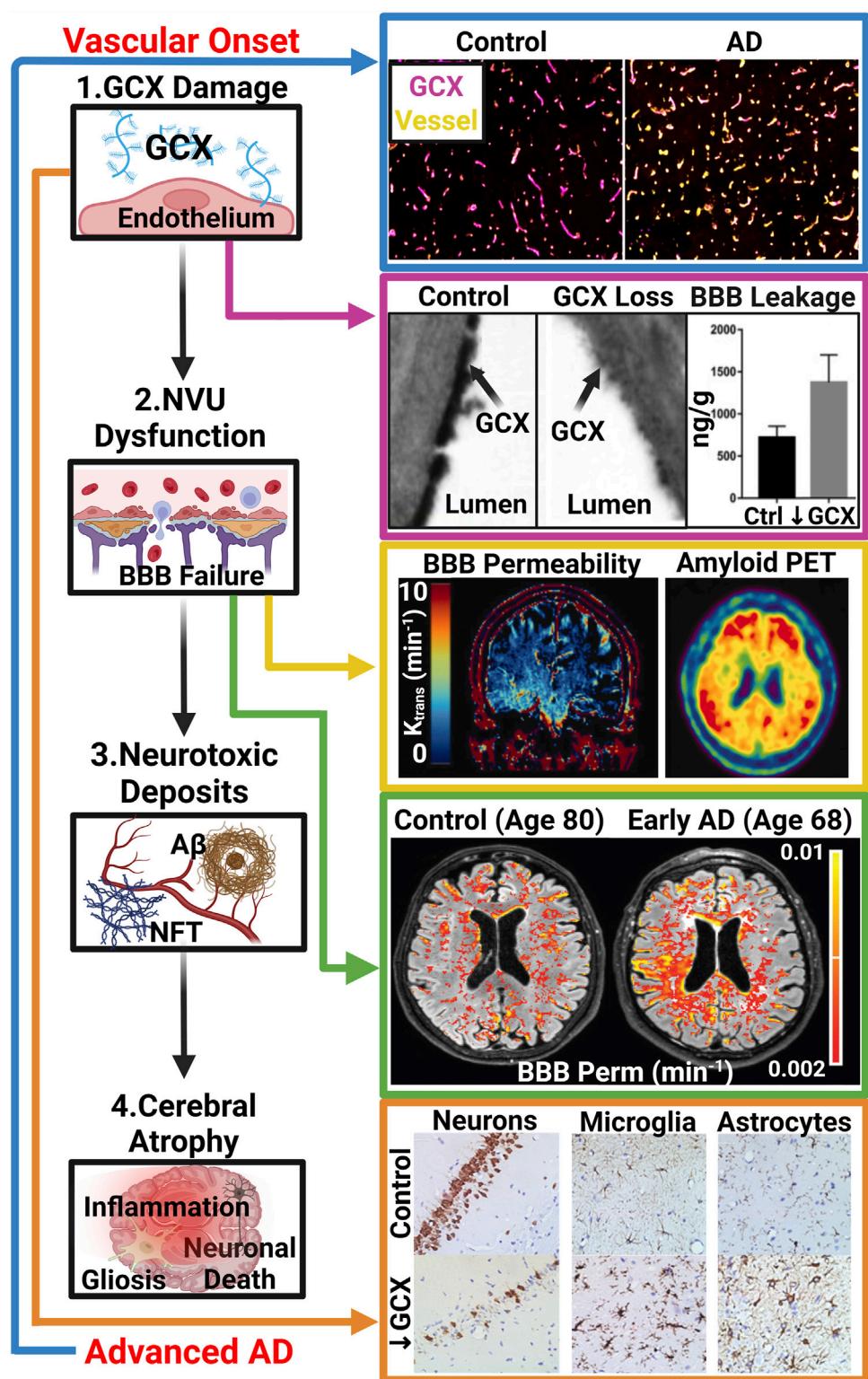
Effective treatments for AD remain elusive due to the complex and poorly understood etiology of the disorder. Traditionally, Amyloid beta (A β) plaques and neurofibrillary tangles (NFTs) are considered the hallmark pathological features of AD (De-Paula et al., 2012). However, targeting these protein aggregates has been challenging, and clinical trials focused on these deposits have shown mixed results, suggesting other mechanisms contribute to the development and progression of AD (Braak and Braak, 1995; Korczyn, 2008; Korczyn, 2012). Thus, it is imperative to investigate additional disease factors for better treatments. This includes exploring the vascular hypothesis of AD, which could significantly advance research and therapies.

Redefining AD as not only a neurological disorder, but rather a combination of vascular abnormalities and neurodegeneration could provide a novel perspective on disease pathology. Despite AD being traditionally considered a non-vascular dementia, virtually all AD patients exhibit impaired vascular function in their brains (De La Torre, 2002). Although this correlation between vascular disease and AD was proposed nearly 30 years ago, the underlying link between vascular dysfunction and AD progression remains relatively understudied (de la Torre and Mussivand, 1993). Epidemiological studies indicate that neurovascular dysfunction often precedes conventional neurocentric pathology in AD patients, offering a promising avenue for novel treatment of AD (Prohovnik et al., 1988; Ruitenberg et al., 2005; Li et al., 2010). This observation is critical to supporting the notion that not only do vascular abnormalities coincide with AD, but they may be promoting the accumulation of toxic protein aggregates in the brain, upstream of symptom manifestation. Future work aims to identify the causal impact of vascular dysfunction to AD symptoms and clarify the weight of these vascular events.

The vascular hypothesis proposes that AD development may originate from neurovascular unit (NVU) breakdown, leading to significant neuronal health changes characteristic of AD. This dysfunction is driven by various interconnected neurovascular pathologies including blood-brain barrier (BBB) breakdown, neurovascular decoupling, and endothelial dysfunction. Structurally, the BBB plays an essential role in facilitating the

transport of nutrients and waste products in and out of the brain's fragile environment (Hussain et al., 2021). When the BBB is damaged, it allows for neuroinflammatory compounds to enter while hindering the clearance of waste products such as AD-associated protein precursors. This leads to protein aggregation, glial inflammation, and neuronal death (Chen and Strickland, 1997; Mhatre et al., 2004; Schachtrup et al., 2007; Sweeney et al., 2018). Congruently, neurovascular decoupling, the process in which the vasculature cannot adequately adapt to the brain's demands, also promotes AD manifestation through inadequate nutrient delivery and impeded waste clearance (Zhu et al., 2022). Finally, endothelial dysfunction within the brain microvasculature induces oxidative stress and cellular inflammation while simultaneously further deteriorating BBB integrity and the responsiveness of blood vessels (Di Marco et al., 2015). All these vascular pathologies have been observed in patients presenting early-stage AD cognitive dysfunction, suggesting their contribution to disease progression (Di Marco et al., 2015; Montagne et al., 2015; van de Haar et al., 2016). Further investigating the impact of these pathologies to AD progression and uncovering the initial insult which causes cerebral vascular dysfunction could dramatically alter treatment approaches to AD.

Although a vascular role appears likely in the pathogenesis of AD, determining a therapeutic target to mitigate or reverse these vascular pathologies remains an unresolved question. Some of the structures at the neurovascular interface under investigation include tight junctions, intracellular transporters, nitric oxide (NO) regulators, and adhesion molecules (Govindpani et al., 2019; Soto-Rojas et al., 2021). However, as of now, no specific blood-vessel based target which significantly modifies the progression of AD has been identified. Thus, in this review, the focus will be drawn to the vascular endothelial glycocalyx (GCX), a relatively understudied structure which stands as a potential master regulator of neurovascular health. The GCX is a sugar-rich nano-structure that resembles a dense bush and lines the inner part of blood vessels (Reitsma et al., 2007). Due to its diverse functionality and structural diversity, it may play a role in regulating several vascular functions that are disrupted in AD, such as stabilizing vessel permeability, promoting neurovascular coupling, and maintaining physiological redox of the endothelium (Curry and Adamson, 2012; Yen et al., 2015; Qu et al., 2021). Research has already demonstrated the significant role of GCX health in preventing various vascular disorders in the body, and recent discoveries regarding its therapeutic potential are rapidly multiplying (Tarbell and Cancel, 2016; Dogné and Flamion, 2020; Zhao et al., 2021). Therefore here, we will present the current knowledge regarding the GCX and its connection to neurovascular health in the context of AD while also outlining future research directions aimed at unraveling its role in disease pathology. Overall, we will propose a GCX-vascular-hypothesis of AD progression where GCX degradation leads to neurovascular unit dysfunction, ultimately promoting abnormal protein accumulation in the brain resulting in AD manifestation (Figure 1). Although still in its early stages and primarily correlational, recent research has notably progressed our comprehension of the connection between cerebral vascular dysfunction and AD, while also shedding light on the involvement of the GCX (Figure 1) (van de Haar et al., 2016; Zhu et al., 2018; Smyth et al., 2022; Moon et al., 2023). However,

**FIGURE 1**

The GCX-Vascular Hypothesis of AD: Hypothesized sequence of events linking GCX degradation to AD pathology: (1.) Various vascular insults induce damage to the GCX. (2.) GCX damage initiates NVU dysfunction, notably affecting the BBB. (3.) NVU dysregulation leads to abnormal accumulation of neurotoxic deposits including A_β and NFTs within the brain, characteristic of AD. (4.) Subsequently, these protein aggregates induce brain inflammation, gliosis, and neuronal death, facilitating cognitive decline. Several recent studies have supported this GCX-rooted hypothesis. (Blue Arrow): Smyth et al. observed a significant reduction in GCX integrity in post-mortem AD tissue compared to healthy controls, suggesting a correlation between late-stage AD and GCX integrity via immunostaining of UEA1 lectin for GCX and Collagen type IV for cerebral vasculature. Adapted from Figure 5 in Smyth LCD, et al. *Acta Neuropathologica Communications*. 2022; 10(1). Copyright (2022) by Acta Neuropathologica Communications (Smyth et al., 2022). This figure is sourced from an open-access publication. (Magenta Arrow): Zhu et al. demonstrated increased leakage of Evans' blue dye into the brain parenchyma (Continued)

FIGURE 1 (Continued)

upon GCX degradation using hyaluronidase treatment in mice, indicating GCX loss triggers NVU dysfunction. Adapted from [Figure 2](#) in Zhu J., et al., *J Cereb Blood Flow Metab.* 2018; 38(11):1979–92, Copyright (2018) by the Journal of Cerebral Blood Flow (32). Used with permission. (Yellow Arrow): Moon et al. showed a regional correlation between increased BBB permeability and AD protein aggregation in early AD patients using DCE MRI and amyloid positron emission tomography. Adapted from [Figure 1](#) in Moon Y., et al. *J Cereb Blood Flow Metab.* 2023; 43(11):1813–25, Copyright (2023) by the Journal of Cerebral Blood Flow ([\(Moon et al., 2023\)](#)). Used with permission. (Green Arrow): Furthermore, van de Haar et al. observed global increases of BBB permeability in patients suffering from early cognitive decline associated with AD compared to controls independent of age. Adapted from van de Haar HJ, et al. *Radiology.* 2016; 281(2):527–35, Copyright (2016) by the Journal of Radiology ([\(van de Haar et al., 2016\)](#)). Used with Permission. (Orange Arrow) Finally, Zhu et al. investigated the impact of GCX degradation on neuronal loss and glial cell activation through immunostaining of neurons (NeuN), microglia (Iba1), and astrocytes (GFAP), revealing significant reductions in neuron count and activated glial cells resembling cerebral pathologies in AD. Adapted from [Figure 5](#) in Zhu J., et al. *J Cereb Blood Flow Metab.* 2018; 38(11):1979–92, Copyright (2018) by the Journal of Cerebral Blood Flow ([\(Zhu et al., 2018\)](#)). Used with permission. While these studies support the proposed cascade, further research is needed to elucidate whether GCX damage is a causative factor in AD manifestation or a consequence of an already pathological state ([De La Torre, 2002; van de Haar et al., 2016; Zhu et al., 2018; Govindpani et al., 2019; Zhao et al., 2021; Smyth et al., 2022; Moon et al., 2023](#)). Created with [BioRender.com](#).

it remains imperative for future research to determine whether the loss of GCX precedes subsequent AD events and elucidate the underlying mechanisms. This review seeks to consolidate existing knowledge regarding the GCX's involvement in advancing neurovascular disease and eventual AD pathology, while also proposing a framework for addressing unanswered questions. In light of the limited therapeutic options available for AD, exploring the GCX for its potential as a novel target presents a source of optimism for improving outcomes in this debilitating condition.

2 The endothelial glycocalyx: a potential master regulator of neurovascular unit function

2.1 What is the cerebral vascular glycocalyx?

The GCX was first visualized by Luft nearly 60 years ago using electron microscopy ([Luft, 1966](#)). Over the past decade, the structure has become increasingly recognized as an essential contributor to vascular homeostasis in health and disease for its diverse roles in vessel permeability, mechanotransduction, and cellular signaling ([Alphonsus and Rodseth, 2014; Uchimido et al., 2019; Potje et al., 2020](#)). In light of accumulating evidence indicating that GCX damage precedes subsequent vascular pathologies, there is a growing focus on exploring the therapeutic potential of this structure upon regeneration not only for cardiovascular disease but neurological conditions as well ([Reed et al., 2019; Yoon et al., 2022](#)). The GCX was only recently considered as an important component of the extended NVU due to its role in regulating neurovascular health ([Stanimirovic and Friedman, 2012](#)). The GCX has been implicated in critical processes within the NVU, including BBB regulation, inflammation control, and NO-mediated neurovascular coupling ([Reitsma et al., 2007; Bartosch et al., 2017; Kutuzov et al., 2018](#)). Based on the GCX's role in these processes, deterioration of the structure may be a precursor to previously described vascular etiologies of AD. Not only are all of these vascular abnormalities present in the majority of AD patients, but initial findings have also indicated that the GCX may be compromised prior to downstream neurological events in these disorders ([Reed et al., 2019; Dogné and Flamion, 2020; Yoon et al., 2022](#)). Protecting and restoring the GCX may therefore be a promising strategy for preventing or treating neurodegenerative diseases.

Structurally, the GCX is composed of numerous proteoglycans, glycoproteins, and associated glycosaminoglycans (GAGs) ([Reitsma et al., 2007; Moore et al., 2021](#)). The major constituents of the GCX include heparan sulfate (HS) with associated core proteins syndecan and glypican, as well as hyaluronic acid (HA) and associated CD44. While these muco-poly-saccharides have been viewed as the main diagnostic markers and mechanistic contributors to GCX function, other complex carbohydrates found at the cell surface including chondroitin sulfate, keratin sulfate, dermatan sulfate, and sialic acid, are also believed to play roles in endothelial function ([Potter et al., 2009; Tarbell and Cancel, 2016](#)). Further investigation is needed to fully differentiate the roles and importance of individual GCX components in regulating NVU physiology and deterring downstream AD pathology ([Pahakis et al., 2007; Yang and Schmidt, 2013](#)).

The nuances of the cerebrovascular GCX compared to other vascular beds are still currently under investigation. This delayed unravelling can be ascribed to the inherent difficulties of imaging a vascular nanostructure imbedded within the skull. However, recent microscopy advances have allowed for visualization of the structure like never before. Preliminary two-photon light scanning microscopy (TPLSM) in mice has demonstrated a robust arterial and capillary GCX along the cerebral arterial tree with a thickness that varies with vessel type and did not correlate with vessel diameter ([Yoon et al., 2017](#)). Interestingly, capillaries exhibited the thickest GCX in relation to vessel diameter perhaps based on the microcirculation being the prominent location of BBB transport. Utilizing another novel imaging modality in side-stream darkfield microscopy (SDF) in human patients, Haeren et al. observed a thicker GCX in cerebral vasculature compared to sublingual vasculature, indicative of the GCX's heterogeneity throughout the body and its prospective critical role to brain physiology ([Haeren et al., 2017; Haeren et al., 2018](#)). While still in its early stages, the ongoing endeavors to differentiate and unravel the intricacies of the cerebrovascular GCX offer promising prospects for future understanding.

2.2 Potential role of the endothelial vascular glycocalyx as a blood-brain barrier function controller

Mounting evidence underscores the pivotal role of the GCX in neurovascular regulation. Further understanding of this relationship

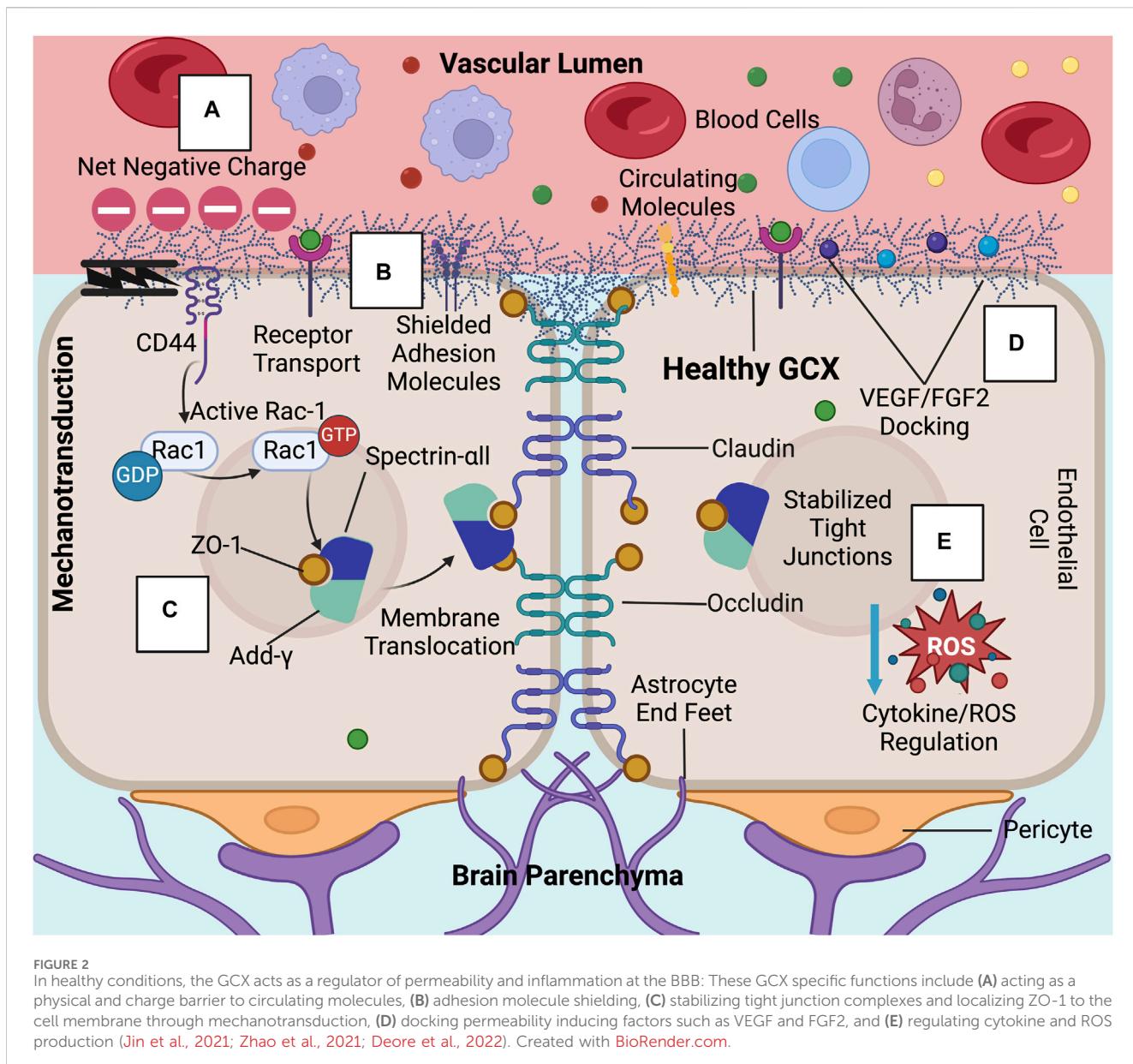


FIGURE 2

In healthy conditions, the GCX acts as a regulator of permeability and inflammation at the BBB: These GCX specific functions include (A) acting as a physical and charge barrier to circulating molecules, (B) adhesion molecule shielding, (C) stabilizing tight junction complexes and localizing ZO-1 to the cell membrane through mechanotransduction, (D) docking permeability inducing factors such as VEGF and FGF2, and (E) regulating cytokine and ROS production (Jin et al., 2021; Zhao et al., 2021; Deore et al., 2022). Created with BioRender.com.

could prove pivotal in combatting AD and related neurodegenerative disorders. First, the integrity of the GCX is essential to the regulation of vascular flux throughout the body, and its role in BBB permeability is increasingly recognized (Zhao et al., 2021). Traditionally, it was believed that BBB permeability was mainly regulated through tight junctions and adherens junctions, for paracellular transport, and membrane channels, pumps, and carrier proteins, for transcellular transport pathways. However, the importance of a robust GCX is only now being appreciated for its relevance to maintaining overall BBB integrity (Zhu et al., 2018; Yang et al., 2021; Zhao et al., 2021). Research has indicated dramatic increases in vascular permeability upon GCX structure damage, strongly supporting the notion that GCX health not only plays a correlative role but a causal one in regulating overall BBB permeability and thus downstream AD pathology (Reed et al., 2019; Jin et al., 2021).

Structurally acting as a physical and charge barrier, the GCX serves as a molecular sieve to the BBB, preventing the entry of neuroinflammatory molecules into the brain while simultaneously allowing essential factors to pass through (Woodcock and Woodcock, 2012; Zhao et al., 2021). The GCX's role in deterring neurotoxic macromolecules present in the plasma, such as albumin, prothrombin, and plasminogen, may be critical to preventing the progression of AD through deterring the entry of neuroinflammatory molecules (Vink and Duling, 2000; Yang and Schmidt, 2013; Kadry et al., 2020). The GCX also regulates BBB permeability due to the high degree of net negative charge on the GAGs HS and chondroitin sulfate as well as sialic acid sugar chains (Walter et al., 2021) (Figure 2, 3). In enzymatic studies, removal of sialic acid residues on these chains resulted in an increase in albumin infiltration likely due to the reduction in electrostatic resistance (Betteridge et al., 2017). Overall, the GCX's dual functionality as a

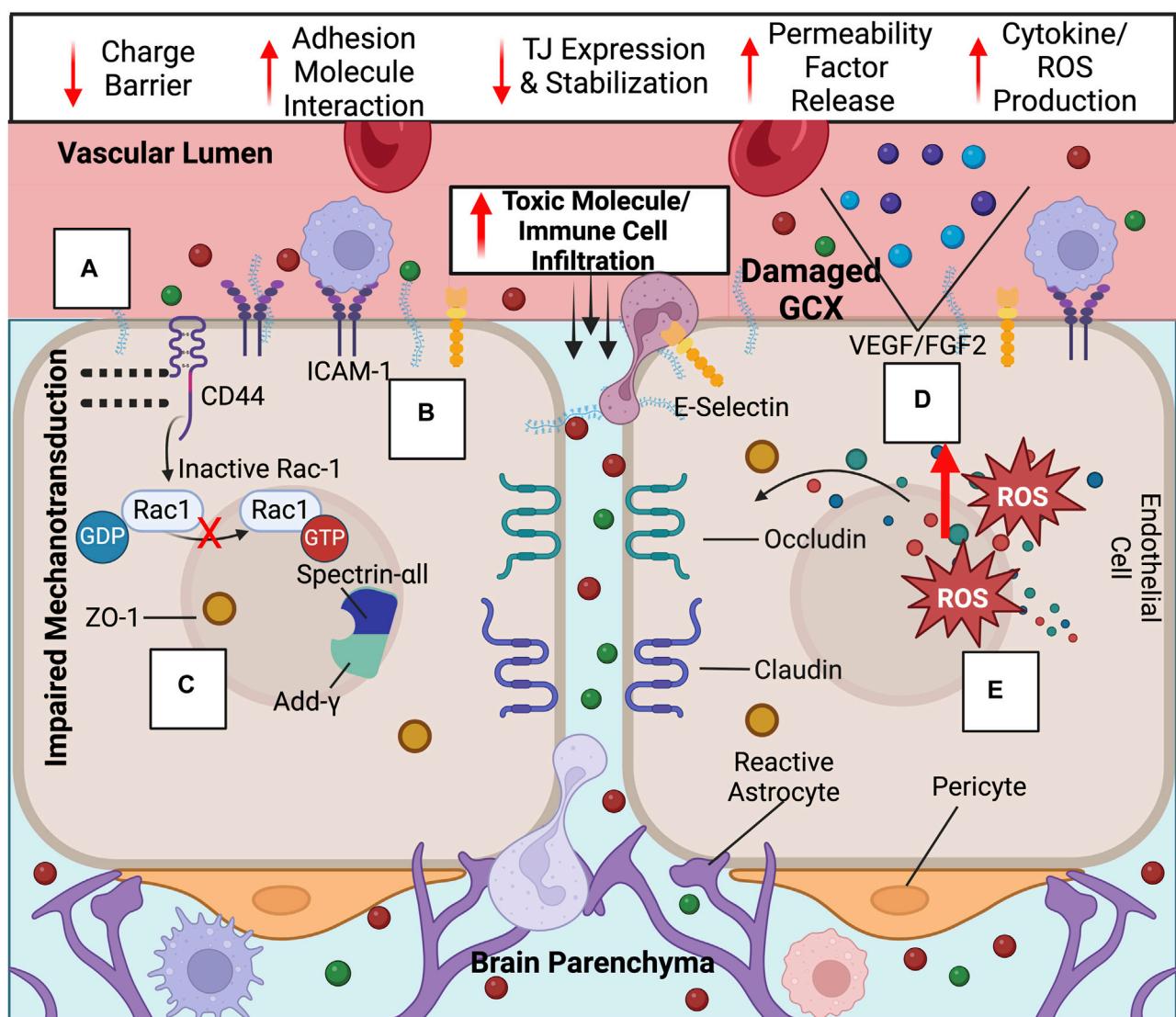


FIGURE 3

Upon GCX degradation, a variety of transport mechanisms are compromised at the BBB, leading to ultimate increased neurotoxic molecule and immune cell infiltration into the brain: Compromised endothelial functions include the loss of the (A) GCX's negative charge barrier, (B) adhesion molecule upregulation and exposure, (C) decreased tight junction expression and ZO-1 membrane localization impairment due to failed mechanotransduction, (D) permeability factor release, and (E) cytokine/ROS production. Created with [BioRender.com](https://biorender.com).

physical and charge barrier underscores its significance in regulating molecular access to the central nervous system (CNS).

The GCX plays a crucial role in maintaining the BBB not only through its physical structure but also by influencing cellular signaling which impacts BBB function. It is known to regulate the production and availability of reactive oxygen species (ROS) and cytokines, which are both linked to the disruption of tight junctions such as claudins and occludins at the BBB (Figure 2, 3) (Goes et al., 2001; Rochfort et al., 2014). Additionally, a thinner GCX is associated with reduced expression of crucial transcellular transporters compromised in AD, such as glucose transporter-1 and p-glycoprotein, essential for glucose transport into the brain and A_β plaque clearance from it, respectively (Gomez González et al., 2011; Winkler et al., 2015; Ding et al., 2021; Santa-Maria et al., 2021). In a study directly examining the effects of GCX impairment on BBB

function, Zhu et al. found that hyaluronidase-induced degradation of HA in a rat model led to increased infiltration of Evan's blue dye into the CNS, reduced tight junction expression, elevated brain water content, and glial activation indicative of neuroinflammation (Figure 1) (Zhu et al., 2018). This highlights the potential involvement of the GCX in BBB permeability regulation and key AD promoting pathologies.

Further research is needed to determine the molecular mechanisms linking GCX loss to BBB disruption and the functions of specific components within the complex, heterogeneous structure. For example, HA removal may trigger a CD44-dependent mechanism of BBB regulation (Al-ahmad et al., 2019). CD44, the primary transmembrane receptor for HA, has been shown by Deore et al. to be critical for BBB formation (Deore et al., 2022). When CD44 is deleted through CRISPR, RAC-1 activation is suppressed whereas

RhoA becomes hyperactivated. Consequently, this prevents the initiation of a pathway that involves the recruitment of ZO-1, the actin filament capping protein adducin- \square , and the cytoskeletal protein spectrin- α II to cell-cell junctions, ultimately impairing the integrity of the BBB. (Figures 2, 3). This pathway is dependent on shear-stress-based mechanotransduction, likely due to HA's role as a mechanosensor (Deore et al., 2022). The interplay of HA, CD44, shear stress, and tight junctions is crucial for maintaining a stable BBB.

Additionally, for HS, which is the most abundant GAG in the GCX, studies show that its enzymatic degradation increases monocyte infiltration into the brain (Floris et al., 2003). HS serves as a docking site for chemokines as well as various other molecules such as fibroblast growth factor 2 (FGF2) and vascular endothelial growth factor (VEGF), all involved in regulating tight junction activity (Figures 2, 3) (Chiodeli et al., 2015; Yang et al., 2021). Furthermore, HS proteoglycans, such as syndecan-1 and glycan-1, may also play critical roles in BBB permeability. Degradation of the GCX enhances syndecan-1 interaction with SRC, promoting caveolae-mediated endocytosis, while glycan-1 loss may reduce A β transport out of the brain via complex formation with low-density lipoprotein receptor-related protein 1 (LRP1) (Leite et al., 2020; Zhu et al., 2021).

Ultimately, gaining a deeper understanding of the mechanisms underlying BBB permeability induced by GCX loss and further elucidating the roles of its individual components could provide valuable insights into its relevance to the BBB-based neurodegeneration associated with AD.

2.3 Potential role of the endothelial glycocalyx as an inflammation controller for the neurovascular unit

While acting as a BBB permeability regulator, the GCX also serves as a key modulator of neuro- and vascular-inflammation. Alterations in immune cell trafficking and endothelial function may lead to AD-related pathologies including A β accumulation, oxidative stress, and neuroinflammation (Govindpani et al., 2019). A feedback loop of neurological and vascular dysfunction results in a severely diseased state of cognitive decline. Could GCX dysfunction precede these downstream events?

In physiological conditions, the GCX plays a crucial role in regulating leukocyte adhesion to the endothelium. The GCX in rat capillaries is believed to range from 200–500 nm, far exceeding the size of endothelial leukocyte receptor and adhesion molecules, such as E-selectin and intercellular adhesion molecule-1 (ICAM-1) (30–40 nm) (van den Berg et al., 2003; Lipowsky, 2012). Due to this size discrepancy, one can logically hypothesize the GCX shields the endothelium from direct interaction with circulating white blood cells (Figures 2, 3). Constantinescu et al. demonstrated that after various heparinase (an enzymatic cleaver of HS) concentrations were administered to mice, the number of adherent leukocytes increased significantly (Constantinescu et al., 2003). Not only does the GCX shield endothelial cells from leukocyte interaction with adhesion molecules, but it may also upregulate adhesion molecule expression upon shedding. In an *in vitro* flow environment, ICAM-1 levels increased by 300% upon enzymatic removal compared to static controls. This increase may be mediated

by a shear-induced NF- κ B pathway regulated by GCX components (McDonald et al., 2016).

In our lab group, we have performed broad RNA-SEQ analysis on human aortic endothelial cells to determine the effects of HS degradation on eliciting endothelial inflammation, a state associated with downstream vascular disease (Figure 4) (Harding et al., 2019a; O'Hare et al., 2022). Through analysis of 60 relevant pro-inflammatory markers, 40 of 60 were upregulated upon HS cleavage via Heparinase III treatment. To our knowledge, no study has characterized the effects of HS loss on such a wide range of inflammatory markers implicated in endothelial activation including adhesion molecules, cytokines/chemokines, and pro-inflammatory transcription factors (Figure 4). This work provides further evidence of the pronounced role HS plays in deterring endothelial dysfunction. However, the significance of HS and other GAGs within the GCX for maintaining normal endothelial function in the cerebral vasculature remains underexplored and warrants further investigation to elucidate their roles in mitigating neurovascular dysfunction associated with AD.

2.4 Potential role of the endothelial glycocalyx as a facilitator of neurovascular coupling through nitric oxide production

A final key function of the GCX which links it to AD pathology is its role in preventing neurovascular decoupling. Neurovascular decoupling, characterized by an impaired link between neuronal activity and cerebral blood flow, is a prominent vascular pathology in AD (Zhu et al., 2022). Although the brain accounts for only 2% of the body by weight, it demands 15%–20% of the body's cardiac output, requiring tightly coordinated neurovascular coupling to facilitate its extensive and dynamic nutrient demands (Williams and Leggett, 1989). In AD, there is compelling evidence indicating a notable decrease in essential nutrients reaching the brain and a rise in the accumulation of neurotoxic deposits. This is believed to stem from the vasculature's inability to meet the brain's transport demands, thus decoupling the systems (Hock et al., 1997; Park et al., 2005). Furthermore, studies aimed at recovering neurovascular coupling resulted in improved cognitive function in AD mouse models, suggesting a causal link to the disease (Tong et al., 2012; Tarantini et al., 2017). Based on this evidence, proper blood-brain signaling is essential to preventing later-stage AD.

The GCX is an essential contributor to preventing AD-associated neurovascular decoupling through regulating the mechano-physiology response of blood vessels in the cerebral vasculature. In particular, it plays a critical role in NO production, the key molecule responsible for vasodilation and subsequent neurovascular coupling (Yen et al., 2015; Dormanns et al., 2016). Endothelial NO generation relies on GCX mechanotransduction, which is compromised when the GCX is degraded (Figures 5, 6). When the GCX is intact, HS senses shear stress applied to the vascular wall and stimulates its attached core protein, glycan, to facilitate a wide range of cellular responses essential to NO production including the opening of calcium ion channels and the activation of the PI3K pathway (Tarbell and Pahakis, 2006; Kumagai et al., 2009; Ebong et al., 2014; Zeng et al., 2015). Although syndecans and glycans bind HS, glycans have demonstrated greater importance to the NO pathways potentially due to their propensity to reside in the caveolae, small invaginations in the cell membrane which associate

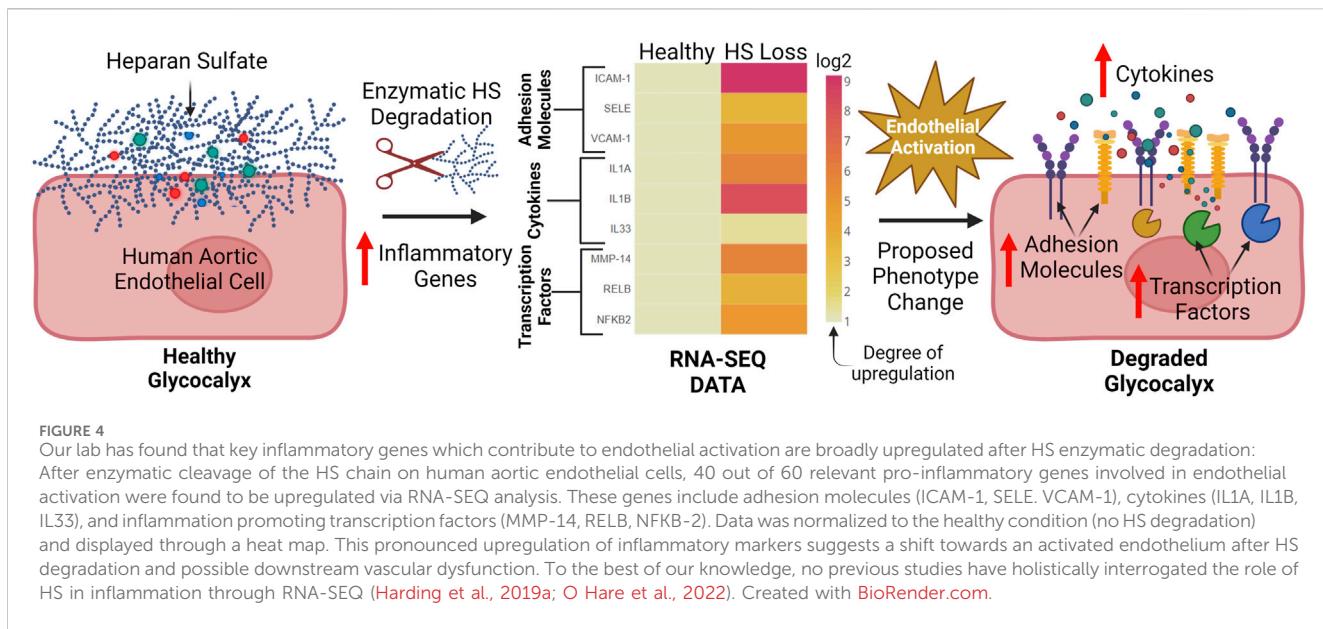


FIGURE 4

Our lab has found that key inflammatory genes which contribute to endothelial activation are broadly upregulated after HS enzymatic degradation: After enzymatic cleavage of the HS chain on human aortic endothelial cells, 40 out of 60 relevant pro-inflammatory genes involved in endothelial activation were found to be upregulated via RNA-SEQ analysis. These genes include adhesion molecules (ICAM-1, SELE, VCAM-1), cytokines (IL1A, IL1B, IL33), and inflammation promoting transcription factors (MMP-14, RELB, NFKB2). Data was normalized to the healthy condition (no HS degradation) and displayed through a heat map. This pronounced upregulation of inflammatory markers suggests a shift towards an activated endothelium after HS degradation and possible downstream vascular dysfunction. To the best of our knowledge, no previous studies have holistically interrogated the role of HS in inflammation through RNA-SEQ (Harding et al., 2019a; O'Hare et al., 2022). Created with BioRender.com.

endothelial nitric oxide synthase (eNOS) (Ebong et al., 2014). With this mechanically transduced increase in Ca^{2+} , a calcium-calmodulin complex induces a conformational change to eNOS, while PI3K phosphorylates the enzyme, both processes ultimately facilitating eNOS activation (Figures 5, 6). Activated eNOS produces NO by catalyzing a reaction that converts the amino acid L-arginine and oxygen into L-citrulline and NO. Consequently, vascular tone is regulated through the release of NO, a potent vasodilator, leading to normal neurovascular coupling.

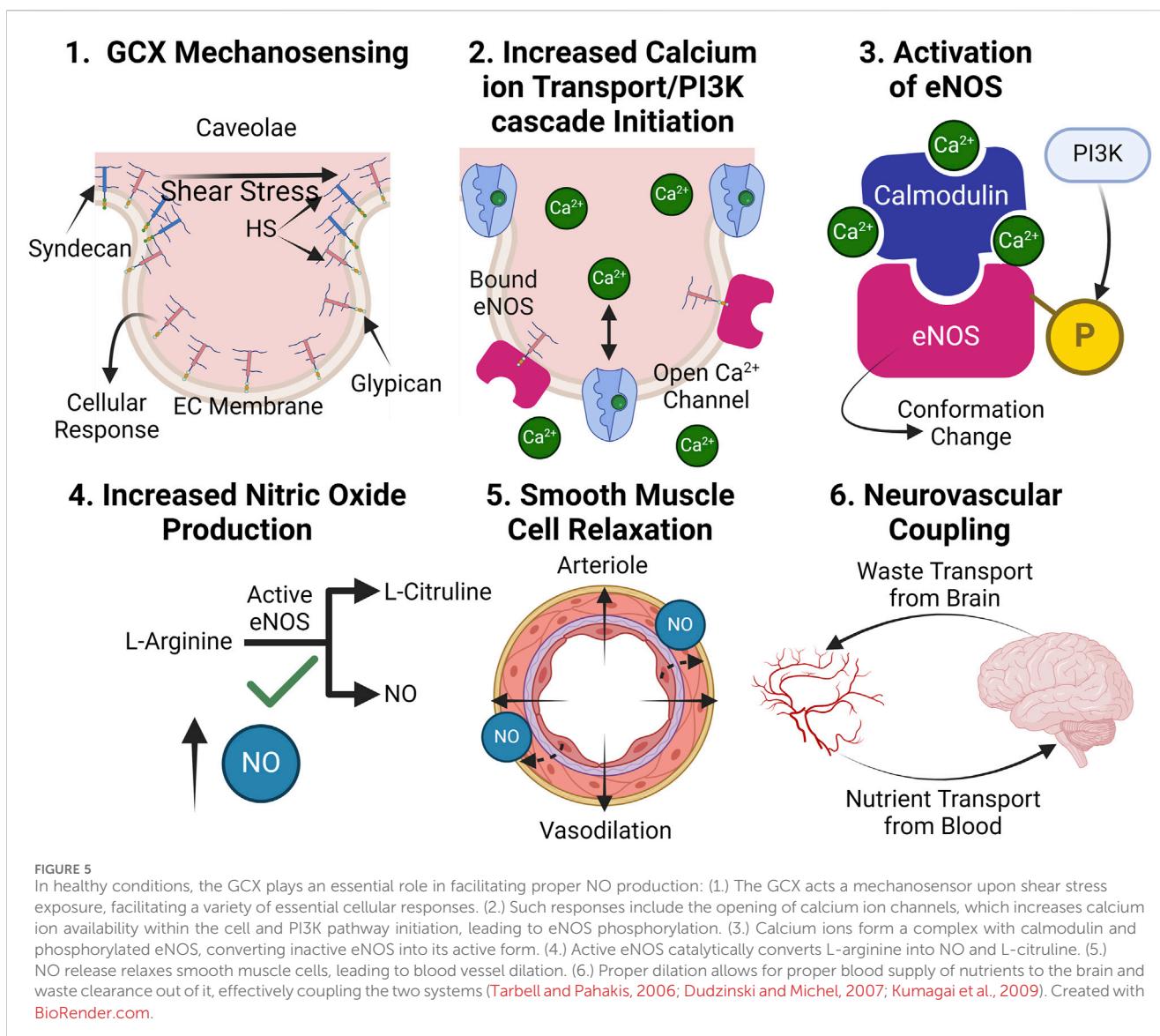
Due to the GCX's involvement in actuating NO production, studies which degrade HS have demonstrated decreased phosphorylated eNOS and subsequent reduced NO production (Ebong et al., 2014; Yen et al., 2015). Subsequent research has utilized atomic force microscopy pulling of key proteoglycans and GAGs of the GCX to identify HS and glypican-1 as being involved in the production of NO, but not syndecan-1, CD44, and HA (Bartosch et al., 2017; Bartosch et al., 2021). These works highlight the GCX's involvement in the pathway as well as the diverse functions of different components in the structure.

Further implicating previously mentioned inflammatory pathologies with NO production, the relationship between ROS, NO, and the GCX also mediates vascular dysfunction. During oxidative stress, ROS like O_2^- interact with NO, forming more precarious species like reactive nitrogen species (RNS) and peroxynitrite (Wang et al., 2018; Schenck et al., 2021). Simultaneously, NO bioavailability is reduced, and the GCX is ameliorated by superoxide dismutase (Rubio-Gayosso et al., 2006). The combination of oxidative stress, reduction in NO, and GCX deterioration creates a vicious cycle of vascular dysfunction, ultimately leading to neurovascular decoupling and the progression of AD symptoms. Future work is needed to clarify mechanisms related to GCX mechanosensing, NO production, and ultimate control of neurovascular coupling in the context of AD. However, with the structure's definitive involvement in the NO pathway, targeted regeneration of the GCX stands as a potential approach for alleviating decoupling-related AD pathology.

3 The endothelial glycocalyx: a link to Alzheimer's disease related vascular pathologies?

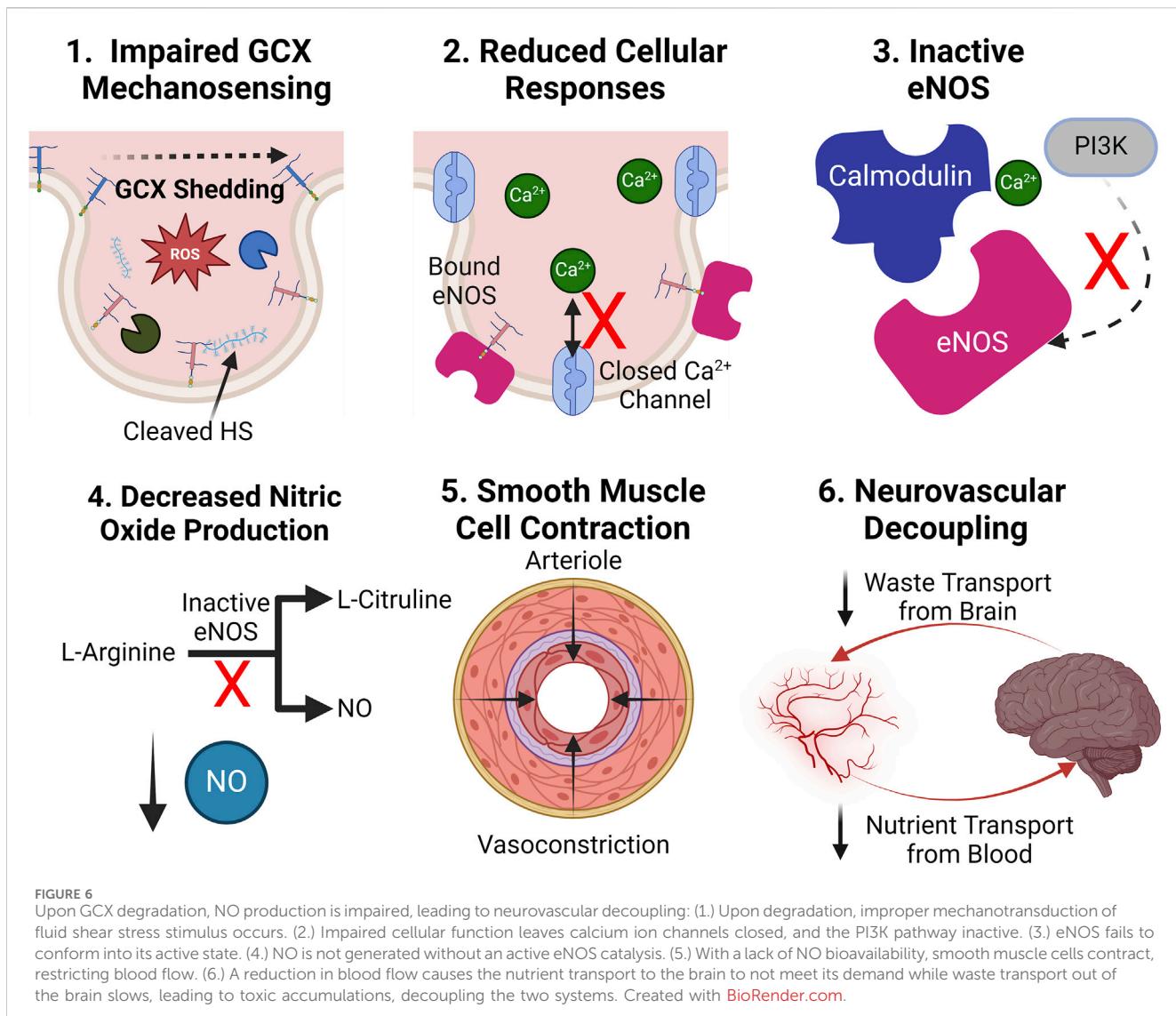
3.1 Evidence of endothelial glycocalyx degradation in Alzheimer's disease and other neurodegenerative diseases

Despite an expanding body of work ascertaining the GCX's role in regulating various vascular functions in AD, the connection between GCX shedding and the progression of AD remains inadequately explored. To establish an argument for the GCX's role in AD, foundational evidence of GCX deterioration must be demonstrated in these conditions. Currently, only a handful of studies directly examine the GCX in AD progression and other neurological disorders. Smyth et al. may have presented the most clinically-relevant evidence for the correlation between GCX shedding and AD (Figure 1). They found reduced vascular density within AD brain tissue compared to healthy controls but normalized this reduction in blood vessels to GCX abundance, further demonstrating that GCX was reduced in brain tissue regardless of the number of vessels. Their findings also elucidated a role of the GCX in deterring neutrophil attachment (Smyth et al., 2022). Others have identified a mechanism of circulating HA and HS proteoglycans in AD neuropathology as well (van Horssen et al., 2001; Reed et al., 2019; Lorente-Gea et al., 2020). Increased levels of heparinase (cleaver of HS) were found in AD brain tissue, providing another conceivable association between GCX shedding and AD progression (Garcia et al., 2017). Loss of HS likely rearranges the BBB, leading to impaired perivascular clearance and subsequent accumulation of $\text{A}\beta$ (Zhang et al., 2021). These published findings provide an indication of GCX involvement in AD, although there is a lack of direct structural measurements of the GCX in AD brain microvasculature as well as mechanistic studies relating GCX loss to AD pathology.



To supplement the few studies that have examined the relationship between GCX function and AD, a growing body of work has demonstrated GCX breakdown in other neurodegenerative disorders with similar vascular pathologies. Therefore, examining the GCX's role in catalyzing a variety of neurodegenerative diseases beyond just AD, could prove widely beneficial. For example, GCX loss has been demonstrated in subcortical vascular dementia, a disease with notably similar vascular components to AD. In an extensive study examining the GCX's role in neurovascular disease, Yoon et al. reported thinning of the GCX along with increased capillary stalling in a vascular dementia animal model (Yoon et al., 2022). Additionally, after enzymatic degradation of the GCX, capillary stalling increased mainly due to leukocyte plugging and an upregulation of endothelial adhesion receptors, suggesting a causal relationship between GCX and vascular dementia pathology (Yoon et al., 2022). Researchers have also investigated the use of GCX components in plasma as an early

biomarker in other neurodegenerative diseases. In a mouse model of multiple sclerosis, HS levels increased before symptom onset, chondroitin sulfate increased prior to symptoms and remained elevated throughout, and HA levels spiked during peak disease severity (DellaValle et al., 2018). Elevated oxidative stress decreased NO levels, and increased glycoprotein deposits in cerebral tissue have also widely been attributed to GCX shedding in neurodegenerative diseases (Pacher et al., 2007; Bonneh-Barkay and Wiley, 2009; Siebert et al., 2014). Although an acute event, ischemic stroke events exhibit similar GCX-related neurovascular pathologies. Ischemic stroke victims display loss of the GCX corresponding with increased neuronal and vascular inflammation (Ko et al., 2020). The initial blockage of blood flow and subsequent reperfusion initiates a rise of ROS followed by matrix metalloproteinases (MMPs) and other proteolytic enzymes which then shed the GCX (Yu et al., 2019). There is substantial evidence of increased levels of shed GCX



components like HS and syndecan in the plasma of stroke victims (Rehm et al., 2007; DellaValle et al., 2019). The deteriorated extracellular GCX structure allows for direct interaction between immune cells and the endothelium, eventually exacerbating neurological impairment (Mockl, 2020; Bettcher et al., 2021a; Bettcher et al., 2021b; Liu et al., 2023). Another acute event that occurs upstream of neurodegenerative diseases, traumatic brain injury, is also known to induce a GCX-related neuropathological cascade (Gonzalez Rodriguez et al., 2018; Zou et al., 2021; Anand et al., 2023). Increased shedding of GCX components after traumatic brain injury is associated with a negative survival rate, implicating the GCX's role in pathology. After injury, an upregulation of ROS, immune cell adhesion, and sheddases results in BBB hyperpermeability, neuroinflammation, and leukocyte attachment (Zou et al., 2021).

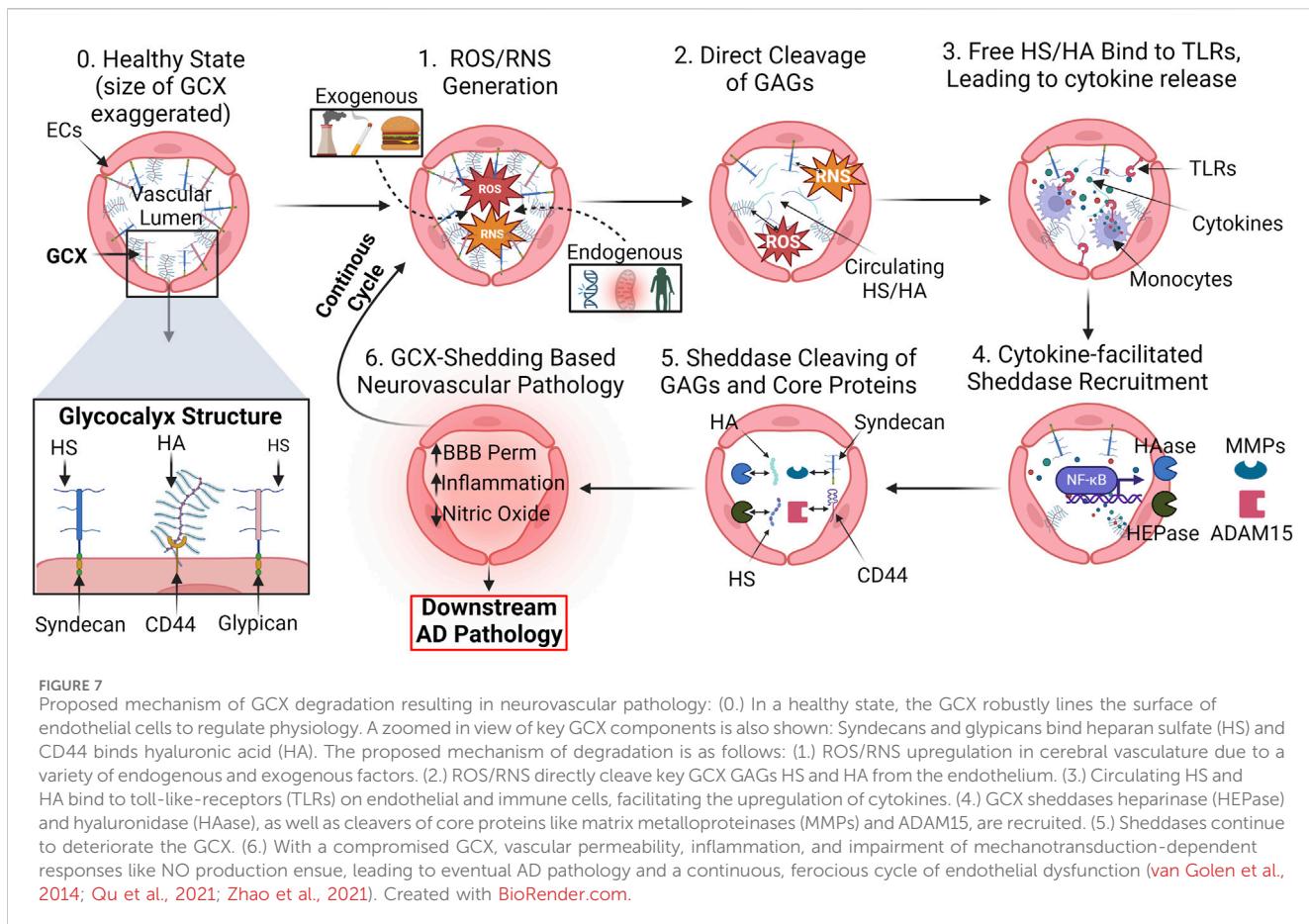
Lastly, the process of aging is worthy of mention due to age being the number one risk factor for the majority of neurodegenerative diseases (Hou et al., 2019) and known to be associated with a diminished GCX (Machin et al., 2018; Majerczak et al., 2019).

Machin et al. proposed the hypothesis that age-related GCX shedding precedes vascular dysfunction, ultimately promoting a variety of cardiovascular and neurological disease pathologies (Machin et al., 2019).

In summary, it is still emerging whether GCX shedding precedes AD and other neurological disorders characterized by vascular abnormalities. The early evidence described here indicates a correlation between GCX integrity and disease severity.

3.2 Proposed mechanism of endothelial glycocalyx shedding in Alzheimer's disease

The means by which the GCX is shed during AD pathology remains a topic of intense investigation. In physiology, GCX homeostasis is meticulously maintained through a dynamic interplay of synthesis, protection, and shear stress sensing mechanisms. Endothelial cells continually synthesize and assemble GCX components, while physiological levels of shear



stress from blood flow reinforce its structure and stimulate GAG synthesis pathways (Alphonsus and Rodseth, 2014; Banerjee et al., 2021). Endogenous factors like NO, prostacyclin, and angiopoietin-1 interact with the GCX to regulate vascular tone, permeability, and stability (Becker et al., 2010; Potje et al., 2020; Qiu et al., 2022). Additionally, antioxidants such as superoxide dismutase help counteract oxidative stress-induced GCX damage, ensuring its integrity and proper function (Beresewicz et al., 1998). However, in pathology, numerous mediators, often interrelated, facilitate GCX degradation, directing a persistent cycle of endothelial dysfunction. These mediators include, but are not limited to ROS/RNS, inflammatory cytokines, sheddases, and MMPs (Masola et al., 2021; Zhao et al., 2021). Thus, understanding the mechanisms underlying the GCX dysregulation cycle is crucial for developing targeted therapeutic interventions to preserve vascular health and potentially prevent downstream neurological dysfunction (Figure 7).

ROS and RNS play a role in direct cleavage of GAGs, as well as the recruitment of other harmful compounds. Three principal reactive species that directly sever HA and HS GAG chains from core protein binding sites include hydroxyl radicals ($\bullet\text{OH}$), carbonate radical anions ($\text{CO}_3\bullet-$), and hypochlorous acid (HOCl) (van Golen et al., 2014). Generation of these reactive species stems from multiple sources but one of the most pertinent endogenous routes involves their secretion by immune cells and the destabilization of the synthase that produces counteractive NO (Alderton et al., 2001; Forman and Torres, 2002). Other sources of ROS/RNS production may stem from the weakening

of free radical neutralizing systems due to aging, as well as environmental factors such as a high-fat diet, air pollution, and smoking (Pérez et al., 2009; Di Meo et al., 2016). ROS/RNS production sets off a cascade of vascular pathologies, among which cleavage of the GCX is a notable factor.

Upon initial cleavage of the HA and HS GCX chains, a cyclic inflammatory cascade promotes further GCX shedding (Figure 7). Circulating HS and low molecular weight HA bind to toll-like receptors on the surface of macrophages and endothelial cells, facilitating the release of inflammatory cytokines such as tumor necrosis factor- α (TNF- α), interleukin-1 β (IL-1 β), and IL-8 (Johnson et al., 2002; Taylor et al., 2004; Scheibner et al., 2006; Qu et al., 2021). In conjunction with ROS/RNS, cytokines recruit a variety of sheddases to the endothelial cell surface including MMPs, hyaluronidase, and heparinase (Dempsey et al., 2000; Monzon et al., 2010; Ramnath et al., 2014). These enzymes not only have the capacity to further shed GAGs but also threaten GAG-anchoring core proteins in the transmembrane domain of the cell. For example, MMP1, MMP9, and MMP14 remove the HS proteoglycan core proteins syndecan-1 and syndecan-4, respectively, while MMP9 also cleaves chondroitin sulfate (Endo et al., 2003; Sieve et al., 2018). A disintegrin and metalloproteinase 15 (ADAM15) is responsible for the scission of CD44, the core protein responsible for HA binding to the endothelial cell surface (Yang et al., 2018). Additionally, heparinases and hyaluronidases continuously remove HS and HA, further complicating GCX recovery attempts by the cell.

Understanding how the GCX degradation cascade is initiated and translating the knowledge to the development of potential avenues for reversal of GCX degradation could prove critical to the amelioration of a plethora of vascular-related cognitive diseases, including AD. Several mediators of GCX dysfunction are found at elevated levels in patients suffering from AD. Excessive generation of GCX cleavers ROS and RNS has been linked to the pathogenesis of AD, as well as variety of age-related neurodegenerative diseases (Manoharan et al., 2016). Notably, elevated levels of ROS production are found to precede late-stage AD events like A β deposits and NFTs, further supporting the notion that vascular dysfunction propagates late-state AD events (Ahmad et al., 2017). Furthermore, postmortem AD brain tissues show an increase in cytokines IL-1 β and IL-6 (Griffin et al., 1989; Hull et al., 1996; Zheng et al., 2016), which suggests both the presence of excess circulating GCX components as well as increased immune cell infiltration and systemic inflammation processes occurring throughout disease progression. MMP-9 was also found at significantly elevated levels in the plasma of AD patients compared to controls (Lorenzl et al., 2003). Although the abovementioned mediators of GCX shedding serve diverse functions, establishing their involvement in the pathophysiology of AD supports the hypothesis of GCX-dependent contributions to the diseases.

4 Current challenges

4.1 Imaging and quantifying cerebrovascular glycocalyx and cerebral vascular changes

To investigate the potential role of GCX integrity in neurodegenerative disease progression, it is crucial to improve both *in vitro* and *in vivo* imaging techniques of the cerebrovascular GCX. Currently, the quantification of GCX components such as HS, HA, and syndecan-1 in plasma is the primary method of assessing GCX shedding. However, additional, more direct approaches should be considered to evaluate the cerebrovascular GCX structure in preclinical, clinical, and postmortem AD tissue.

Transmission electron microscopy (TEM) is a principal technique for imaging the GCX in fixed, preserved conditions. However, one of the main challenges in TEM is preserving the sample in a way that accurately reflects the GCX ultrastructure (Haeren et al., 2016). Traditional preparation processes typically involve dehydration of the sample, causing the GCX to collapse in on itself and drastically altering its organization and thickness (Hempel et al., 2020). To address this issue, rapid freezing/freeze substitution (RF/FS) has emerged as a promising technique for sample preparation. Ebong et al. demonstrated that using RF/FS, the GCX of bovine aortic endothelial cells measured an average of 11 μ m, compared to an insubstantial 0.040 μ m using traditional TEM (Eborg et al., 2011). Despite the promise of RF/FS in accurately preserving the GCX, it remains a labor-intensive, delicate process that is unable to image specific GCX components.

To address the limitations of traditional TEM, confocal laser scanning microscopy (CLSM) is a promising method that can fluorescently label specific GCX components. While the characterization of GCX GAGs remains a challenge due to their

high degree of heterogeneity and lack of antigenicity, innovative antibodies and lectins (such as wheat germ agglutinin) can be utilized to tag domains of GAG components of the GCX, for fluorescent labeling and subsequent CLSM visualization (Mockl, 2020). By combining RF/FS with immunolabeling-based CLSM, Twamley et al. were able to analyze the morphology of specific GCX components (Twamley et al., 2021), advancing the state-of-the-art in labeling and quantification of the GCX.

Despite the recent advances of *in vitro* techniques, development of unique solutions to microscopic imaging of the cerebrovascular GCX *in vivo* in functional tissue remains essential for elucidating the nature and function of the structure in the body. Two-photon light scanning microscopy (TPLSM) utilizes dual-photon absorption to increase penetration depth and allow for fluorescent imaging of living tissue (Denk et al., 1990). By contrasting fluorescent dextran in the lumen with wheat germ agglutinin-tagged GCX, TPLSM has been used to assess GCX thickness, BBB permeability, and vascular geometry (Yoon et al., 2017; Kucharz et al., 2022). Results indicate that the use of *in vivo* TPLSM leads to a more robust GCX than what is observed *ex vivo* and offers a functional approach to assessing GCX thickness, BBB permeability, and vascular geometry. However, this imaging technique will likely be limited to pre-clinical animal studies for cerebrovascular GCX imaging due to its invasiveness, high cost, slow imaging speed, and the need for fluorescent contrast agents.

Another option for *in vivo* assessment of the GCX that demonstrates more clinical potential is side-stream dark field imaging (SDF). SDF measures the degree of penetration of red blood cells into the GCX through a reflected LED light. Based on the extent of penetration, computer programs like Glycocheck can correlate the perfused boundary region (the permeable part of the GCX) to a GCX thickness (Iba and Levy, 2019). A recent protocol developed by Haeren et al. was the first to utilize SDF of the cerebrovascular GCX in a clinical setting. Based on the study, SDF could effectively quantify GCX thickness in the cerebral vasculature, was straightforward to operate, and safe for the use on human patients (Haeren et al., 2018). SDF and Glycocheck offer the advantage of intraoperative measurement of GCX thickness without the need for fluorescent dyes, long processing times, and image acquisition expertise (Haeren et al., 2017; Berhouma et al., 2020). However, SDF and Glycocheck still pose several drawbacks due to indirect GCX measurement and may be affected by motion-induced blurring. With the applicability of SDF and Glycocheck, an increasing amount of data on cerebrovascular GCX integrity in neurodegenerative disease is expected to emerge in the coming years, offering valuable insights for tailored pre-clinical and clinical applications.

Finally, due to the substantial difficulties of directly quantifying the GCX *in vivo*, novel, translational imaging modalities which can broadly quantify vascular changes will be instrumental in elucidating the causal role, if any, of GCX impairment on AD progression during longitudinal studies. Some studies have utilized dynamic contrast-enhanced magnetic resonance imaging (DCE MRI) to investigate BBB leakage in early AD (Figure 1) (van de Haar et al., 2016; Moon et al., 2023). However, DCE MRI has limitations in studying BBB permeability primarily due to its inability to distinguish between blood flow and permeability changes accurately (Manning et al., 2021). To overcome this, a

TABLE 1 Choosing the Right BBB Model: Key Considerations and Comparative Analysis: This table offers a thorough evaluation of recent BBB models, examining each model against five pivotal factors essential for achieving physiological relevance as well as experimental feasibility. “Ease of fabrication” assesses the simplicity and technical expertise required for model construction. Consideration is given to the length of cell culture required for each model, impacting experimental throughput as well. “Physiological Flow” evaluates the model’s ability to simulate hemodynamic flow conditions experienced by cerebral blood vessels. “Physiological geometry” examines how closely the model’s structure resembles the innate architecture of the neurovasculature. “Post-experimental Analysis” assesses the ease and effectiveness of conducting post-experimental biological assays including barrier functionality, protein quantification, and fluorescent microscopy. “Model Versatility” is determined by the model’s capability to adapt to different mechanical environments, culture conditions, and experimental factors, enabling diverse applications. Based on this assessment, a millifluidic, transwell-compatible model shows promise for future *in vitro* studies exploring the role of the GCX in AD.

Model type	Graphical representation	Ease of fabrication	Physiological flow	Physiological geometry	Post-experimental analysis	Versatility	Ref
Millifluidic Transwell (Ebong Model)		✓	✓		✓	✓	Harding et al. (2022)
Transwell		✓			✓		Ito et al. (2019), Li et al. (2019)
Organoid		?		✓	✓		Cho et al. (2017b), Berhouma et al. (2020)
Organ-on-a-Chip			✓		✓	✓	Brown et al. (2015), Boghdeh et al. (2022)
3D Vessel			✓	✓	?	?	Marino et al. (2021), Yan et al. (2023)

promising approach is to utilize quantitative ultra-short time-to-echo contrast-enhanced magnetic resonance imaging (QUTE-CE MRI) to holistically quantify BBB permeability changes over time more accurately. QUTE-CE MRI offers a safe, accurate, and non-invasive method of quantifying BBB leakage making it an ideal *in vivo* quantitative method for longitudinal studies (Gharagouzloo et al., 2015; Leaston et al., 2021a; Leaston et al., 2021b). Furthermore, QUTE-CE MRI’s precise quantification of BBB changes facilitates targeted investigation into key regions affected in AD—like the hippocampus, thalamus, amygdala, and cerebral cortex, while still encompassing the entire brain in the study (Kapasi and Schneider, 2016; Nelson et al., 2016). Compared with traditional MRI, QUTE-CE MRI allows for more precise measurement of BBB permeability due to its ability to capture ultra-short echo times, which are sensitive to subtle changes in tissue microstructure and permeability. This enables researchers and clinicians to obtain more accurate and reliable data on BBB integrity, facilitating the possibility of prolonged longitudinal studies on a key read out for

neurovascular health. With the continued development of these revolutionary imaging methods, unprecedented insight into GCX integrity, dynamics of BBB permeability, and overall cerebral vascular health are positioned to be elucidated, heralding a transformative era in vascular and AD research.

4.2 *In Vitro* modeling of the BBB

Another substantial challenge with elucidating the GCX’s role in AD involves the development of suitable *in vitro* models of the BBB. Access to preclinical and clinical *in vivo* settings that incorporate the complex nature of the BBB is limited. Therefore, the development of accurate yet high throughput *in vitro* models of the BBB structure remain indispensable to the field for their reproducibility, cost-effectiveness, and accessibility. *In vitro* BBB models which can accurately reproduce the interplay between the GCX, barrier function, and pathophysiology are critical to examining the

GCX-vascular hypothesis of AD. Despite significant research in this area, recent models vary widely in their fabrication methods, flow conditions, model geometry, applicability, and versatility (Table 1). Developing an *in vitro* system which balances these parameters to fit the experimental goals continues to be a priority.

Today, the field largely agrees that the primitive *in vitro* BBB model consisting of a static, brain microvascular endothelial cell monolayer cultured in a transwell insert falls short of the needed complexity to be considered a viable candidate to study GCX-BBB-AD interactions (Table 1). A primary shortcoming of this model is the lack of physiological shear stress the cells are subjected to. Hemodynamic flow conditions are increasingly recognized as essential for capturing the intricate microenvironment of the NVU *in vitro*, as endothelial cells respond dynamically to shear stress (Williams-Medina et al., 2020). Under shear stress, brain endothelial cell phenotype alterations occur, with flow models demonstrating decreased BBB permeability compared to static cultures (Santaguida et al., 2006; Cucullo et al., 2011). In specific relation to the GCX, thickness of the structure in flow conditions has been reported to increase 70%–80% compared to static conditions and approach thicknesses observed *in vivo* (Ueda et al., 2004; Tsvirkun et al., 2017; Harding et al., 2019b). Selecting a model which generates a robust GCX is essential for future studies aimed at investigating the GCX role in deterring AD pathology.

Similar to the importance of flow conditions, *in vitro* BBB models which can incorporate many cell types to recreate the complex multicellular interface are essential to accurately studying AD pathogenesis. Different cells, including primary human endothelial cells, induced pluripotent stem cell-derived (iPSC) endothelial cells, animal-derived endothelial cells, and immortalized endothelial cells, mimic the cellular lining of the brain vascular lumen (He et al., 2014; Destefano et al., 2018). Adding another layer of complexity, supportive cells such as astrocytes, pericytes, microglia, and neurons are also often incorporated and deemed essential to achieving a functional BBB model, depending on the purpose (Daneman and Prat, 2015). These diverse cell types enable the study of neurovascular function in healthy conditions, neurovascular function in neuroinflammatory and neurodegenerative conditions, therapeutic targets, and drug transport across the BBB, but it is still unclear the exact importance of them to a functional model.

In terms of fabricating a model which captures the innate geometric properties of the BBB, several innovative fabrication methods are being investigated (Table 1). Certain microfluidic systems utilize channel culturing techniques to replicate 3D blood vessels and allow for physiological flow and cell-cell interactions, providing increased physiological relevance (Bonakdar et al., 2017; Hajal et al., 2022). However, these systems can be technically complex to fabricate and operate, requiring specialized expertise and equipment. Scaffold-based approaches create three-dimensional frameworks using porous membranes or hydrogels, promoting cell-cell interactions and barrier formation but may lack the ability to precisely control spatial organization and have limited scalability (Marino et al., 2021). Novel 3D organoid models enable *in vivo* like formations of complex BBB structures (Cho CF. et al., 2017; Bergmann et al., 2018; van Dyck et al., 2023). However, organoid culturing techniques are still under development, and challenges remain in

achieving reproducibility, scalability, and mechanically accurate microenvironments. The preferred option for fabrication method often depends on research expertise, experimental design, and throughput considerations.

With the goal of taking all of these factors into consideration, our lab has developed a custom-made millifluidic device that is compatible with standard transwell inserts to develop a physiologically relevant study while maintaining high throughput for analysis of the role of the GCX in AD (Table 1) (Harding et al., 2022). This design combines standard transwell inserts with a microfluidic device featuring validated shear stress profiles. It also incorporates primary human brain endothelial cells, pericytes, and astrocytes, enhancing fidelity for studying BBB dynamics and neurological consequences of GCX impairment. The advantages of this model include its straightforward and reusable fabrication via 3D printing and insertion of transwell inserts, physiological flow rate, robust post-experimental analysis, and versatility with the ability to culture different cell types and flow rate changes. Although there are more physiological fabrication approaches which achieve 3D geometries and direct cell-cell contact, this transwell system is comparable to previous studies (Table 1). Initial studies demonstrate that GCX thickness and tight junction expression increase in tri-cultures containing endothelial cells, astrocytes, and pericytes. Furthermore, exposure to 12 dyne/cm² of shear stress increased GCX coverage, potentially supporting the lower observed permeability in flow conditions (Harding et al., 2022). This model is well suited for balancing the needed physiological relevance with ease of use required to study the GCX's role in AD.

4.3 Experimental methods of studying GCX-related Alzheimer's disease pathology

Due to the GCX's heterogeneity and complex functionality to the NVU, devising experimental strategies to study the relationship between GCX degradation and vascular-related AD pathology is a critical consideration. Traditionally, the GCX is degraded through enzymatic cleavage of its GAGs (McDonald et al., 2016; Yalcin et al., 2018). Although this technique may be sufficient for other vascular-organ interfaces, the multicellular, dynamic nature of the BBB may lead to the enzymes having several off-target effects on brain cell function, resulting in questions of causation during downstream biological analysis (Yang et al., 2021; Zhang et al., 2021).

For this reason, recent gene editing advancements offer practical means to investigate the GCX's role in AD-vascular pathology (Reitsma et al., 2007). Techniques like CRISPR/Cas9, shRNA, and siRNA effectively knockdown or knockout GCX core protein expression, facilitating the study of their functional significance (Song and Yang, 2010; Abrahimi et al., 2015). By employing these techniques *in vitro* and *in vivo*, researchers can investigate the functional consequences of depleting GCX core protein expression on cellular processes i.e., barrier integrity, inflammation control, and responsiveness to the extracellular environment. While effective *in vitro*, utilizing *in vivo* knockout models which broadly target GCX core proteins presents challenges due to their expression in cell types beyond

endothelial cells, increasing the risk of off-target effects (Couchman and Pataki, 2012). To address this issue, efforts should be directed towards generating conditional transgenic animal models with endothelial cell-specific GCX core protein knockouts (Liao et al., 2001; Fahs et al., 2014). These genomic editing approaches would contribute to a deeper understanding of the roles played by GCX core proteins in disease processes, including neurodegenerative disorders like AD.

The impact of GCX-specific knockout mice will be powerful when combined with the *in vivo* imaging techniques described in Section 4.1. For example, *in vivo* imaging techniques such as TPLSM offer the potential to track the neurovasculature and GCX over time with greater depth and precision compared to traditional techniques (Haeren et al., 2016; Kilic et al., 2020). Additionally, the use of QUTE-CE MRI provides a promising method to track neurovascular abnormalities, BBB permeability, cerebral blood flow, and neuroinflammation over the course of disease progression in a non-invasive, clinically-relevant way (Leaston et al., 2021a). Finally, post-mortem-tissue analysis utilizing RF/FS preservation techniques can visualize GCX thickness and coverage in a manner previously unobtainable with traditional preservatives. Given the relatively recent appreciation for GCX involvement in neurodegenerative disease and the development of these groundbreaking imaging modalities, the foundation is set to firmly establish a prospective relationship between GCX degradation and downstream AD pathology.

A legitimate concern with evaluating the direct relationship between GCX impairment and AD progression *in vitro* and *in vivo* is modelling complex disease in a relevant way. To generate AD-like pathology both *in vitro* and *in vivo*, researchers may employ various techniques to mimic key aspects of the disease. *In vitro*, AD-like pathology can be induced in cultured cells or blood-brain organoids by exposing them to A β peptides, tau protein aggregates, or neuroinflammatory cytokines (Sharma et al., 2021; Schreiner et al., 2022). This can lead to the formation of A β plaques, tau hyperphosphorylation, synaptic dysfunction, and neuronal cell death, resembling key features of AD pathology (Chen et al., 2021). *In vivo*, there are several commercially available transgenic rodent models of AD but are a constant subject of debate, primarily because of the disease's complex etiology and the fact that rodents cannot naturally develop it (Yokoyama et al., 2022). Some commonly deployed models for studying AD which are well-characterized for inducing neuropathological features and cognitive deficits characteristic of the disease include the mutant amyloid precursor protein and presenilin 1 (APP/PS1), Tg2576, and 5xFAD lines (Gotz et al., 2018; Sanchez-Varo et al., 2022). A major advantage of these AD models is abnormal protein folding in the brain, appreciable vascular changes, and cognitive deficits witnessed and progressing between 6 and 12 months of age in most models, setting a feasible time scale for longitudinal studies of AD (Webster et al., 2014). However, one of the persistent shortcomings of these transgenic models is their neuro-centric focus and failure to exhibit likely vascular changes present in AD. However, inducing impairment of the GCX in these models and subsequently longitudinally examining the animals for variations in cognitive function would be valuable in establishing the GCX's role as a contributor to the propagation of AD. This would be

particularly significant if AD models with GCX deficiency were to exhibit exacerbated symptoms.

Moreover, beyond studying AD models, exploring the role of the GCX in AD also involves considering the aging aspect, the number one risk factor of the disease. Aged cells and rodent models provide valuable insights into the dynamic changes occurring within the GCX as the brain matures, shedding light on its relevance to AD pathology. Aged cells exhibit physiological changes and molecular alterations that mimic aspects of aging-related neurodegeneration observed in AD while also paralleling the believed thinning of the GCX with age (Ting et al., 2021; Sun et al., 2023). *In vitro*, cell lines derived from aged individuals can be used to model age-related changes in cellular function and susceptibility to AD pathology. These cells may exhibit increased oxidative stress, protein misfolding, mitochondrial dysfunction, and altered signaling pathways associated with aging and neurodegeneration (Chen et al., 2020). Likewise, investigating aged rodents *in vivo* provides insight into the effects of aging on disease progression and its potential correlation with GCX loss. Such research, considering traditional AD amyloid-based pathology, age, and GCX alterations, not only explores the potential impact of GCX loss as a major contributing factor but also potentially informs the development of novel therapeutic interventions for AD.

To address whether GCX loss exacerbates AD pathology, specific techniques can be employed. *In vitro*, GCX loss can be induced in endothelial cells as described above, followed by exposure to AD-related stimuli. *In vivo*, genetic manipulation or pharmacological approaches targeting GCX components can be combined with AD-inducing interventions in transgenic mouse models. By assessing changes in BBB integrity, A β clearance, neuroinflammation, and cognitive function, researchers can elucidate the role of GCX loss in worsening AD pathology and identify potential therapeutic targets for intervention.

5 Future work and conclusion

Neurodegenerative diseases such as AD are among the most urgent public health concerns of the 21st century, with the demand for effective therapies expected to increase alongside the aging population. Given the complex etiology of AD involving both neurological and vascular components, it is essential to advance research on the understanding of the interplay of both systems. Targeting the vascular endothelial GCX emerges as a promising avenue, offering the potential to prolong or restore critical neurovascular functions impaired in AD. Despite the ongoing uncertainty regarding whether GCX degradation precedes vascular insult and AD pathology, recent advancements in evaluating the GCX in the cerebral vasculature open doors for future work to definitively determine its viability as a therapeutic target for neurodegenerative diseases.

After conducting an extensive literature review and considering experimental feasibility, the initial approach to determining whether the GCX could serve as a viable therapeutic target for delaying, mitigating, or preventing AD symptoms involves thoroughly assessing the functionality of the GCX in relation to countering

AD-related neurovascular pathologies in both *in vitro* and *in vivo* settings.

Tangible methods of accomplishing this task *in vitro* include: 1. Utilize a physiologically relevant, reliable, high degree of post-experimental analysis BBB model such as the one described by our group in a previous publication (Harding et al., 2022). 2. Use a highly transfectable shRNA knockdown technique to reduce the expression of individual GCX core proteins selectively and stably such as CD44, syndecan-1, and glypican-1 in brain microvascular endothelial cells. 3. Assess the effects of these GCX core protein knockdowns on NVU functionality using techniques such as dextran permeability assays, trans-endothelial electric resistance, and cell-cell communication. Additionally, analyzing the protein and gene expression of markers associated with AD-related neurovascular pathology to uncover potential mechanistic insights, including tight junctions, intracellular transporters, adhesion molecules, cytokines, and NO production mediators. This approach aims to elucidate the role of individual GCX components in maintaining physiological function.

In vivo methods of examining the role of the GCX in AD should aim to corroborate *in vitro* findings while further capturing complexities unobtainable through solely cell-based observations such as: 1. Examine distinctions among healthy mice, CD44 knockout models (Due to this knockout model's commercial availability and minimal off-target effects), and amyloid-based AD models like APP/PS1, to understand the ramifications of GCX impairment in contrast to the traditional neurocentric manifestation of AD, particularly concerning AD-related neurovascular dysfunction (Protin et al., 1999; McKallip et al., 2002). 2. Longitudinally interrogate changes in BBB permeability using translationally relevant QUTE-CE MRI until reaching an appropriate advanced age, to consider the compounded effect of aging in relation to GCX impairment and AD progression. 3. Perform immunological analysis on brain sections and plasma samples, comparing baseline samples to those obtained after aging in the three groups, and probing for similar markers as in the *in vitro* study. These findings will explore whether the knockout of a crucial GCX core protein, CD44, triggers BBB dysfunction, while also distinguishing the effects of BBB damage stemming from abnormal protein folding linked to AD.

If the data from an initial study were to show promise, further investigations could be pursued on determining the GCX's relevance in AD, involving more labor, cost, and time-intensive approaches as those highlighted in Section 4.3. If evidence were to mount indicating the significant involvement of the GCX in AD pathology, it becomes imperative to explore novel GCX-regenerating therapeutics in these models, assessing their potential to ameliorate the severe pathological symptoms of the disease (Banerjee et al., 2021). While significant efforts are still required to ascertain the role, if any, of this vascular-based structure in protecting against AD, recent advancements in biotechnology have undoubtedly opened the door to feasibly address this question like never before. As we continue to navigate the complexities of AD and the potential of a substantial vascular role, the pursuit of a deeper understanding of the GCX's relationship to the disease stands as a critical step toward effective treatments and improved public health outcomes.

Author contributions

NO'H: Conceptualization, Funding acquisition, Writing–review and editing, Data curation, Formal Analysis, Investigation, Visualization, Writing–original draft. KM: Investigation, Visualization, Writing–original draft, Writing–review and editing, Data curation, Formal Analysis. EEE: Conceptualization, Investigation, Writing–review and editing, Data curation, Funding acquisition, Project administration, Resources, Supervision.

Funding

The author(s) declare that financial support was received for the research, authorship, and/or publication of this article. This work was also funded by the National Science Foundation Graduate Research Fellowship, DGE-1938052 (to NO'H), to support PhD dissertation research focused on studying the role of the mechanically regulated endothelial glycocalyx in blood-brain barrier function and Alzheimer's Disease. We acknowledge that this work was also funded by the National Science Foundation CAREER Award, CMMI-1846962 (to EEE), to support endothelial mechanobiology research and education.

Acknowledgments

We would like to thank faculty in the chemical engineering, bioengineering, and psychology department at Northeastern University for providing invaluable input on certain aspects of this review including Dr. Abbas Yaseen, Dr. Craig Ferris, Dr. Ryan Koppes, and Dr. Guohao Dai. We acknowledge support from Kate Kryder, who is a Data Analysis & Visualization Specialist in the Northeastern University Library. Additionally, we would like to thank current and past members of the Ebong lab group for contributing with the conceptualization, focus, and editing of this review including Ian Harding, Mark Vigliotti, Ronodeep Mitra, Mohammad Hamrangsekachae, Chinedu Okorafor, and Kaleigh Pentland.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

References

Abrahimi, P., Chang, W. G., Kluger, M. S., Qyang, Y., Tellides, G., Saltzman, W. M., et al. (2015). Efficient gene disruption in cultured primary human endothelial cells by CRISPR/Cas9. *Circ. Res.* 117 (2), 121–128. doi:10.1161/CIRCRESAHA.117.306290

Ahmad, W., Ijaz, B., Shabbiri, K., Ahmed, F., and Rehman, S. (2017). Oxidative toxicity in diabetes and Alzheimer's disease: mechanisms behind ROS/RNS generation. *J. Biomed. Sci.* 24 (1), 76. doi:10.1186/s12929-017-0379-z

Al-Ahmad, A. J., Patel, R., Palecek, S. P., and Shusta, E. V. (2019). Hyaluronan impairs the barrier integrity of brain microvascular endothelial cells through a CD44-dependent pathway. *J. Cereb. Blood Flow. Metab.* 39 (9), 1759–1775. doi:10.1177/0271678X18767748

Alderton, W. K., Cooper, C. E., and Knowles, R. G. (2001). Nitric oxide synthases: structure, function and inhibition. *Biochem. J.* 357 (Pt 3), 593–615. doi:10.1042/0264-6021:3570593

Alphonsus, C. S., and Rodseth, R. N. (2014). The endothelial glycocalyx: a review of the vascular barrier. *Anaesthesia* 69 (7), 777–784. doi:10.1111/anae.12661

Alzheimer's disease facts and figures (2021). Alzheimer's disease facts and figures. *Alzheimer's Dementia* 17 (3), 327–406. doi:10.1002/azld.12328

Anand, T., Reyes, A. A., Sjöquist, M. C., Magnotti, L., and Joseph, B. (2023). Resuscitating the endothelial glycocalyx in trauma and hemorrhagic shock. *Ann. Surg. Open* 4 (3), e298. doi:10.1097/AS9.0000000000000298

Banerjee, S., Mwangi, J. G., Stanley, T. K., Mitra, R., and Ebong, E. E. (2021). Regeneration and assessment of the endothelial glycocalyx to address cardiovascular disease. *Industrial Eng. Chem. Res.* 60 (48), 17328–17347. doi:10.1021/acs.iecr.1c03074

Bartosch, A. M. W., Mathews, R., Mahmoud, M. M., Cancel, L. M., Haq, Z. S., and Tarbell, J. M. (2021). Heparan sulfate proteoglycan glycan-1 and PECAM-1 cooperate in shear-induced endothelial nitric oxide production. *Sci. Rep.* 11 (1), 11386. doi:10.1038/s41598-021-90941-w

Bartosch, A. M. W., Mathews, R., and Tarbell, J. M. (2017). Endothelial glycocalyx-mediated nitric oxide production in response to selective AFM pulling. *Biophys. J.* 113 (1), 101–108. doi:10.1016/j.bpj.2017.05.033

Becker, B. F., Chappell, D., Bruegger, D., Annecke, T., and Jacob, M. (2010). Therapeutic strategies targeting the endothelial glycocalyx: acute deficits, but great potential. *Cardiovasc. Res.* 87 (2), 300–310. doi:10.1093/cvr/cvq137

Beresewicz, A., Czarnowska, E., and Maczewski, M. (1998). Ischemic preconditioning and superoxide dismutase protect against endothelial dysfunction and endothelium glycocalyx disruption in the postischemic Guinea-pig hearts. *Mol. Cell. Biochem.* 186 (1–2), 87–97. doi:10.1007/978-1-4615-4979-6_11

Bergmann, S., Lawler, S. E., Qu, Y., Fadzen, C. M., Wolfe, J. M., Regan, M. S., et al. (2018). Blood-brain-barrier organoids for investigating the permeability of CNS therapeutics. *Nat. Protoc.* 13 (12), 2827–2843. doi:10.1038/s41596-018-0066-x

Berhouma, M., Picart, T., Dumot, C., Pelissou-Guyotat, I., Meyronet, D., Ducray, F., et al. (2020). Alterations of cerebral microcirculation in peritumoral edema: feasibility of *in vivo* sidestream dark-field imaging in intracranial meningiomas. *Neuro-Oncology Adv.* 2 (1), vdaa108. doi:10.1093/noajnl/vdaa108

Bettcher, B. M., Tansey, M. G., Dorothee, G., and Heneka, M. T. (2021a). Peripheral and central immune system crosstalk in Alzheimer disease - a research prospectus. *Nat. Rev. Neurol.* 17 (11), 689–701. doi:10.1038/s41582-021-00549-x

Bettcher, B. M., Tansey, M. G., Dorothee, G., and Heneka, M. T. (2021b). Publisher Correction: peripheral and central immune system crosstalk in Alzheimer disease - a research prospectus. *Nat. Rev. Neurol.* 17 (11), 724. doi:10.1038/s41582-021-00579-5

Betteridge, K. B., Arkill, K. P., Neal, C. R., Harper, S. J., Foster, R. R., Satchell, S. C., et al. (2017). Sialic acids regulate microvessel permeability, revealed by novel *in vivo* studies of endothelial glycocalyx structure and function. *J. Physiol.* 595 (15), 5015–5035. doi:10.1111/jp274167

Boghdeh, N. A., Risner, K. H., Barrera, M. D., Britt, C. M., Schaffer, D. K., Alem, F., et al. (2022). Application of a human blood brain barrier organ-on-a-chip model to evaluate small molecule effectiveness against Venezuelan equine encephalitis virus. *Viruses* 14 (12), 2799. doi:10.3390/v14122799

Bonakdar, M., Graybill, P. M., and Davalos, R. V. (2017). A microfluidic model of the blood-brain barrier to study permeabilization by pulsed electric fields. *RSC Adv.* 7 (68), 42811–42818. doi:10.1039/C7RA07603G

Bonneh-Barkay, D., and Wiley, C. A. (2009). Brain extracellular matrix in neurodegeneration. *Brain Pathol.* 19 (4), 573–585. doi:10.1111/j.1750-3639.2008.00195.x

Braak, H., and Braak, E. (1995). Staging of Alzheimer's disease-related neurofibrillary changes. *Neurobiol. Aging* 16 (3), 271–278. ; discussion 8–84. doi:10.1016/0197-4580(95)00021-6

Brown, J. A., Pensabene, V., Markov, D. A., Allwardt, V., Neely, M. D., Shi, M., et al. (2015). Recreating blood-brain barrier physiology and structure on chip: a novel neurovascular microfluidic bioreactor. *Biomicrofluidics* 9 (5), 054124. doi:10.1063/1.4934713

Chen, M. B., Yang, A. C., Yousef, H., Lee, D., Chen, W., Schaum, N., et al. (2020). Brain endothelial cells are exquisite sensors of age-related circulatory cues. *Cell. Rep.* 30 (13), 4418–4432. doi:10.1016/j.celrep.2020.03.012

Chen, X., Sun, G., Tian, E., Zhang, M., Davtyan, H., Beach, T. G., et al. (2021). Modeling sporadic Alzheimer's disease in human brain organoids under serum exposure. *Adv. Sci.* 8 (18), 2101462. doi:10.1002/advs.202101462

Chen, Z. L., and Strickland, S. (1997). Neuronal death in the hippocampus is promoted by plasmin-catalyzed degradation of laminin. *Cell.* 91 (7), 917–925. doi:10.1016/s0092-8674(00)80483-3

Chiodelli, P., Bugatti, A., Urbinati, C., and Rusnati, M. (2015). Heparin/heparan sulfate proteoglycans glycanome in angiogenesis: biological implications and therapeutic use. *Molecules* 20 (4), 6342–6388. doi:10.3390/molecules20046342

Cho, C. F., Wolfe, J. M., Fadzen, C. M., Calligaris, D., Hornburg, K., Chiocca, E. A., et al. (2017a). Blood-brain-barrier spheroids as an *in vitro* screening platform for brain-penetrating agents. *Nat. Commun.* 8, 15623. doi:10.1038/ncomms15623

Cho, C.-F., Wolfe, J. M., Fadzen, C. M., Calligaris, D., Hornburg, K., Chiocca, E. A., et al. (2017b). Blood-brain-barrier spheroids as an *in vitro* screening platform for brain-penetrating agents. *Nat. Commun.* 8 (1), 15623. doi:10.1038/ncomms15623

Constantinescu, A. A., Vink, H., and Spaan, J. A. E. (2003). Endothelial cell glycocalyx modulates immobilization of leukocytes at the endothelial surface. *Arteriosclerosis, Thrombosis, Vasc. Biol.* 23 (9), 1541–1547. doi:10.1161/01.ATV.0000085630.24353.3D

Couchman, J. R., and Pataki, C. A. (2012). An introduction to proteoglycans and their localization. *J. Histochem. Cytochem.* 60 (12), 885–897. doi:10.1369/0022155412464638

Cucullo, L., Hossain, M., Puvenna, V., Marchi, N., and Janigro, D. (2011). The role of shear stress in Blood-Brain Barrier endothelial physiology. *BMC Neurosci.* 12, 40. doi:10.1186/1471-2202-12-40

Curry, F. E., and Adamson, R. H. (2012). Endothelial glycocalyx: permeability barrier and mechanosensor. *Ann. Biomed. Eng.* 40 (4), 828–839. doi:10.1007/s10439-011-0429-8

Daneman, R., and Prat, A. (2015). The blood-brain barrier. *Cold Spring Harb. Perspect. Biol.* 7 (1), a020412. doi:10.1101/cshperspect.a020412

De La Torre, J. C. (2002). Alzheimer disease as a vascular disorder: nosological evidence. *Stroke* 33 (4), 1152–1162. doi:10.1161/01.str.0000014421.15948.67

de la Torre, J. C., and Mussivand, T. (1993). Can disturbed brain microcirculation cause Alzheimer's disease? *Neurol. Res.* 15 (3), 146–153. doi:10.1080/01616412.1993.11740127

DellaValle, B., Hasselblad, H., Johansen, F. F., Iversen, H. K., Rungby, J., and Hempel, C. (2019). Multiple soluble components of the glycocalyx are increased in patient plasma after ischemic stroke. *Stroke* 50 (10), 2948–2951. doi:10.1161/STROKEAHA.119.025953

DellaValle, B., Manresa-Arraut, A., Hasselblad, H., Stensballe, A., Rungby, J., Larsen, A., et al. (2018). Detection of glycan shedding in the blood: new class of multiple sclerosis biomarkers? *Front. Immunol.* 9, 1254. doi:10.3389/fimmu.2018.01254

Dempsey, L. A., Brunn, G. J., and Platt, J. L. (2000). Heparanase, a potential regulator of cell-matrix interactions. *Trends Biochem. Sci.* 25 (8), 349–351. doi:10.1016/s0968-0004(00)01619-4

Denk, W., Strickler, J. H., and Webb, W. W. (1990). Two-photon laser scanning fluorescence microscopy. *Science* 248 (4951), 73–76. doi:10.1126/science.2321027

Deore, B. J., Partyka, P. P., Fan, F., and Galie, P. A. (2022). CD44 mediates shear stress mechanotransduction in an *in vitro* blood-brain barrier model through small GTPases RhoA and Rac1. *FASEB J.* 36 (5), e22278. doi:10.1096/fj.202100822RR

De-Paula, V. J., Radanovic, M., Diniz, B. S., and Forlenza, O. V. (2012). *Alzheimer's disease*. Netherlands: Springer, 329–352.

Destefano, J. G., Jamieson, J. J., Linville, R. M., and Seaton, P. C. (2018). Benchmarking *in vitro* tissue-engineered blood-brain barrier models. *Fluids Barriers CNS* 15 (1), 32. doi:10.1186/s12987-018-0117-2

Di Marco, L. Y., Venneri, A., Farkas, E., Evans, P. C., Marzo, A., and Frangi, A. F. (2015). Vascular dysfunction in the pathogenesis of Alzheimer's disease—A review of endothelium-mediated mechanisms and ensuing vicious circles. *Neurobiol. Dis.* 82, 593–606. doi:10.1016/j.nbd.2015.08.014

Di Meo, S., Reed, T. T., Venditti, P., and Victor, V. M. (2016). Role of ROS and RNS sources in physiological and pathological conditions. *Oxid. Med. Cell. Longev.* 2016, 1245049. doi:10.1155/2016/1245049

Ding, Y., Zhong, Y., Baldesswiler, A., Abner, E. L., Bauer, B., and Hartz, A. M. S. (2021). Protecting P-glycoprotein at the blood-brain barrier from degradation in an Alzheimer's disease mouse model. *Fluids Barriers CNS* 18 (1), 10. doi:10.1186/s12987-021-00245-4

Dogné, S., and Flamion, B. (2020). Endothelial glycocalyx impairment in disease: focus on hyaluronan shedding. *Am. J. Pathol.* 190 (4), 768–780. doi:10.1016/j.ajpath.2019.11.016

Dormanns, K., Brown, R. G., and David, T. (2016). The role of nitric oxide in neurovascular coupling. *J. Theor. Biol.* 394, 1–17. doi:10.1016/j.jtbi.2016.01.009

Dudzinski, D. M., and Michel, T. (2007). Life history of eNOS: partners and pathways. *Cardiovasc. Res.* 75 (2), 247–260. doi:10.1016/j.cardiores.2007.03.023

Ebong, E. E., Lopez-Quintero, S. V., Rizzo, V., Spray, D. C., and Tarbell, J. M. (2014). Shear-induced endothelial NOS activation and remodeling via heparan sulfate, glypcan-1, and syndecan-1. *Integr. Biol. (Camb.)* 6 (3), 338–347. doi:10.1039/c3ib40199e

Ebong, E. E., Macaluso, F. P., Spray, D. C., and Tarbell, J. M. (2011). Imaging the endothelial glycocalyx *in vitro* by rapid freezing/freeze substitution transmission electron microscopy. *Arterioscler. Thromb. Vasc. Biol.* 31 (8), 1908–1915. doi:10.1161/ATVBAHA.111.225268

Endo, K., Takino, T., Miyamori, H., Kinsen, H., Yoshizaki, T., Furukawa, M., et al. (2003). Cleavage of syndecan-1 by membrane type matrix metalloproteinase-1 stimulates cell migration. *J. Biol. Chem.* 278 (42), 40764–40770. doi:10.1074/jbc.M306736200

Fahs, S. A., Hille, M. T., Shi, Q., Weiler, H., and Montgomery, R. R. (2014). A conditional knockout mouse model reveals endothelial cells as the principal and possibly exclusive source of plasma factor VIII. *Blood* 123 (24), 3706–3713. doi:10.1182/blood-2014-02-555151

Floris, S., Van Den Born, J., Van Der Pol, S. M. A., Dijkstra, C. D., and De Vries, H. E. (2003). Heparan sulfate proteoglycans modulate monocyte migration across cerebral endothelium. *J. Neuropathology Exp. Neurology* 62 (7), 780–790. doi:10.1093/jnen/62.7.780

Forman, H. J., and Torres, M. (2002). Reactive oxygen species and cell signaling: respiratory burst in macrophage signaling. *Am. J. Respir. Crit. Care Med.* 166 (12 Pt 2), S4–S8. doi:10.1164/rccm.2206007

Garcia, B., Martin, C., Garcia-Suarez, O., Muniz-Alonso, B., Ordiales, H., Fernandez-Menendez, S., et al. (2017). Upregulated expression of heparanase and heparanase 2 in the brains of Alzheimer's disease. *J. Alzheimers Dis.* 58 (1), 185–192. doi:10.3233/JAD-160198

Gharagouzloo, C. A., McMahon, P. N., and Sridhar, S. (2015). Quantitative contrast-enhanced MRI with superparamagnetic nanoparticles using ultrashort time-to-echo pulse sequences. *Magnetic Reson. Med.* 74 (2), 431–441. doi:10.1002/mrm.25426

Goes, A., Wouters, D., Pol, S. M. A., Huijzinga, R., Ronken, E., Adamson, P., et al. (2001). Reactive oxygen species enhance the migration of monocytes across the blood-brain barrier *in vitro*. *FASEB J.* 15 (10), 1852–1854. doi:10.1096/fj.00-0881fje

Gomez Gonzalez, B., Larios, H. M., and Escobar, A. (2011). Increased transvascular transport of WGA-peroxidase after chronic perinatal stress in the hippocampal microvasculature of the rat. *Int. J. Dev. Neurosci.* 29 (8), 839–846. doi:10.1016/j.ijdevneu.2011.08.003

Gonzalez Rodriguez, E., Cardenas, J. C., Cox, C. S., Kitagawa, R. S., Stensballe, J., Holcomb, J. B., et al. (2018). Traumatic brain injury is associated with increased syndecan-1 shedding in severely injured patients. *Scand. J. trauma, Resusc. Emerg. Med.* 26 (1), 102. doi:10.1186/s13049-018-0565-3

Gotz, J., Bodea, L. G., and Goedert, M. (2018). Rodent models for Alzheimer disease. *Nat. Rev. Neurosci.* 19 (10), 583–598. doi:10.1038/s41583-018-0054-8

Govindpani, K., McNamara, L. G., Smith, N. R., Vinnakota, C., Waldvogel, H. J., Faull, R. L., et al. (2019). Vascular dysfunction in Alzheimer's disease: a prelude to the pathological process or a consequence of it? *J. Clin. Med.* 8 (5), 651. doi:10.3390/jcm8050651

Griffin, W. S., Stanley, L. C., Ling, C., White, L., MacLeod, V., Perrot, L. J., et al. (1989). Brain interleukin 1 and S-100 immunoreactivity are elevated in Down syndrome and Alzheimer disease. *Proc. Natl. Acad. Sci. U. S. A.* 86 (19), 7611–7615. doi:10.1073/pnas.86.19.7611

Haeren, R. H., van de Ven, S. E., van Zandvoort, M. A., Vink, H., van Overbeeke, J. J., Hoogland, G., et al. (2016). Assessment and imaging of the cerebrovascular glycocalyx. *Curr. Neurovasc. Res.* 13 (3), 249–260. doi:10.2174/156720261366160504104434

Haeren, R. H., Vink, H., Staals, J., van Zandvoort, M. A., Dings, J., van Overbeeke, J. J., et al. (2017). Protocol for intraoperative assessment of the human cerebrovascular glycocalyx. *BMJ Open* 7 (1), e013954. doi:10.1136/bmjjopen-2016-013954

Haeren, R. H. L., Rijkers, K., Schijns, O., Dings, J., Hoogland, G., van Zandvoort, M., et al. (2018). *In vivo* assessment of the human cerebral microcirculation and its glycocalyx: a technical report. *J. Neurosci. Methods* 303, 114–125. doi:10.1016/j.jneumeth.2018.03.009

Hajal, C., Offeddu, G. S., Shin, Y., Zhang, S., Morozova, O., Hickman, D., et al. (2022). Engineered human blood-brain barrier microfluidic model for vascular permeability analyses. *Nat. Protoc.* 17 (1), 95–128. doi:10.1038/s41596-021-00635-w

Harding, I. C., Cisneros, W., and Ebong, E. (2019a). The role of the endothelial glycocalyx in atherosclerosis development via ROS production and downstream endothelial activation. *Proc. Conf. Arteriosclerosis, Thrombosis, Vasc. Biol.* doi:10.1161/atvb.39.suppl_1.733

Harding, I. C., Mitra, R., Mensah, S. A., Nersesyan, A., Bal, N. N., and Ebong, E. E. (2019b). Endothelial barrier reinforcement relies on flow-regulated glycocalyx, a potential therapeutic target. *Biorheology* 56 (2-3), 131–149. doi:10.3233/BIR-180205

Harding, I. C., O'Hare, N. R., Vigliotti, M., Caraballo, A., Lee, C. I., Millican, K., et al. (2022). Developing a transwell millifluidic device for studying blood-brain barrier endothelium. *Lab a Chip* 22 (23), 4603–4620. doi:10.1039/d2lc00657j

He, Y., Yao, Y., Tsirka, S. E., and Cao, Y. (2014). Cell-culture models of the blood-brain barrier. *Stroke* 45 (8), 2514–2526. doi:10.1161/STROKEAHA.114.005427

Hempel, C., Kapishnikov, S., Perez-Berna, A. J., Werner, S., Guttmann, P., Pereiro, E., et al. (2020). The need to freeze—dehydration during specimen preparation for electron microscopy collapses the endothelial glycocalyx regardless of fixation method. *Microcirculation* 27 (7), e12643. doi:10.1111/micc.12643

Hock, C., Villringer, K., Muller-Spahn, F., Wenzel, R., Heeker, H., Schuh-Hofer, S., et al. (1997). Decrease in parietal cerebral hemoglobin oxygenation during performance of a verbal fluency task in patients with Alzheimer's disease monitored by means of near-infrared spectroscopy (NIRS)—correlation with simultaneous rCBF-PET measurements. *Brain Res.* 755 (2), 293–303. doi:10.1016/s0006-8993(97)00122-4

Hou, Y., Dan, X., Babbar, M., Wei, Y., Hasselbalch, S. G., Croteau, D. L., et al. (2019). Ageing as a risk factor for neurodegenerative disease. *Nat. Rev. Neurol.* 15 (10), 565–581. doi:10.1038/s41582-019-0244-7

Hull, M., Berger, M., Volk, B., and Bauer, J. (1996). Occurrence of interleukin-6 in cortical plaques of Alzheimer's disease patients may precede transformation of diffuse into neuritic plaques. *Ann. N. Y. Acad. Sci.* 777, 205–212. doi:10.1111/j.1749-6632.1996.tb34420.x

Hussain, B., Fang, C., and Chang, J. (2021). Blood-brain barrier breakdown: an emerging biomarker of cognitive impairment in normal aging and dementia. *Front. Neurosci.* 15, 688090. doi:10.3389/fnins.2021.688090

Iba, T., and Levy, J. H. (2019). Derangement of the endothelial glycocalyx in sepsis. *J. Thromb. Haemost.* 17 (2), 283–294. doi:10.1111/jth.14371

Ito, R., Umehara, K., Suzuki, S., Kitamura, K., Nunoya, K. I., Yamaura, Y., et al. (2019). A human immortalized cell-based blood-brain barrier triculture model: development and characterization as a promising tool for drug-brain permeability studies. *Mol. Pharm.* 16 (11), 4461–4471. doi:10.1021/acs.molpharmaceut.9b00519

Jin, J., Fang, F., Gao, W., Chen, H., Wen, J., Wen, X., et al. (2021). The structure and function of the glycocalyx and its connection with blood-brain barrier. *Front. Cell. Neurosci.* 15, 739699. doi:10.3389/fncel.2021.739699

Johnson, G. B., Brunn, G. J., Kodaira, Y., and Platt, J. L. (2002). Receptor-mediated monitoring of tissue well-being via detection of soluble heparan sulfate by Toll-like receptor 4. *J. Immunol.* 168 (10), 5233–5239. doi:10.4049/jimmunol.168.10.5233

Kadry, H., Noorani, B., and Cucullo, L. (2020). A blood-brain barrier overview on structure, function, impairment, and biomarkers of integrity. *Fluids Barriers CNS* 17 (1), 69. doi:10.1186/s12987-020-00230-3

Kapasi, A., and Schneider, J. A. (2016). Vascular contributions to cognitive impairment, clinical Alzheimer's disease, and dementia in older persons. *Biochim. Biophys. Acta* 1862 (5), 878–886. doi:10.1016/j.bbadi.2015.12.023

Kilic, K., Desjardins, M., Tang, J., Thunemann, M., Sunil, S., Erdener, S. E., et al. (2020). Chronic cranial windows for long term multimodal neurovascular imaging in mice. *Front. Physiol.* 11, 612678. doi:10.3389/fphys.2020.612678

Ko, K., Suzuki, T., Ishikawa, R., Hattori, N., Ito, R., Umehara, K., et al. (2020). Ischemic stroke disrupts the endothelial glycocalyx through activation of pro-HPSE via acrolein exposure. *J. Biol. Chem.* 295 (52), 18614–18624. doi:10.1074/jbc.RA120.015105

Korczyn, A. D. (2008). The amyloid cascade hypothesis. *Alzheimer's Dementia.* 4 (3), 176–178. doi:10.1016/j.jalz.2007.11.008

Korczyn, A. D. (2012). Why have we failed to cure Alzheimer's disease? *J. Alzheimers Dis.* 29 (2), 275–282. doi:10.3233/JAD-2011-110359

Kucharcz, K., Mathiesen Janiurek, M., Christoffersen, C., and Lauritzen, M. (2022). Two-photon microscopy *in vivo* reveals brain vessel type-specific loss of glycocalyx caused by apoM/SIP signaling impairment.

Kumagai, R., Lu, X., and Kassab, G. S. (2009). Role of glycocalyx in flow-induced production of nitric oxide and reactive oxygen species. *Free Radic. Biol. Med.* 47 (5), 600–607. doi:10.1016/j.freeradbiomed.2009.05.034

Kumar, A., Sidhu, J., Goyal, A., and Tsao, J. W. (2022). *Alzheimer disease. StatPearls.* USA: Treasure Island (FL).

Kutuzov, N., Flyvbjerg, H., and Lauritzen, M. (2018). Contributions of the glycocalyx, endothelium, and extravascular compartment to the blood-brain barrier. *Proc. Natl. Acad. Sci. U. S. A.* 115 (40), E9429–E9438–E38. doi:10.1073/pnas.1802155115

Leaston, J., Ferris, C. F., Kulkarni, P., Chandramohan, D., Van De Ven, A. L., Qiao, J., et al. (2021a). Neurovascular imaging with QUTE-CE MRI in APOE rats reveals early vascular abnormalities. *PLOS ONE* 16 (8), e0256749. doi:10.1371/journal.pone.0256749

Leaston, J., Qiao, J., Harding, I. C., Kulkarni, P., Gharagouzloo, C., Ebong, E., et al. (2021b). Quantitative imaging of blood-brain barrier permeability following repetitive mild head impacts. *Front. Neurology* 12, 729464. doi:10.3389/fneur.2021.729464

Leite, D. M., Matias, D., and Battaglia, G. (2020). The role of BAR proteins and the glycocalyx in brain endothelium transcytosis. *Cells* 9 (12), 2685. doi:10.3390/cells9122685

Li, N. C., Lee, A., Whitmer, R. A., Kivipelto, M., Lawler, E., Kazis, L. E., et al. (2010). Use of angiotensin receptor blockers and risk of dementia in a predominantly male population: prospective cohort analysis. *BMJ* 340, b5465. doi:10.1136/bmj.b5465

Li, Y., Sun, X., Liu, H., Huang, L., Meng, G., Ding, Y., et al. (2019). Development of human *in vitro* brain-blood barrier model from induced pluripotent stem cell-derived endothelial cells to predict the *in vivo* permeability of drugs. *Neurosci. Bull.* 35 (6), 996–1010. doi:10.1007/s12264-019-00384-7

Liao, Y., Day, K. H., Damon, D. N., and Duling, B. R. (2001). Endothelial cell-specific knockout of connexin 43 causes hypotension and bradycardia in mice. *Proc. Natl. Acad. Sci.* 98 (17), 9989–9994. doi:10.1073/pnas.171305298

Lipowsky, H. H. (2012). The endothelial glycocalyx as a barrier to leukocyte adhesion and its mediation by extracellular proteases. *Ann. Biomed. Eng.* 40 (4), 840–848. doi:10.1007/s10439-011-0427-x

Liu, C., Xu, S., Liu, Q., Chai, H., Luo, Y., and Li, S. (2023). Identification of immune cells infiltrating in hippocampus and key genes associated with Alzheimer's disease. *BMC Med. Genomics* 16 (1), 53. doi:10.1186/s12920-023-01458-2

Lorente-Gea, L., Garcia, B., Martin, C., Ordiales, H., Garcia-Suarez, O., Pina-Batista, K. M., et al. (2020). Heparan sulfate proteoglycans undergo differential expression alterations in alzheimer disease brains. *J. Neuropathol. Exp. Neurol.* 79 (5), 474–483. doi:10.1093/jnen/nlaa016

Lorenzl, S., Albers, D. S., Relkin, N., Ngyuen, T., Hilgenberg, S. L., Chirichigno, J., et al. (2003). Increased plasma levels of matrix metalloproteinase-9 in patients with Alzheimer's disease. *Neurochem. Int.* 43 (3), 191–196. doi:10.1016/s0197-0186(03)00004-4

Luft, J. H. (1966). Fine structures of capillary and endocapillary layer as revealed by ruthenium red. *Fed. Proc.* 25 (6), 1773–1783.

Machin, D. R., Bloom, S. I., Campbell, R. A., Phuong, T. T. T., Gates, P. E., Lesniewski, L. A., et al. (2018). Advanced age results in a diminished endothelial glycocalyx. *Am. J. Physiol. Heart Circ. Physiol.* 315 (3), H531–H539–H9. doi:10.1152/ajpheart.00104.2018

Machin, D. R., Phuong, T. T., and Donato, A. J. (2019). The role of the endothelial glycocalyx in advanced age and cardiovascular disease. *Curr. Opin. Pharmacol.* 45, 66–71. doi:10.1016/j.coph.2019.04.011

Majerczak, J., Grandys, M., Frolow, M., Szkutnik, Z., Zakrzewska, A., Nizankowski, R., et al. (2019). Age-dependent impairment in endothelial function and arterial stiffness in former high class male athletes is No different to that in men with No history of physical training. *J. Am. Heart Assoc.* 8 (18), e012670. doi:10.1161/JAHA.119.012670

Manning, C., Stringer, M., Dickie, B., Clancy, U., Valdés Hernandez, M. C., Wiseman, S. J., et al. (2021). Sources of systematic error in DCE-MRI estimation of low-level blood-brain barrier leakage. *Magnetic Reson. Med.* 86 (4), 1888–1903. doi:10.1002/mrm.28833

Manoharan, S., Guillemin, G. J., Abiramasundari, R. S., Essa, M. M., Akbar, M., and Akbar, M. D. (2016). The role of reactive oxygen species in the pathogenesis of Alzheimer's disease, Parkinson's disease, and huntington's disease: a mini review. *Oxid. Med. Cell. Longev.* 2016, 8590578. doi:10.1155/2016/8590578

Marino, A., Baronio, M., Buratti, U., Mele, E., and Ciofani, G. (2021). Porous optically transparent cellulose acetate scaffolds for biomimetic blood-brain barrier *in vitro* models. *Front. Bioeng. Biotechnol.* 9, 630063. doi:10.3389/fbioe.2021.630063

Masola, V., Zaza, G., Arduini, A., Onisto, M., and Gambaro, G. (2021). Endothelial glycocalyx as a regulator of fibrotic processes. *Int. J. Mol. Sci.* 22 (6), 2996. doi:10.3390/ijms22062996

McDonald, K. K., Cooper, S., Danielzak, L., and Leask, R. L. (2016). Glycocalyx degradation induces a proinflammatory phenotype and increased leukocyte adhesion in cultured endothelial cells under flow. *PLoS ONE* 11 (12), e0167576. doi:10.1371/journal.pone.0167576

McKallip, R. J., Do, Y., Fisher, M. T., Robertson, J. L., Nagarkatti, P. S., and Nagarkatti, M. (2002). Role of CD44 in activation-induced cell death: CD44-deficient mice exhibit enhanced T cell response to conventional and superantigens. *Int. Immunol.* 14 (9), 1015–1026. doi:10.1093/intimm/dxf068

Mhatre, M., Nguyen, A., Kashani, S., Pham, T., Adesina, A., and Grammas, P. (2004). Thrombin, a mediator of neurotoxicity and memory impairment. *Neurobiol. Aging* 25 (6), 783–793. doi:10.1016/j.neurobiolaging.2003.07.007

Mockl, L. (2020). The emerging role of the mammalian glycocalyx in functional membrane organization and immune system regulation. *Front. Cell. Dev. Biol.* 8, 253. doi:10.3389/fcell.2020.00253

Montagne, A., Barnes, S. R., Sweeney, M. D., Halliday, M. R., Sagare, A. P., Zhao, Z., et al. (2015). Blood-brain barrier breakdown in the aging human hippocampus. *Neuron* 85 (2), 296–302. doi:10.1016/j.neuron.2014.12.032

Monzon, M. E., Fregien, N., Schmid, N., Falcon, N. S., Campos, M., Casalino-Matsuda, S. M., et al. (2010). Reactive oxygen species and hyaluronidase 2 regulate airway epithelial hyaluronan fragmentation. *J. Biol. Chem.* 285 (34), 26126–26134. doi:10.1074/jbc.M110.135194

Moon, Y., Jeon, H. J., Han, S. H., Min-Young, N., Kim, H. J., Kwon, K. J., et al. (2023). Blood-brain barrier breakdown is linked to tau pathology and neuronal injury in a differential manner according to amyloid deposition. *J. Cereb. Blood Flow. Metab.* 43 (11), 1813–1825. doi:10.1177/0271678X231180035

Moore, K. H., Murphy, H. A., and George, E. M. (2021). The glycocalyx: a central regulator of vascular function. *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 320 (4), R508–R518. doi:10.1152/ajpregu.00340.2020

Nelson, A. R., Sweeney, M. D., Sagare, A. P., and Zlokovic, B. V. (2016). Neurovascular dysfunction and neurodegeneration in dementia and Alzheimer's disease. *Biochim. Biophys. Acta* 1862 (5), 887–900. doi:10.1016/j.bbadi.2015.12.016

O'Hare, N. R., Harding, I. C., Herman, I. M., and Ebong, E. E. (2022). "The roles of shear stress and heparan sulfate degradation in endothelial cell activation," in Proceeding of the Summer Biomechanics, Bioengineering, & Biotransport Conference, China, June 28, 2023 (IEEE).

Pacher, P., Beckman, J. S., and Liaudet, L. (2007). Nitric oxide and peroxynitrite in health and disease. *Physiol. Rev.* 87 (1), 315–424. doi:10.1152/physrev.00029.2006

Pahakis, M. Y., Kosky, J. R., Dull, R. O., and Tarbell, J. M. (2007). The role of endothelial glycocalyx components in mechanotransduction of fluid shear stress. *Biochem. Biophysical Res. Commun.* 355 (1), 228–233. doi:10.1016/j.bbrc.2007.01.137

Park, L., Anrather, J., Zhou, P., Frys, K., Pitstick, R., Younkin, S., et al. (2005). NADPH oxidase-derived reactive oxygen species mediate the cerebrovascular dysfunction induced by the amyloid peptide. *J. Neurosci.* 25 (7), 1769–1777. doi:10.1523/JNEUROSCI.5207-04.2005

Pérez, V. I., Bokov, A., Remmen, H. V., Mele, J., Ran, Q., Ikeno, Y., et al. (2009). Is the oxidative stress theory of aging dead? *Biochimica Biophysica Acta (BBA) - General Subj.* 1790 (10), 1005–1014. doi:10.1016/j.bbagen.2009.06.003

Potje, S. R., Paula, T. D., Paulo, M., and Bendhack, L. M. (2020). The role of glycocalyx and caveolae in vascular homeostasis and diseases. *Front. Physiol.* 11, 620840. doi:10.3389/fphys.2020.620840

Potter, D. R., Jiang, J., and Damiano, E. R. (2009). The recovery time course of the endothelial cell glycocalyx *in vivo* and its implications *in vitro*. *Circulation Res.* 104 (11), 1318–1325. doi:10.1161/CIRCRESAHA.108.191585

Prohovnik, I., Mayeux, R., Sackeim, H. A., Smith, G., Stern, Y., and Alderson, P. O. (1988). Cerebral perfusion as a diagnostic marker of early Alzheimer's disease. *Neurology* 38 (6), 931–937. doi:10.1212/wnl.38.6.931

Protin, U., Schweighoffer, T., Jochum, W., and Hilberg, F. (1999). CD44-deficient mice develop normally with changes in subpopulations and recirculation of lymphocyte subsets. *J. Immunol.* 163 (9), 4917–4923. doi:10.4049/jimmunol.163.9.4917

Qiu, Y., Buffonge, S., Ramnath, R., Jenner, S., Fawaz, S., Arkill, K. P., et al. (2022). Endothelial glycocalyx is damaged in diabetic cardiomyopathy: angiopoietin 1 restores glycocalyx and improves diastolic function in mice. *Diabetologia* 65 (5), 879–894. doi:10.1007/s00125-022-05650-4

Qu, J., Cheng, Y., Wu, W., Yuan, L., and Liu, X. (2021). Glycocalyx impairment in vascular disease: focus on inflammation. *Front. Cell. Dev. Biol.* 9, 730621. doi:10.3389/fcell.2021.730621

Ramnath, R., Foster, R. R., Qiu, Y., Cope, G., Butler, M. J., Salmon, A. H., et al. (2014). Matrix metalloproteinase 9-mediated shedding of syndecan 4 in response to tumor necrosis factor α : a contributor to endothelial cell glycocalyx dysfunction. *FASEB J.* 28 (11), 4686–4699. doi:10.1096/fj.14-252221

Reed, M. J., Damodarasamy, M., Pathan, J. L., Chan, C. K., Spiekerman, C., Wight, T. N., et al. (2019). Increased hyaluronan and TSG-6 in association with neuropathologic changes of Alzheimer's disease. *J. Alzheimers Dis.* 67 (1), 91–102. doi:10.3233/JAD-180797

Rehm, M., Bruegger, D., Christ, F., Conzen, P., Thiel, M., Jacob, M., et al. (2007). Shedding of the endothelial glycocalyx in patients undergoing major vascular surgery with global and regional ischemia. *Circulation* 116 (17), 1896–1906. doi:10.1161/CIRCULATIONAHA.106.684852

Reitsma, S., Slaaf, D. W., Vink, H., van Zandvoort, M. A., and oude Egbrink, M. G. (2007). The endothelial glycocalyx: composition, functions, and visualization. *Pflugers Arch.* 454 (3), 345–359. doi:10.1007/s00424-007-0212-8

Rochfort, K. D., Collins, L. E., Murphy, R. P., and Cummins, P. M. (2014). Downregulation of blood-brain barrier phenotype by proinflammatory cytokines involves NADPH oxidase-dependent ROS generation: consequences for interendothelial adherens and tight junctions. *PLoS ONE* 9 (7), e101815. doi:10.1371/journal.pone.0101815

Rubio-Gayoso, I., Platts, S. H., and Duling, B. R. (2006). Reactive oxygen species mediate modification of glycocalyx during ischemia-reperfusion injury. *Am. J. Physiol. Heart Circ. Physiol.* 290 (6), H2247–H2256. doi:10.1152/ajpheart.00796.2005

Ruitenberg, A., den Heijer, T., Bakker, S. L., van Swieten, J. C., Koudstaal, P. J., Hofman, A., et al. (2005). Cerebral hypoperfusion and clinical onset of dementia: the Rotterdam Study. *Ann. Neurol.* 57 (6), 789–794. doi:10.1002/ana.20493

Sanchez-Varo, R., Mejias-Ortega, M., Fernandez-Valenzuela, J. J., Nunez-Diaz, C., Caceres-Palomo, L., Vegas-Gomez, L., et al. (2022). Transgenic mouse models of Alzheimer's disease: an integrative analysis. *Int. J. Mol. Sci.* 23 (10), 5404. doi:10.3390/ijms23105404

Santaguida, S., Janigro, D., Hossain, M., Oby, E., Rapp, E., and Cucullo, L. (2006). Side by side comparison between dynamic versus static models of blood-brain barrier *in vitro*: a permeability study. *Brain Res.* 1109 (1), 1–13. doi:10.1016/j.brainres.2006.06.027

Santa-Maria, A. R., Walter, F. R., Figueiredo, R., Kincses, A., Vigh, J. P., Heymans, M., et al. (2021). Flow induces barrier and glycocalyx-related genes and negative surface charge in a lab-on-a-chip human blood-brain barrier model. *J. Cereb. Blood Flow. Metab.* 41 (9), 2201–2215. doi:10.1177/0271678X21992638

Schachtrup, C., Lu, P., Jones, L. L., Lee, J. K., Lu, J., Sachs, B. D., et al. (2007). Fibrinogen inhibits neurite outgrowth via beta 3 integrin-mediated phosphorylation of the EGF receptor. *Proc. Natl. Acad. Sci. U. S. A.* 104 (28), 11814–11819. doi:10.1073/pnas.0704045104

Scheibner, K. A., Lutz, M. A., Boodoo, S., Fenton, M. J., Powell, J. D., and Horton, M. R. (2006). Hyaluronan fragments act as an endogenous danger signal by engaging TLR2. *J. Immunol.* 177 (2), 1272–1281. doi:10.4049/jimmunol.177.2.1272

Schenck, H., Netti, E., Teernstra, O., De Ridder, I., Dings, J., Niemela, M., et al. (2021). The role of the glycocalyx in the pathophysiology of subarachnoid hemorrhage-induced delayed cerebral ischemia. *Front. Cell. Dev. Biol.* 9, 731641. doi:10.3389/fcell.2021.731641

Schreiner, T. G., Creangă-Murariu, I., Tamba, B. I., Lucanu, N., and Popescu, B. O. (2022). *In vitro* modeling of the blood-brain barrier for the study of physiological conditions and Alzheimer's disease. *Biomolecules* 12 (8), 1136. doi:10.3390/biom12081136

Sharma, N. S., Karan, A., Lee, D., Yan, Z., and Xie, J. (2021). Advances in modeling Alzheimer's disease *in vitro*. *Adv. NanoBiomed Res.* 1 (12), 2100097. doi:10.1002/anbr.20200097

Sheppard, O., and Coleman, M. (2020). *Alzheimer's disease: etiology, neuropathology and pathogenesis*. China: Exon Publications, 1–22.

Siebert, J. R., Conta Steencken, A., and Osterhout, D. J. (2014). Chondroitin sulfate proteoglycans in the nervous system: inhibitors to repair. *BioMed Res. Int.* 2014, 845323–845415. doi:10.1155/2014/845323

Sieve, I., Munster-Kuhnel, A. K., and Hilfiker-Kleiner, D. (2018). Regulation and function of endothelial glycocalyx layer in vascular diseases. *Vasc. Pharmacol.* 100, 26–33. doi:10.1016/j.vph.2017.09.002

Smyth, L. C. D., Murray, H. C., Hill, M., Van Leeuwen, E., Hight, B., Magon, N. J., et al. (2022). Neutrophil-vascular interactions drive myeloperoxidase accumulation in the brain in Alzheimer's disease. *Acta Neuropathol. Commun.* 10 (1), 38. doi:10.1186/s40478-022-01347-2

Song, H., and Yang, P. C. (2010). Construction of shRNA lentiviral vector. *N. Am. J. Med. Sci.* 2 (12), 598–601. doi:10.4297/najms.2010.2598

Soto-Rojas, L. O., Pacheco-Herrero, M., Martínez-Gómez, P. A., Campa-Córdoba, B. B., Apártiga-Pérez, R., Villegas-Rojas, M. M., et al. (2021). The neurovascular unit dysfunction in Alzheimer's disease. *Int. J. Mol. Sci.* 22 (4), 2022. doi:10.3390/ijms22042202

Stanimirovic, D. B., and Friedman, A. (2012). Pathophysiology of the neurovascular unit: disease cause or consequence? *J. Cereb. Blood Flow. Metab.* 32 (7), 1207–1221. doi:10.1038/jcbfm.2012.25

Sun, L., Wang, L., Ye, K. X., Wang, S., Zhang, R., Juan, Z., et al. (2023). Endothelial glycocalyx in aging and age-related diseases. *Aging Dis.* 14 (5), 1606–1617. doi:10.14336/AD.2023.0131

Sweeney, M. D., Sagare, A. P., and Zlokovic, B. V. (2018). Blood-brain barrier breakdown in Alzheimer disease and other neurodegenerative disorders. *Nat. Rev. Neurol.* 14 (3), 133–150. doi:10.1038/nrnuro.2017.188

Tarantini, S., Tran, C. H. T., Gordon, G. R., Ungvari, Z., and Csizsar, A. (2017). Impaired neurovascular coupling in aging and Alzheimer's disease: contribution of astrocyte dysfunction and endothelial impairment to cognitive decline. *Exp. Gerontol.* 94, 52–58. doi:10.1016/j.exger.2016.11.004

Tarbell, J. M., and Cancel, L. M. (2016). The glycocalyx and its significance in human medicine. *J. Intern. Med.* 280 (1), 97–113. doi:10.1111/joim.12465

Tarbell, J. M., and Pahakis, M. Y. (2006). Mechanotransduction and the glycocalyx. *J. Intern. Med.* 259 (4), 339–350. doi:10.1111/j.1365-2796.2006.01620.x

Taylor, K. R., Trowbridge, J. M., Rudisill, J. A., Termeer, C. C., Simon, J. C., and Gallo, R. L. (2004). Hyaluronan fragments stimulate endothelial recognition of injury through TLR4. *J. Biol. Chem.* 279 (17), 17079–17084. doi:10.1074/jbc.M310859200

Ting, K. K., Coleman, P., Zhao, Y., Vadas, M. A., and Gamble, J. R. (2021). The aging endothelium. *Vasc. Biol.* 3 (1), R35–R47. doi:10.1530/VB-20-0013

Tong, X. K., Lecrux, C., Rosa-Neto, P., and Hamel, E. (2012). Age-dependent rescue by simvastatin of Alzheimer's disease cerebrovascular and memory deficits. *J. Neurosci.* 32 (14), 4705–4715. doi:10.1523/JNEUROSCI.0169-12.2012

Tsvirkun, D., Grichine, A., Duperray, A., Misbah, C., and Bureau, L. (2017). Microvasculature on a chip: study of the endothelial surface layer and the flow structure of red blood cells. *Sci. Rep.* 7 (1), 45036. doi:10.1038/srep45036

Twamley, S. G., Stach, A., Heilmann, H., Sohl-Kielcynski, B., Stangl, V., Ludwig, A., et al. (2021). Immuno-electron and confocal laser scanning microscopy of the glycocalyx. *Biol. (Basel).* 10 (5), 402. doi:10.3390/biology10050402

Uchimido, R., Schmidt, E. P., and Shapiro, N. I. (2019). The glycocalyx: a novel diagnostic and therapeutic target in sepsis. *Crit. Care* 23 (1), 16. doi:10.1186/s13054-018-2292-6

Ueda, A., Shimomura, M., Ikeda, M., Yamaguchi, R., and Tanishita, K. (2004). Effect of glycocalyx on shear-dependent albumin uptake in endothelial cells. *Am. J. Physiol. Heart Circ. Physiol.* 287 (5), H2287–H2294. doi:10.1152/ajpheart.00808.2003

van de Haar, H. J., Burgmans, S., Jansen, J. F., van Osch, M. J., van Buchem, M. A., Muller, M., et al. (2016). Blood-brain barrier leakage in patients with early Alzheimer disease. *Radiology* 281 (2), 527–535. doi:10.1148/radiol.2016152244

van den Berg, B. M., Vink, H., and Spaan, J. A. (2003). The endothelial glycocalyx protects against myocardial edema. *Circ. Res.* 92 (6), 592–594. doi:10.1161/01.RES.0000065917.53950.75

van Dyck, C. H., Swanson, C. J., Aisen, P., Bateman, R. J., Chen, C., Gee, M., et al. (2023). Lecanemab in early Alzheimer's disease. *N. Engl. J. Med.* 388 (1), 9–21. doi:10.1056/NEJMoa2212948

van Golen, R. F., Reiniers, M. J., Vrisekoop, N., Zuurbier, C. J., Olthof, P. B., van Rheenen, J., et al. (2014). The mechanisms and physiological relevance of glycocalyx degradation in hepatic ischemia/reperfusion injury. *Antioxid. Redox Signal.* 21 (7), 1098–1118. doi:10.1089/ars.2013.5751

van Horssen, J., Otte-Holler, I., David, G., Maat-Schieman, M. L., van den Heuvel, L. P., Wesseling, P., et al. (2001). Heparan sulfate proteoglycan expression in cerebrovascular amyloid beta deposits in Alzheimer's disease and hereditary cerebral hemorrhage with amyloidosis (Dutch) brains. *Acta Neuropathol.* 102 (6), 604–614. doi:10.1007/s004010100414

Vink, H., and Duling, B. R. (2000). Capillary endothelial surface layer selectively reduces plasma solute distribution volume. *Am. J. Physiol. Heart Circ. Physiol.* 278 (1), H285–H289. doi:10.1152/ajpheart.2000.278.1.H285

Walter, F. R., Santa-Maria, A. R., Meszaros, M., Veszelka, S., Der, A., and Deli, M. A. (2021). Surface charge, glycocalyx, and blood-brain barrier function. *Tissue Barriers* 9 (3), 1904773. doi:10.1080/21688370.2021.1904773

Wang, Y., Branicky, R., Noe, A., and Hekimi, S. (2018). Superoxide dismutases: dual roles in controlling ROS damage and regulating ROS signaling. *J. Cell. Biol.* 217 (6), 1915–1928. doi:10.1083/jcb.201708007

Webster, S. J., Bachstetter, A. D., Nelson, P. T., Schmitt, F. A., and Van Eldik, L. J. (2014). Using mice to model Alzheimer's dementia: an overview of the clinical disease and the preclinical behavioral changes in 10 mouse models. *Front. Genet.* 5, 88. doi:10.3389/fgene.2014.00088

Williams, L. R., and Leggett, R. W. (1989). Reference values for resting blood flow to organs of man. *Clin. Phys. Physiol. Meas.* 10 (3), 187–217. doi:10.1088/0143-0815/10/3/001

Williams-Medina, A., Deblock, M., and Janigro, D. (2020). *In vitro* models of the blood-brain barrier: tools in translational medicine. *Front. Med. Technol.* 2, 623950. doi:10.3389/fmed.2020.623950

Winkler, E. A., Nishida, Y., Sagare, A. P., Rege, S. V., Bell, R. D., Perlmuter, D., et al. (2015). GLUT1 reductions exacerbate Alzheimer's disease vasculo-neuronal dysfunction and degeneration. *Nat. Neurosci.* 18 (4), 521–530. doi:10.1038/nn.3966

Woodcock, T. E., and Woodcock, T. M. (2012). Revised Starling equation and the glycocalyx model of transvascular fluid exchange: an improved paradigm for prescribing intravenous fluid therapy. *Br. J. Anaesth.* 108 (3), 384–394. doi:10.1093/bja/aer515

Yalcin, O., Jani, V. P., Johnson, P. C., and Cabrales, P. (2018). Implications enzymatic degradation of the endothelial glycocalyx on the microvascular hemodynamics and the arteriolar red cell free layer of the rat cremaster muscle. *Front. Physiol.* 9, 168. doi:10.3389/fphys.2018.00168

Yan, L., Dwiggins, C. W., Gupta, U., and Stroka, K. M. (2023). A rapid-patterning 3D vessel-on-chip for imaging and quantitatively analyzing cell-cell junction phenotypes. *Bioeng. (Basel)* 10 (9), 1080. doi:10.3390/bioengineering10091080

Yang, R., Chen, M., Zheng, J., Li, X., and Zhang, X. (2021). The role of heparin and glycocalyx in blood-brain barrier dysfunction. *Front. Immunol.* 12, 754141. doi:10.3389/fimmu.2021.754141

Yang, X., Meegan, J. E., Jannaway, M., Coleman, D. C., and Yuan, S. Y. (2018). A disintegrin and metalloproteinase 15-mediated glycocalyx shedding contributes to vascular leakage during inflammation. *Cardiovasc. Res.* 114 (13), 1752–1763. doi:10.1093/cvr/cvy167

Yang, Y., and Schmidt, E. P. (2013). The endothelial glycocalyx: an important regulator of the pulmonary vascular barrier. *Tissue Barriers* 1 (1), e23494. doi:10.4161/tisb.23494

Yen, W., Cai, B., Yang, J., Zhang, L., Zeng, M., Tarbell, J. M., et al. (2015). Endothelial surface glycocalyx can regulate flow-induced nitric oxide production in microvessels *in vivo*. *PLoS One* 10 (1), e0117133. doi:10.1371/journal.pone.0117133

Yokoyama, M., Kobayashi, H., Tatsumi, L., and Tomita, T. (2022). Mouse models of Alzheimer's disease. *Front. Mol. Neurosci.* 15, 912995. doi:10.3389/fnmol.2022.912995

Yoon, J. H., Lee, E. S., and Jeong, Y. (2017). *In vivo* imaging of the cerebral endothelial glycocalyx in mice. *J. Vasc. Res.* 54 (2), 59–67. doi:10.1159/000457799

Yoon, J. H., Shin, P., Joo, J., Kim, G. S., Oh, W. Y., and Jeong, Y. (2022). Increased capillary stalling is associated with endothelial glycocalyx loss in subcortical vascular dementia. *J. Cereb. Blood Flow. Metab.* 42 (8), 1383–1397. doi:10.1177/0271678X221076568

Yu, H., Kalogeris, T., and Korthuis, R. J. (2019). Reactive species-induced microvascular dysfunction in ischemia/reperfusion. *Free Radic. Biol. Med.* 135, 182–197. doi:10.1016/j.freeradbiomed.2019.02.031

Zeng, Y., Liu, X. H., Tarbell, J., and Fu, B. (2015). Sphingosine 1-phosphate induced synthesis of glycocalyx on endothelial cells. *Exp. Cell. Res.* 339 (1), 90–95. doi:10.1016/j.yexcr.2015.08.013

Zhang, X., O'Callaghan, P., Li, H., Tan, Y., Zhang, G., Barash, U., et al. (2021). Heparanase overexpression impedes perivascular clearance of amyloid- β from murine brain: relevance to Alzheimer's disease. *Acta Neuropathol. Commun.* 9 (1), 84. doi:10.1186/s40478-021-01182-x

Zhao, F., Zhong, L., and Luo, Y. (2021). Endothelial glycocalyx as an important factor in composition of blood-brain barrier. *CNS Neurosci. Ther.* 27 (1), 26–35. doi:10.1111/cns.13560

Zheng, C., Zhou, X.-W., and Wang, J.-Z. (2016). The dual roles of cytokines in Alzheimer's disease: update on interleukins, TNF- α , TGF- β and IFN- γ . *Transl. Neurodegener.* 5 (1), 7. doi:10.1186/s40035-016-0054-4

Zhu, J., Li, X., Yin, J., Hu, Y., Gu, Y., and Pan, S. (2018). Glycocalyx degradation leads to blood-brain barrier dysfunction and brain edema after asphyxia cardiac arrest in rats. *J. Cereb. Blood Flow. Metab.* 38 (11), 1979–1992. doi:10.1177/0271678X17726062

Zhu, J., Li, Z., Ji, Z., Wu, Y., He, Y., Liu, K., et al. (2021). Glycocalyx is critical for blood-brain barrier integrity by suppressing caveolin1-dependent endothelial transcytosis following ischemic stroke. *Brain Pathol.* 32, e13006. doi:10.1111/bpa.13006

Zhu, W. M., Neuhaus, A., Beard, D. J., Sutherland, B. A., and DeLuca, G. C. (2022). Neurovascular coupling mechanisms in health and neurovascular uncoupling in Alzheimer's disease. *Brain* 145 (7), 2276–2292. doi:10.1093/brain/awac174

Zou, Z., Li, L., Schafer, N., Huang, Q., Maegele, M., and Gu, Z. (2021). Endothelial glycocalyx in traumatic brain injury associated coagulopathy: potential mechanisms and impact. *J. Neuroinflammation* 18 (1), 134. doi:10.1186/s12974-021-02192-1