



# Strategic vaccine stockpiles for regional epidemics of emerging viruses: A geospatial modeling framework

Colin J. Carlson<sup>a,b</sup>, Romain Garnier<sup>a</sup>, Andrew Tiu<sup>a</sup>, Stephen P. Luby<sup>c</sup>, Shweta Bansal<sup>a,\*</sup>

<sup>a</sup> Department of Biology, Georgetown University

<sup>b</sup> Department of Epidemiology of Microbial Diseases, Yale University School of Public Health

<sup>c</sup> School of Medicine, Stanford University

## ABSTRACT

Multinational epidemics of emerging infectious diseases are increasingly common, due to anthropogenic pressure on ecosystems and the growing connectivity of human populations. Early and efficient vaccination can contain outbreaks and prevent mass mortality, but optimal vaccine stockpiling strategies are dependent on pathogen characteristics, reservoir ecology, and epidemic dynamics. Here, we model major regional outbreaks of Nipah virus and Middle East respiratory syndrome, and use these to develop a generalized framework for estimating vaccine stockpile needs based on spillover geography, spatially-heterogeneous healthcare capacity and spatially-distributed human mobility networks. Because outbreak sizes were highly skewed, we found that most outbreaks were readily contained (median stockpile estimate for MERS-CoV: 2,089 doses; Nipah: 1,882 doses), but the maximum estimated stockpile need in a highly unlikely large outbreak scenario was 2–3 orders of magnitude higher (MERS-CoV: ~87,000 doses; Nipah ~ 1.1 million doses). Sensitivity analysis revealed that stockpile needs were more dependent on basic epidemiological parameters (i.e., death and recovery rate) and healthcare availability than any uncertainty related to vaccine efficacy or deployment strategy. Our results highlight the value of descriptive epidemiology for real-world modeling applications, and suggest that stockpile allocation should consider ecological, epidemiological, and social dimensions of risk.

## 1. Introduction

Recent decades have witnessed a marked increase in animal-to-human spillover of pathogens with epidemic and pandemic potential, likely as a result of changes in ecosystem function, agriculture, climate, and human-wildlife contact [10,18,39,53,54,65]. Due to rising global mobility, a greater proportion of these outbreaks have become serious epidemics [11,63,84,66,89]. The 2012 emergence of the Middle East respiratory syndrome coronavirus (MERS-CoV) marked the start of a decade of prolonged multinational outbreaks of viruses that have conventionally been seen as readily contained by a rapid response [19,38]. In the time since, this pattern has recurred in regional epidemics of Ebola virus [46], explosive epidemics of mosquito-borne diseases in the Americas [72], the Covid-19 pandemic, and most recently, the 2022 global mpox outbreak [3].

The scale of these outbreaks—and the challenges of non-pharmaceutical interventions at those scales—have increasingly made vaccination the first (and sometimes only) line of defense. After the 2014 epidemic in West Africa, vaccines have become a standard component of Ebola virus outbreak response [81,100,102]; similarly, *Vaccinia*-based vaccines developed for smallpox are also effective against mpox [80],

and were instrumental in containing the 2022 outbreak. For novel pathogens, lineages, or variants, the speed of the R&D pipeline has been a subject of historical concern [104], but the Covid-19 pandemic has demonstrated the feasibility of comparatively rapid manufacturing and rollout, as well as the value of investments in novel vaccine platform technologies [101,22, 70]. Anticipated progress towards the development of universal vaccines will further improve preparedness for novel pathogens [7,23].

Proactive investments in vaccine stockpiles could help contain outbreaks earlier, and prevent larger epidemics and pandemics altogether [52]. Equally important, they could reduce the drastic inequity that has resulted from Global North monopolization of intellectual property, manufacturing capacity, and vaccine allocation that has been a hallmark of both the Covid-19 and mpox responses. Though the optimal design of vaccine stockpiles is a familiar problem in quantitative epidemiology [67, 91, 93], less work has explored application to emerging pathogens [60].

Here, we develop a stochastic geospatial modeling framework that accounts for multiscale dynamics of disease emergence, including spillover risk, spatial heterogeneity in population distribution and healthcare infrastructure, and spatial connectivity via human mobility.

\* Corresponding author.

E-mail address: [shweta.bansal@georgetown.edu](mailto:shweta.bansal@georgetown.edu) (S. Bansal).

<https://doi.org/10.1016/j.vaccine.2024.06.019>

Received 29 January 2024; Received in revised form 3 June 2024; Accepted 5 June 2024

Available online 19 June 2024

0264-410X/© 2024 Elsevier Ltd. All rights reserved, including those for text and data mining, AI training, and similar technologies.

We demonstrate how vaccine stockpiling can be approached as a spatial design problem, and highlight where future work can reduce model uncertainty.

## 2. Methods

Our project was conducted in 2019 as a quantitative exercise in support of the Coalition for Epidemic Preparedness Innovations (CEPI), which coordinates a global effort to develop novel vaccines and vaccine technologies that can reduce epidemic and pandemic risk. Since 2017, CEPI has funded the development of several dozen vaccine candidates, with a core focus on six priority pathogens (Lassa virus, MERS-CoV, Rift Valley fever, Nipah virus, chikungunya virus, and Ebola virus), as well as (since 2020) Covid-19 and other coronaviruses yet to emerge. In service of that effort, our project modeled the vaccine stockpile needs that would arise in a major regional multi-national epidemic of two of these pathogens (Nipah virus and MERS-CoV). Our models are intended to reproduce the characteristics of real, historical outbreaks of these pathogens. While concerns have been raised that new lineages of either pathogen could arise with greater capacity for human-to-human transmission [87,64,87,64], we focus here on regional epidemics rather than high transmissibility scenarios in which global mass vaccination would be necessary. To address other aspects of uncertainty, we conduct sensitivity analysis on over a dozen epidemiological parameters, and highlight the impact of data gaps.

Features our focal pathogens share include (1) frequent zoonotic (animal-to-human) spillover, (2) an anticipated risk of unexpectedly-large regional outbreaks, and (3) ongoing vaccine development efforts. Nipah virus is a bat-origin virus that was first described from a 1998 outbreak in Malaysia. Since then, there have been sixteen re-emergence events with most occurring in Bangladesh (10 outbreaks) or India (6 outbreaks) [35]. Secondary transmission is limited but nontrivial, with outbreaks usually limited to a few dozen cases [74]. Most recently, in August 2023, an unusually large outbreak occurred in Kerala, India, with 30 cases of infection, and approximately half involving human-to-human transmission [92]. Similarly, MERS-CoV also likely originated in bats [6], but is maintained in domesticated camels. Though its capacity for human-to-human spread has so far been middling (particularly compared to SARS-CoV-2) [31], over 2,600 cases have been reported since 2012 in over two dozen countries, with 84 % of cases occurring in Saudi Arabia [107]. Despite limited evidence of community spread, both Nipah and MERS-CoV have a notable track record of spreading in healthcare settings: in the 2015 outbreak of MERS-CoV, a single infected patient seeking treatment at multiple facilities in South Korea sparked an outbreak of 186 cases, including 25 healthcare workers [59,49]; during the 2018 Kerala outbreak of Nipah virus, nosocomial transmission accounted for most or all secondary cases [8], and more limited instances have been reported in smaller outbreaks [90,86].

We focus our attention on a geographic region for each pathogen: for Nipah, our focal region consists of India and Bangladesh in the Indian subcontinent; for MERS-CoV, our focal region consists of 21 countries of the Middle East and Northeast Africa (Bahrain, Djibouti, Egypt, Eritrea, Ethiopia, Iran, Iraq, Israel, Jordan, Kuwait, Lebanon, Oman, Palestine, Qatar, Saudi Arabia, Somalia, South Sudan, Sudan, Syria, United Arab Emirates, and Yemen).

To estimate vaccine stockpile needs for these two pathogens, we developed a multilayer modeling framework that predicts local and regional epidemiological dynamics post-emergence. Our approach accounts for geographic heterogeneity in both spillover risk and human-to-human transmission, drawing on previous frameworks developed for Ebola virus and influenza [84,77,15]. As a novel extension, we added a third layer for vaccination, with multiple strategies represented. All simulations and analyses were conducted in Python 3.7 and R 3.5.

### Stage 1. Ecological risk factors for emergence.

Fine-scale mapping of spillover risk poses a substantial challenge,

particularly for pathogens with only a few dozen recorded animal-to-human transmission events. Machine learning is often used to predict hotspots of spillover intensity, including for Nipah [26,76,98] and MERS-CoV [85], but these approaches are sensitive to surveillance gaps, and may be prone to surprises (i.e., outbreak risk may be underestimated in areas where spillover has not been previously observed). For a more flexible approach, we examined a set of basic geospatial datasets that captured animal reservoirs and their contact patterns with humans, and aimed to identify a subset that were reflective of known outbreak history [26,83]. (More details can be found in the [Supporting Information](#)).

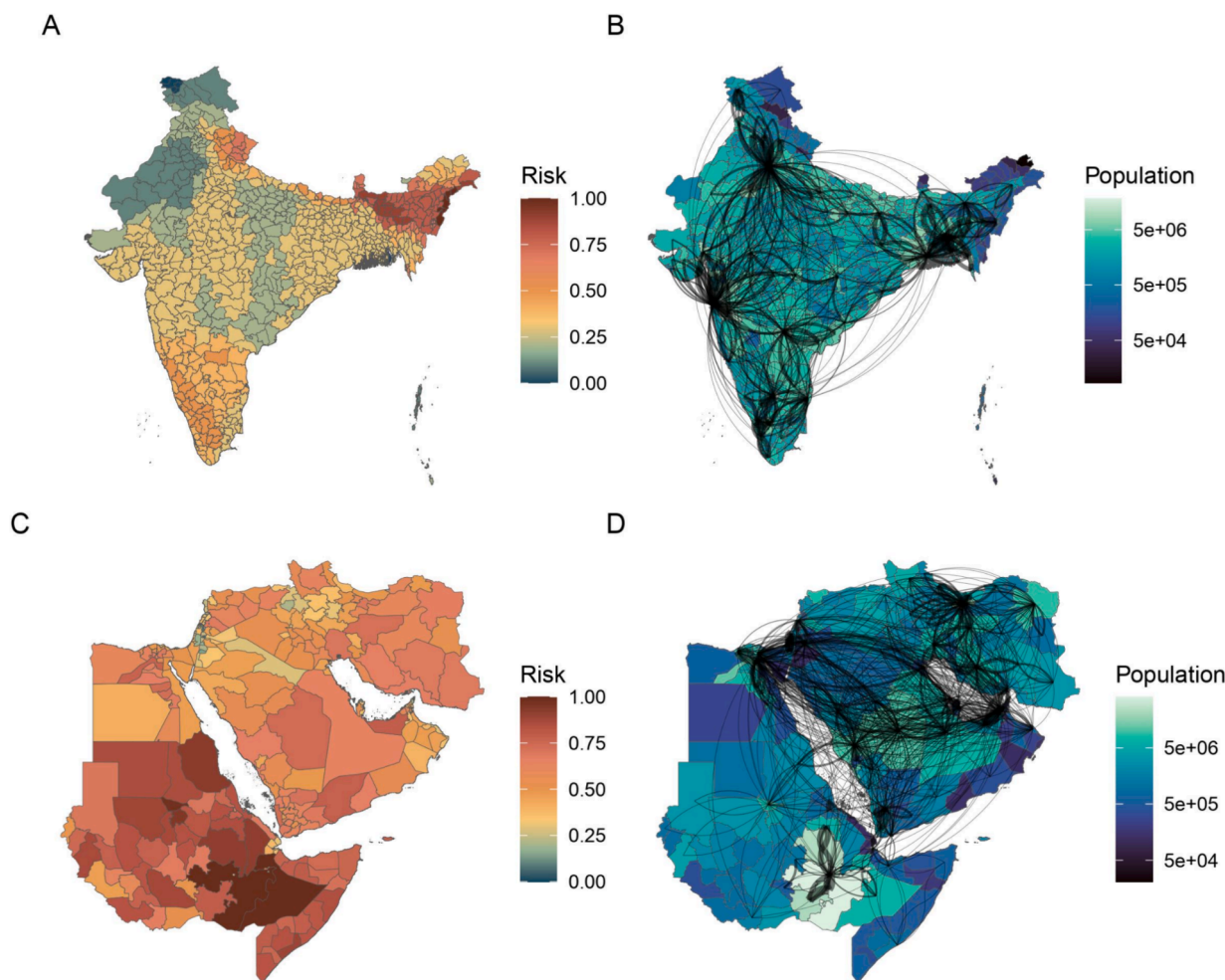
Since its emergence in 1998, Nipah virus has exhibited two broad syndromes of spillover risk. Sometimes, the virus reaches humans through an intermediate host, such as pigs (in the 1998 outbreak in Malaysia) [82] or horses (in the 2014 outbreak in the Philippines [21]). In Bangladesh, where Nipah virus spillover occurs more regularly, the virus has reached humans directly from bat reservoirs, usually through date palm sap contaminated with bat secretions. We chose to use the richness of known bat host species as a coarse proxy for viral circulation in reservoirs, and therefore as a reasonable first-principles approximation of risk [78]. We also explored additional predictors that captured the conversion of intact forest to agricultural mosaic landscapes or palm monoculture. [68,33,14]. These datasets correctly approximated some facets of outbreak history: in particular, forest cover (and so deforestation) is relatively high in Kerala, and date palm production is moderately high in southern Bangladesh. However, in aggregate, these factors would suggest a high level of risk in eastern and northeastern India, and low risk in Bangladesh or west Bengal, where most known spillovers have occurred; therefore, we decided not to use them in our analysis (see [Supporting Information](#)).

Although MERS-like coronaviruses are found in bats worldwide [6,75,71], the origin of MERS-CoV appears to have been a single bat-to-camel transmission event; the virus has since jumped from camels to humans at least 200 times [83], almost exclusively in livestock keeper populations. Although MERS-CoV circulates in camels throughout northern Africa, the Arabian peninsula, and even southern Asia [27]), reported spillover events have historically been confined to Saudi Arabia, Qatar, Oman, and the United Arab Emirates [83]. However, serological studies have found evidence of MERS-CoV exposure in Kenyan livestock handlers [62,55], and during the preparation of the study, an active case cluster indicative of spillover was finally identified in Kenya [73]. We estimated the risk of spillover based on the average of two scaled, log-transformed predictors ( [Figure S2](#)): the number of camels, based on unpublished data from the United Nations (UN) Food and Agriculture Organization (FAO)'s Gridded Livestock of the World study [40]; and the human population employed in the agricultural sector [97].

Each predictor layer was aggregated from raster layers to an administrative polygon selected based on the spatial extent and population density in the region (state or equivalent first subnational level [i.e., ADM-1] for MERS-CoV, and second subnational administrative level [i.e., ADM-2] for Nipah). These predictor layers were rescaled between zero and one, and for MERS-CoV, the two component layers were averaged ( [Fig. 1](#)). Both of the final risk surfaces reasonably approximate the geography of historical spillover events (see [Appendix S1](#)), while identifying much broader areas where outbreaks could plausibly start someday. We therefore used these layers to drive our simulations, and seeded outbreaks based on a random multinomial sample of administrative units weighted by their scaled emergence risk.

### Stage 2. Epidemiological dynamics post-emergence.

For both pathogens, we modeled human-to-human transmission using a standard susceptible – exposed – infectious – resistant (SEIR) compartmental model, simulated on a metapopulation of administrative units (ADM-1 for MERS and ADM-2 for Nipah) with significant within-population structure. Unless otherwise specified, epidemiological parameters used in the model were estimated from clinical reports (e.g., [9]) or gray literature (e.g., fact sheets from the U.S. Centers for Disease



**Fig. 1.** Spatial heterogeneity driving simulated epidemic dynamics. For both Nipah virus (A,B) and MERS-CoV (C,D), simulations happen in two stages: outbreaks are generated, and then spread on a human metapopulation. In the emergence stage, spillover risk is generated from mechanistic predictions and scaled from 0 to 1 (A,C). In the epidemic stage, spread is governed by population density, healthcare access, and a gravity model for traveler frequency (B,D; network edge weight indicates relative traveler volume; and only larger population centers are shown for visualization purposes). The countries included in the regions are listed in [Tables S1 and S2](#)

Control and Prevention or the World Health Organization). The same set of parameters was included in both models ([Table 1](#)), with the exception of asymptomatic rate: whereas subclinical cases of Nipah virus are generally presumed rare [57,74], we assumed that a significant portion of MERS-CoV cases are likely to be subclinical [16,42,5]. For these subclinical MERS cases, we assumed that they would be both non-infectious (transitioning directly from the exposed to recovered class) and functionally asymptomatic (and therefore do not trigger ring vaccination) given limited evidence for asymptomatic MERS transmission [42,30].

Each administrative unit was treated as a distinct population, with a total estimated size based on Gridded Population of the World version 4 [20]. Outbreaks spread spatially based on a gravity model ([Fig. 1](#); additional details in the [Supporting Information](#)), which scales probability of infection based on the distance between and population sizes of any two areas; this approach has been previously shown to be a reasonably accurate proxy for epidemic spread at large scales [11]. Outbreaks spread across populations based on either travel of infected (but not hospitalized) persons to new areas, or travel of susceptible persons from new areas to areas with ongoing outbreaks.

Within each population, we modeled multiple layers of transmission. All individuals started in the community layer (i.e., population), with a single infected case to initiate an outbreak. Frequency-dependent transmission was allowed to occur across the entire community, where new infections were generated at time  $t$  a rate of  $\beta I(t)/N$ . Transmission

within closer contact networks was modeled as a second layer, where each new infection from community transmission would expose a new “household” (close contact network) to elevated risk, based on an average number of within-household or otherwise close contacts  $\tau$  estimated from previous outbreaks of both Nipah virus and MERS-CoV [44,30]. This higher-risk proportion of the social network expanded as a function of the cumulative number of infections from community transmission  $I(\infty)$ , and new infections were generated at a rate of  $\beta I(t)/(\tau I(\infty))$ .

Finally, we treated hospitalization and nosocomial transmission as a distinct layer of population structure. Infected persons were randomly hospitalized at a daily rate, based on literature estimates of average time to hospitalization for MERS-CoV in Saudi Arabia [30] and, as a proxy for Nipah virus, malaria in India [108,88]; new data on Nipah virus outbreaks in Bangladesh, published during our study, support this approximation [74]. We assume that patients are not isolated immediately, and thus allow nosocomial transmission was also assumed to be frequency dependent, where hospitalized patients  $H$  infected a mix of other patients and healthcare workers  $M$  at a rate of  $\beta H(t)/M$ . We used World Bank estimates of beds per 1,000 persons at a national level to estimate hospital capacity [103], and assumed 25 % of beds were occupied at the start of an outbreak. We used the same source for national estimates of the number of clinicians per 1,000 persons, and assumed that supplemented with additional data from the World Health Organization and local ministries of health when necessary (e.g., for

**Table 1**

Key epidemiological parameter estimates used in the simulations, and parameter ranges explored in the global sensitivity analysis. (Abbreviations: HCW = healthcare worker; s.d. = standard deviation.).

Parameter	MERS-CoV		Nipah virus	
	Estimate	Range	Estimate	Range
Transmission rate (per day per person)( $\beta$ )	0.225	–	0.40	–
Rate of progression to infectious (per day)	1/6	(1/14, 1/2)	1/8	(1/14, 1/5)
Recovery rate (per day)	1/30	(1/40, 1/2)	1/10	(1/15, 1/3)
Case fatality rate (per day)	1/12	(1/20, 1/5)	1/3	(1/10, 1/2)
Asymptomatic rate	0.70	(0.30, 0.90)	0.0	–
Number of contacts (mean)( $\tau$ )	11	–	20	–
Number of contacts (s.d.)	4	–	16	–
Delay for contact tracing (days)	7	(1, 14)	7	(1, 14)
Number of missed contacts (mean)	3	–	3	–
Number of missed contacts (s.d.)	5	–	5	–
Mean time to hospital (days)	6	(3, 12)	3	(1, 10)
Proportion of risky contacts for HCWs	0.05	(0, 0.5)	0.05	(0, 0.5)
Vaccine efficacy	0.75	(0.30, 0.95)	0.75	(0.30, 0.95)
Time from vaccination to protection (days)	14	(7, 28)	14	(7, 28)
Cases to initiate vaccination (per district)	2	(1, 10)	2	(1, 10)
Time until vaccine availability (days)	7	(1, 28)	7	(1, 28)
Ring vaccination rate	1/10	(1/14, 1/4)	1/10	(1/14, 1/4)
Healthcare worker vaccination rate	1/3	(1/7, 1)	1/3	(1/7, 1)

South Sudan and Palestine; [Tables S1 and S2](#)). Because no finer-scale data was available, we assumed that healthcare workers were distributed proportionally to population within countries across administrative units. We also assumed that, due to behavioral risk reduction, access to personal protective equipment, and other factors [\[45\]](#), patient-clinician contact is significantly lower risk than contact within the general population.

### Stage 3. Vaccination

A number of vaccine candidates are already in various stages of development and clinical trials for both MERS-CoV and Nipah virus [\[36,29,17,69,4,51\]](#), but so far, none have been licensed for use in human populations. Given the resulting uncertainty, and for modeling simplicity, we assumed a one-dose regimen for both vaccines, as indicated in the target product profile from the WHO [\[105, 106\]](#). We also assume that vaccination results in sterilizing immunity (vaccinated individuals are no longer susceptible), but that immunity takes time to be conferred, that efficacy is less than 100 %, and that there could be delays in vaccine availability after public health response is first mobilized.

In epidemic simulations, we examined three possible strategies for vaccination: (1) Vaccination of healthcare workers only: once vaccination thresholds are passed and vaccines are available, all healthcare workers are rapidly vaccinated. (2) Healthcare worker vaccination as in scenario 1, plus ring vaccination: once vaccination thresholds are passed and vaccines are available, the individuals in contact with a sick person (i.e., all those within the household layer) are vaccinated within 10 days. However, a small proportion of contacts are assumed to be missed by this process, allowing some onward transmission to slip through. (3) Healthcare worker and ring vaccination as in scenario 2, plus catch-up vaccination: after an additional delay to allow for contact tracing, all “missed contacts” are also identified and vaccinated.

### Model implementation and sensitivity analysis.

Epidemic simulations were stochastic at every level, requiring a substantial number of iterations to fully sample the possible range of outbreaks. For both pathogens, we simulated 10,000 outbreaks until 10 days after the last case appeared (in communities or hospitals), or for a maximum of one calendar year (365 days), at which point simulations were interrupted. In our model,  $R_0$  is dependent not only on transmission rate, but also on recovery rate, death rate, and time to hospitalization. We approximated  $R_0$  in the community layer for a deterministic formulation of our model, and found that the  $R_0$  value ranged from 0.31 to 1.03 (mean  $\pm$  s.d.:  $0.57 \pm 0.15$ ) for Nipah virus and 0.07 to 0.41 (mean  $\pm$  s.d.:  $0.21 \pm 0.09$ ) for MERS-CoV. (The large difference between these estimates is due to the effect of including non-infectious asymptomatic cases of MERS-CoV.).

Because simulations had several layers of complexity, and not all parameters were equally supported by peer-reviewed literature, we conducted a global sensitivity analysis to evaluate how vaccine stockpile needs changed in response to model parameterization. We selected 12 parameters for Nipah virus (excluding asymptomatic rate, which is negligible) and 13 parameters for MERS-CoV, and chose not to examine variation in transmission rate  $\beta$ , given that estimates were based on data from previous outbreaks. We executed the sensitivity analysis using Latin hypercube sampling (LHS) to select 50 parameter sets with non-overlapping values from the ranges reported in [Table 1](#). We then analyzed the results of the sensitivity analysis using generalized linear mixed models (GLMMs) to characterize how the number of vaccine doses was explained by each parameter.

## 3. Results

### Vaccine stockpile needs.

Regardless of pathogen or vaccination strategy, we found that the majority of simulated outbreaks only required a few thousand vaccine doses for containment. This was unsurprising given the epidemiology of both pathogens, which has historically been self-limiting except in rare cases. Our simulations reproduced this dynamic intentionally: in roughly a quarter of MERS-CoV simulations, and nearly a third of Nipah virus simulations, index cases failed to produce any secondary cases (and so no vaccines were deployed).

We examined the number of doses needed across a range of quantiles as a way of capturing different levels of acceptable risk for stockpile size ([Table 2](#)). Even with full ring and catch-up vaccination, only  $\sim 2,000$  doses were needed to contain a median-sized outbreak of either pathogen. However, our simulation also reproduced a strong overdispersion of outbreak sizes, where the largest epidemics were several orders of magnitude larger than the median. In the 99th percentile of outbreaks, around  $\sim 150\text{--}180,000$  doses of Nipah vaccines and  $\sim 50,000$  doses of

**Table 2**

Number of doses used at the end of one year (average across simulations). Full distributions are visualized in [Figure S3](#).

Percentile of outbreak size	Vaccination strategy 1: healthcare workers only	Vaccination strategy 2: 1 + ring vaccination	Vaccination strategy 3: 2 + catch-up vaccination
MERS-CoV			
Median	1,236	1,523	2,089
75th	5,944	6,561	6,826
90th	15,191	15,426	16,321
95th	21,096	20,929	21,647
99th	53,638	48,542	50,212
Maximum	84,367	102,788	86,690
Nipah virus			
Median	1,246	1,859	1,882
75th	5,260	6,207	6,168
90th	12,313	14,202	14,181
95th	23,396	29,507	29,532
99th	185,747	162,400	151,320
Maximum	2,105,972	1,019,554	1,128,042



MERS-CoV vaccines could be needed; in the maximum-sized outbreaks we simulated, up to  $\sim 1$ –2 million doses of Nipah vaccines and  $\sim 85$ –100,000 doses of MERS-CoV vaccines could be needed during the first year of an epidemic. These estimates were generally independent of vaccination strategy; the increased number of doses needed for more intensive strategies were generally compensated for by reductions in transmission. However, in the largest Nipah virus outbreaks, we found that community vaccination substantially reduced total needs compared to only vaccinating healthcare workers.

Along this gradient of outbreak sizes, the geography of vaccination needs changed substantially (Fig. 2). Larger outbreaks were more likely to reach major population centers, especially for Nipah virus, for which simulated spillover risk was independent of population density. As a result, vaccination needs in smaller outbreaks more closely tracked spillover risk, while in larger outbreaks, the greatest needs were in cities like Riyadh, Saudi Arabia or Delhi, India. In a major outbreak, vaccination needs in these cities could easily be 3–5 orders of magnitude greater than in rural areas. While this pattern reflected an obvious relationship with population density, it was also heavily driven by the geography of healthcare: for example, vaccination needs were much higher in the high-income parts of the Arabian peninsula than in comparable African cities, due to much higher numbers of healthcare workers.

#### Sensitivity analysis.

In both models, sensitivity analyses found that the strongest determinants of stockpile size were those that shaped  $R_0$  and therefore final outbreak size (Fig. 3). Both a higher death rate and a higher recovery rate could substantially decrease transmission and shorten outbreak duration, while increased delays in hospitalization left more time for uninterrupted community transmission and spatial spread

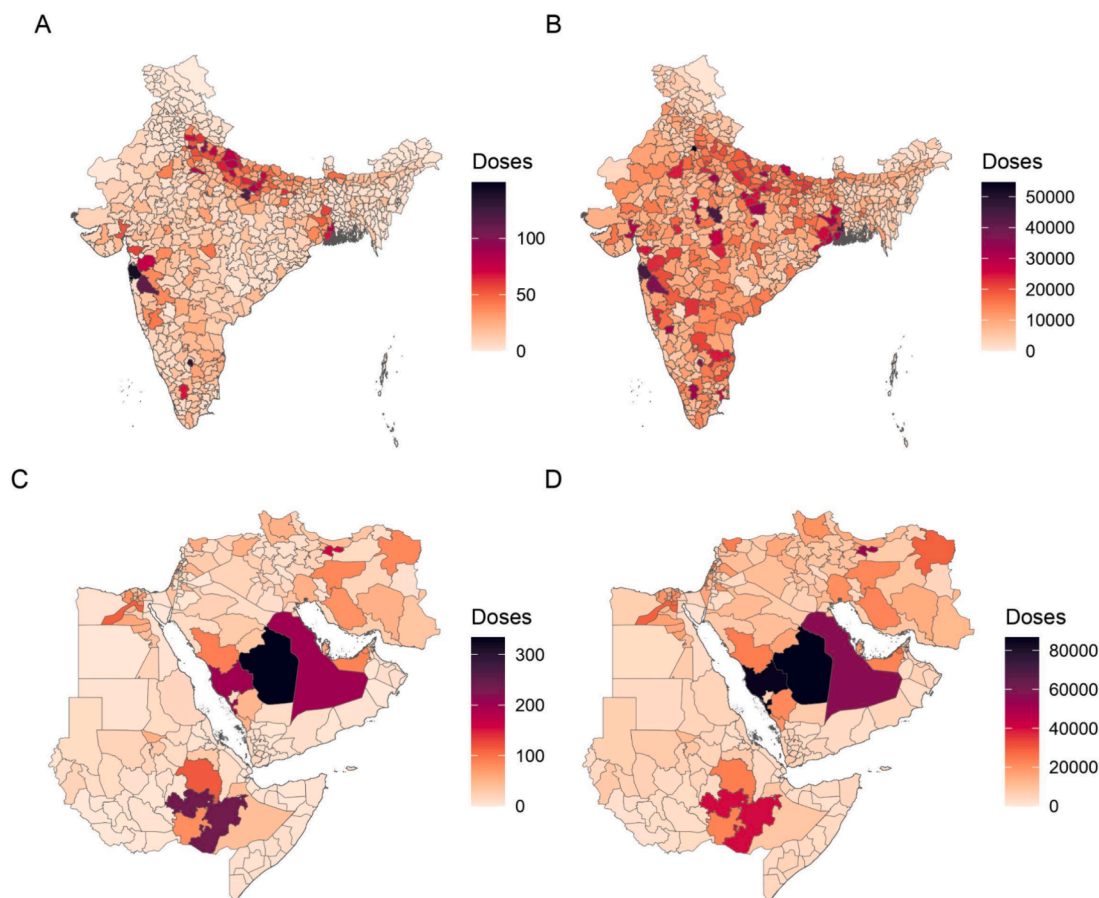
across administrative areas.

In the Nipah virus simulations, the total community vaccination was also somewhat increased by aspects of vaccine rollout, including delays in vaccine availability and, to a minor degree, delays in protection conferred. The effect of both was even less pronounced in the full model, confirming that their effect was through vaccination mediating  $R_0$  in the community.

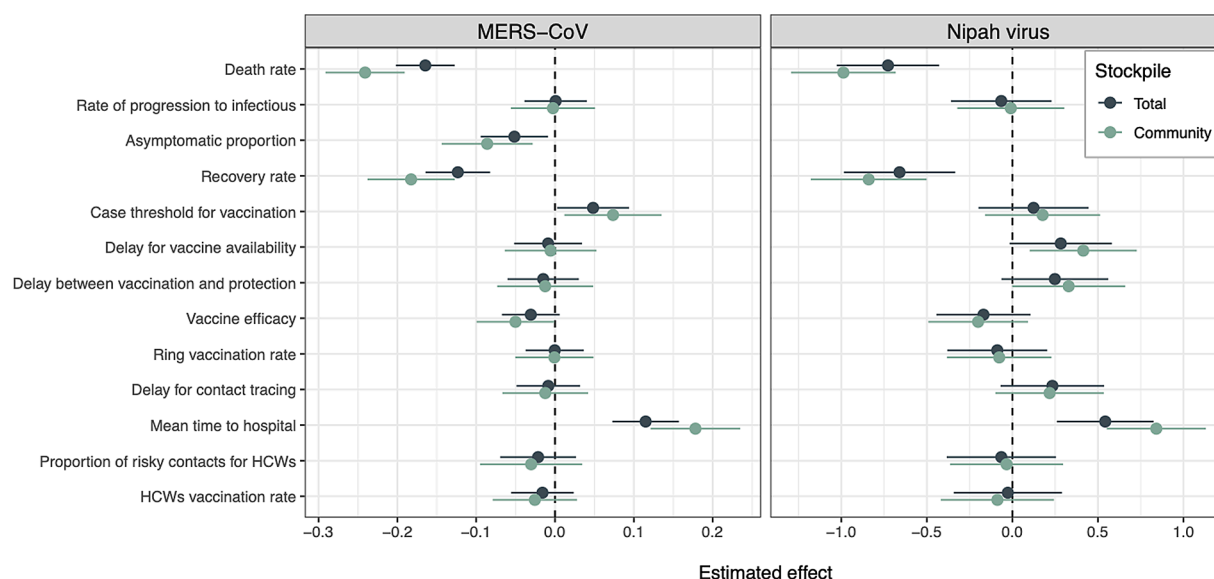
For MERS-CoV, the effects of other parameters were more idiosyncratic. Higher asymptomatic rates and, to a minor degree, higher vaccine efficacy both had a similar effect to death and recovery rates, in both cases by creating more dead ends for onward transmission. Increasing the case threshold for vaccination similarly resulted in delayed containment and increased total community vaccination needs. When healthcare worker vaccination was included, this pattern inverted, due to the design of the simulation exercise: because nosocomial transmission dominated the simulated MERS-CoV outbreaks, higher case thresholds for vaccination and longer delays to vaccine availability both reduced total doses needed, simply by shortening the period over which healthcare worker vaccination was occurring. Total vaccination needs also decreased with a higher asymptomatic rate, and to a lesser degree, with incubation period.

#### 4. Discussion

Using our multiscale modeling framework, we estimated that a few thousand doses would probably be able to suppress a typical outbreak of either pathogen, but an active stockpile of fewer than 1 million doses of MERS-CoV vaccine and at most 2–3 million doses of Nipah virus vaccine will very likely be sufficient for the first year of an acute regional epidemic. Actual needs will be determined by basic epidemiology,



**Fig. 2.** Spatial heterogeneity in predicted outcomes. Vaccination needs for Nipah virus (A,B) and MERS-CoV (C,D), based on the mean (A,C) or maximum (B,D) estimated stockpile size (including ring, catch-up, and healthcare worker vaccination) across 10,000 simulations.



**Fig. 3.** Sensitivity analysis. For each parameter, the mean estimated effect and 95% confidence interval is shown, where effects that do not overlap zero are statistically significant. Values are shown for both the full vaccine stockpile (blue) and the community stockpile (i.e., excluding healthcare worker vaccination; green).

vaccine quality, and deployment strategy, but overall, we found that our estimates were generally robust to these uncertainties. This places our estimated needs comfortably within the range of comparable stockpiles; for example, the WHO's global stockpiles of oral cholera vaccine and yellow fever vaccine both began with a baseline of 2 million doses, and the latter was increased to 6 million doses in 2016 to manage the growing number of outbreaks in both Africa and South America.

More broadly, our study demonstrates that stockpile design is a spatially-structured problem. A significant body of work recognizes that epidemic dynamics of many infectious diseases are heavily shaped by several aspects of human behavior, including animal-human contact [79,31,43], multiscale mobility [11,24], social contact networks [13], patient-clinician interactions [25,96], and several facets of epidemic response. Our study establishes a parsimonious and generalizable framework for simulating these dynamics on real landscapes, parameterized with multiple sources of real-world data, without the opacity, complexity, and computational cost frequently associated with agent-based approaches [2,95]. While our quantitative predictions are dependent on the model structure and parameters driven by data availability, and social and environmental disruptions that increase population density, contact, or mobility may alter these predictions [94,48], our work qualitatively highlights that a spatially-structured emergence and transmission risk landscape leads to geographically heterogeneous stockpile needs.

Developing this modeling framework also highlights how scientific uncertainty is unevenly distributed across different stages of the disease emergence process [77]. For both pathogens, we found that mechanistic proxies had a limited ability to generate realistic landscapes of spillover risk. Datasets cataloging prior spillovers are also workable as a proxy for risk [60], but would have failed to anticipate major recent outbreaks of both pathogens we examined here [8,73]. In the future, global databases of wildlife disease prevalence could be used to develop more precise maps of spillover risk. However, our simulations suggested that this layer of uncertainty is not the most important for stockpile design; in a regional emergency, optimal vaccine allocation will mostly need to track population density and healthcare availability. This finding tracks with previous work simulating Ebola virus epidemics, which found that only 14 % of variation in final spatial extent was determined by outbreak origin [56].

Within the process of human-to-human transmission, we found that the strongest predictors of final epidemic size (and therefore vaccine

stockpile needs) were basic disease characteristics like case fatality and symptomatic rates. This finding underscores the importance of descriptive epidemiology [61,37], and the value of studies that revisit historical outbreaks, in order to better calibrate stockpile needs. Ability to access healthcare was also a key determinant of outbreak progression, underscoring the risk that devastating epidemics will start at the edge of surveillance systems and go undetected for weeks to months [41,99,50]. In contrast, we found that stockpile needs were largely insensitive to the specific features of different vaccines or their distribution. While basic principles of outbreak response generally hold—vaccines that confer sterilizing immunity at a high rate are the most useful for containing outbreaks, and effective community vaccination will best reduce the burden on healthcare systems—a sufficiently large stockpile of any vaccine could be an essential life-saving countermeasure in the coming decade. Given concerns that more transmissible viral variants or relatives exist in reservoirs, and the high probability of another pandemic comparable to Covid-19 in the coming decades [66], the ethical imperatives and financial incentives to establish these stockpiles are substantial. With future progress on the global health security challenges of vaccine equity, vaccine hesitancy, supply chain and cold chain infrastructure, and global health data sharing and coordination, vaccine stockpiles can hold the promise of rapid and effective response to emerging disease outbreaks.

#### CRediT authorship contribution statement

**Colin J. Carlson:** Writing – review & editing, Writing – original draft, Visualization, Validation, Methodology, Investigation, Formal analysis, Data curation, Conceptualization. **Romain Garnier:** Writing – review & editing, Visualization, Supervision, Software, Methodology, Investigation, Funding acquisition, Formal analysis, Data curation, Conceptualization. **Andrew Tiu:** Writing – review & editing, Visualization, Validation, Software, Formal analysis, Data curation. **Stephen P. Luby:** Writing – review & editing. **Shweta Bansal:** Writing – review & editing, Supervision, Resources, Project administration, Methodology, Investigation, Funding acquisition, Formal analysis, Conceptualization.

#### Declaration of competing interest

CJC received funding from the Rockefeller Foundation and the Carnegie Corporation of New York, PAX Sapiens and Open

Philanthropy. SB received funding from the Merck Investigator Studies Program and FluLab Foundation. The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

## Data availability

Code to reproduce the analyses is available at [github.com/bansal-lab/vaccinestockpile](https://github.com/bansal-lab/vaccinestockpile).

## Acknowledgements

This project was made possible with financial support provided by the Coalition for Epidemic Preparedness Innovations, Norway. We thank Alison Bettis, Georgina Murphy, and Mauro Bernuzzi of CEPI for their support during the development of our modeling framework. We are grateful to Emily Gurley (Johns Hopkins University, USA) and B.L. (Bart) Haagmans (Erasmus MC, The Netherlands) for serving as Nipah and MERS experts, respectively; to Steph Seifert (Washington State University, USA) and Dan Becker (University of Oklahoma, USA) for thoughtful feedback on the project; and to Tim Robinson (United Nations Food and Agriculture Organization) and Marius Gilbert (Universite Libre de Bruxelles) for sharing unpublished data on camel population density. CJC was additionally supported by the National Science Foundation, USA grant number BII 2213854, and thanks colleagues in the Verena program for thoughtful conversations about mapping landscapes of zoonotic risk. SB was additionally supported by the National Institutes of Health, USA grant number R01GM123007.

## Appendix A. Supplementary data

[1,12,28,34,32,47,58,76].

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.vaccine.2024.06.019>.

## References

- Ahmed W, Bivins A, Simpson SL, Bertsch PM, Ehret J, Hosegood I, et al. Wastewater Surveillance Demonstrates High Predictive Value for COVID-19 Infection on Board Repatriation Flights to Australia. *Environ Int* 2022;158 (January):106938.
- Ajelli M, Gonçalves B, Balcan D, Colizza V, Hao Hu, Ramasco JJ, et al. Comparing Large-Scale Computational Approaches to Epidemic Modeling: Agent-Based versus Structured Metapopulation Models. *BMC Infect Dis* 2010. <https://doi.org/10.1186/1471-2334-10-190>.
- Alakunle EF, Okeke MI. Monkeypox Virus: A Neglected Zoonotic Pathogen Spreads Globally. *Nat Rev Microbiol* 2022;20(9):507–8.
- Alharbi NK, Padron-Regalado E, Thompson CP, Kupke A, Wells D, Sloan MA, et al. ChAdOx1 and MVA Based Vaccine Candidates against MERS-CoV Elicit Neutralising Antibodies and Cellular Immune Responses in Mice. *Vaccine* 2017; 35(30):3780–3788.
- Al-Tawfiq JA, Gautret P. Asymptomatic Middle East Respiratory Syndrome Coronavirus (MERS-CoV) Infection: Extent and Implications for Infection Control: A Systematic Review. *Travel Med Infect Dis* 2019;27:27–32.
- Anthony SJ, Johnson CK, Greig DJ, Kramer S, Che X, Wells H, et al. Global Patterns in Coronavirus Diversity. *Virus Evol* 2017;3(1):vex012.
- Arevalo CP, Bolton MJ, Le Sage V, Ye N, Furey C, Muramatsu H, et al. A Multivalent Nucleoside-Modified mRNA Vaccine against All Known Influenza Virus Subtypes. *Science* 2022;378(6622):899–904.
- Arun Kumar, Govindakarnavar, Radhakrishnan Chandni, Devendra T. Mourya, Sajeet K. Singh, Rajeev Sadanandan, Preeti Sudan, Balram Bhargava, and Nipah Investigators People and Health Study Group. Outbreak Investigation of Nipah Virus Disease in Kerala, India, 2018. *J Infect Dis* 2019;219(12):1867–78.
- Assiri A, McGeer A, Perl TM, Price CS, Al AA, Rabeeah DAT, et al. Hospital Outbreak of Middle East Respiratory Syndrome Coronavirus. *N Engl J Med* 2013; 369(5):407–16.
- Baker RE, Mahmud AS, Miller IF, Rajeev M, Rasambainarivo F, Rice BL, et al. Infectious Disease in an Era of Global Change. *Nat Rev Microbiol* 2022;20(4): 193–205.
- Balcan D, Colizza V, Gonçalves B, Hao Hu, Ramasco JJ, Vespignani A. Multiscale Mobility Networks and the Spatial Spreading of Infectious Diseases. *PNAS* 2009; 106(51):21484–2149.
- Bangladesh Bureau of Statistics. 2018. *Yearbook of Agricultural Statistics-2017*. Government of the People's Republic of Bangladesh.
- Bansal S, Grenfell BT, Meyers LA. When Individual Behaviour Matters: Homogeneous and Network Models in Epidemiology. *Journal of the Royal Society, Interface / the Royal Society* 2007;4(16):879–91.
- Becker, Daniel J., Peggy Eby, Wyatt Madden, Alison J. Peel, and Raina K. Plowright. 2021. "Ecological Conditions Experienced by Bat Reservoir Hosts Predict the Intensity of Hendra Virus Excretion over Space and Time." *bioRxiv*. Doi: 10.1101/2021.08.19.457011.
- Berger KA, Pigott DM, Tomlinson F, Godding D, Maurer-Stroh S, Taye B, et al. The Geographic Variation of Surveillance and Zoonotic Spillover Potential of Influenza Viruses in Domestic Poultry and Swine. *Open Forum Infect Dis* 2018;5 (12):ofy318.
- Bernard-Stoecklin S, Nikolay B, Assiri A, Saeed AAB, Embarek PKB, El Bushra H, et al. Comparative Analysis of Eleven Healthcare-Associated Outbreaks of Middle East Respiratory Syndrome Coronavirus (MERS-Cov) from 2015 to 2017. *Sci Rep* 2019;9(1):7385.
- Bosaeed M, Balkhy HH, Almaziad S, Aljami HA, Alhatmi H, Alanazi H, et al. Safety and Immunogenicity of ChAdOx1 MERS Vaccine Candidate in Healthy Middle Eastern Adults (MERS002): An Open-Label, Non-Randomised, Dose-Escalation, Phase 1b Trial. *The Lancet Microbe* 2022. [https://doi.org/10.1016/s2666-5247\(21\)00193-2](https://doi.org/10.1016/s2666-5247(21)00193-2).
- Carlson CJ, Albery GF, Merow C, Trisos CH, Zipfel CM, Eskew EA, et al. Climate Change Increases Cross-Species Viral Transmission Risk. *Nature* 2022;607(7919): 555–62.
- Carlson, Colin J., Matthew R. Boyce, Margaret Dunne, Ellie Graeden, Jessica Lin, Yasser Omar Abdellatif, Max A. Palys, Munir Pavez, Alexandra L. Phelan, and Rebecca Katz. 2022. "The World Health Organization's Disease Outbreak News: A Retrospective Database." *PLOS Global Public Health*. Doi: <https://doi.org/10.1371/journal.pgph.0001083>.
- Center for International Earth Science Information Network - CIESIN - Columbia University. 2016. "Gridded Population of the World, Version 4 (GPWv4): Population Count." Palisades, NY: NASA Socioeconomic Data and Applications Center (SEDAC). Doi: 10.7927/H4X63JVC.
- Ching PK, G., Vikki Carr de los Reyes, Maria Nemias Sucaldito, Enrique Tayag, Alah Baby Columna-Vingno, Fedelino F. Malbas Jr, Gilbert C. Bolo Jr, et al. Outbreak of Henipavirus Infection, Philippines, 2014. *Emerg Infect Dis* 2015;21 (2):328–31.
- Cleve M. What the Lightning-Fast Quest for Covid Vaccines Means for Other Diseases. *Nature* 2021;589:16–8.
- Cohen AA, Gnanapragasam PNP, Lee YE, Hoffman PR, Susan Ou, Kakutani LM, et al. Mosaic Nanoparticles Elicit Cross-Reactive Immune Responses to Zoonotic Coronaviruses in Mice. *Science* 2021;371(6530):735–41.
- Colizza V, Barrat A, Barthélemy M, Valleron A-J, Vespignani A. Modeling the Worldwide Spread of Pandemic Influenza: Baseline Case and Containment Interventions. *PLoS Med* 2007;4(1):e13.
- Cooper BS, Medley GF, Scott GM. Preliminary Analysis of the Transmission Dynamics of Nosocomial Infections: Stochastic and Management Effects. *J Hosp Infect* 1999;43(2):131–47.
- Deka MA, Morshed N. Mapping Disease Transmission Risk of Nipah Virus in South and Southeast Asia. *Tropical Medicine and Infectious Disease* 2018;3(2). <https://doi.org/10.3390/tropicalmed3020057>.
- Dighe A, Jombart T, Van Kerkhove MD, Ferguson N. A Systematic Review of MERS-CoV Seroprevalence and RNA Prevalence in Dromedary Camels: Implications for Animal Vaccination. *Epidemics* 2019;29(December):100350.
- Directorate of Economics & Statistics. 2018. *Agricultural Statistics at a Glance 2017*. Government of India, Ministry of Agriculture & Farmers Welfare.
- van Doremalen N, Lambe T, Sebastian S, Bushmaker T, Fischer R, Feldmann F, et al. A Single-Dose ChAdOx1-Vectored Vaccine Provides Complete Protection against Nipah Bangladesh and Malaysia in Syrian Golden Hamsters. *PLoS Negl Trop Dis* 2019;13(6):e0007462.
- Drosten C, Meyer B, Müller MA, Corman VM, Al-Masri M, Hossain R, et al. Transmission of MERS-Coronavirus in Household Contacts. *N Engl J Med* 2014; 371(9):828–35.
- Dudas, Gytis, Luiz Max Carvalho, Andrew Rambaut, and Trevor Bedford. 2018. "MERS-CoV Spillover at the Camel-Human Interface." *eLife* 7 (January). Doi: 10.7554/eLife.31257.
- Eby P, Peel AJ, Hoegh A, Madden W, Giles JR, Hudson PJ, et al. Pathogen Spillover Driven by Rapid Changes in Bat Ecology. *Nature* 2023;613(7943): 340–434.
- Epstein JH, Anthony SJ, Ariful Islam A, Kilpatrick M, Khan SA, Balkey MD, et al. Nipah Virus Dynamics in Bats and Implications for Spillover to Humans. *PNAS* 2020;117(46):29190–201.
- Epstein JH, Prakash V, Smith CS, Daszak P, McLaughlin AB, Meehan G, et al. Henipavirus Infection in Fruit Bats (*Pteropus Giganteus*), India. *Emerg Infect Dis* 2008;14(8):1309–11.
- European Centre for Disease Prevention and Control (ECDC). Factsheet on Nipah virus disease. Accessed May 27, 2024. <https://www.ecdc.europa.eu/en/infectious-diseases-topics/nipah-virus-disease/factsheet-nipah-virus-disease>; 2023.
- Foster, Stephanie L., Courtney Woolsey, Viktoriya Borisevich, Krystle N. Agans, Abhishek N. Prasad, Daniel J. Deer, Joan B. Geisbert, et al. 2022. "A Recombinant VSV-Vectored Vaccine Rapidly Protects Nonhuman Primates against Lethal Nipah Virus Disease." *Proceedings of the National Academy of Sciences*. Doi: 10.1073/pnas.2200065119.
- Fox MP, Murray EJ, Lesko CR, Sealy-Jefferson S. On the Need to Revitalize Descriptive Epidemiology. *Am J Epidemiol* 2022;191(7):1174–2119.
- Fraser C, Riley S, Anderson RM, Ferguson NM. Factors That Make an Infectious Disease Outbreak Controllable. *PNAS* 2004;101(16):6146–51.



- [39] Gibb, Rory, Sadie J. Ryan, David Pigott, Maria del Pilar Fernandez, Renata L. Muylaert, Gregory F. Albery, Daniel J. Becker, et al. 2024. "The Anthropogenic Fingerprint on Emerging Infectious Diseases." *bioRxiv*. Doi: 10.1101/2024.05.22.24307684.
- [40] Gilbert M, Nicolas G, Cinardi G, Van Boeckel TP, Vanwambeke SO, William Wint GR, et al. Global Distribution Data for Cattle, Buffaloes, Horses, Sheep, Goats, Pigs, Chickens and Ducks in 2010. *Sci Data* 2018;5(October):180227.
- [41] Glennon EE, Jephcott FL, Restif O, Wood JLN. Estimating Undetected Ebola Spillovers. *PLoS Negl Trop Dis* 2019;13(6):e0007428.
- [42] Grant R, Malik MR, Elkholy A, Van Kerkhove MD. A Review of Asymptomatic and Subclinical Middle East Respiratory Syndrome Coronavirus Infections. *Epidemiol Rev* 2019;41(1):69–81.
- [43] Gurley ES, Hegde ST, Hossain K, Hossain MS, Sazzad MJ, Hossain MR, et al. Convergence of Humans, Bats, Trees, and Culture in Nipah Virus Transmission, Bangladesh. *Emerg Infect Dis* 2017;23(9):1446–53.
- [44] Gurley ES, Montgomery JM, Jahangir Hossain M, Bell M, Azad AK, Islam MR, et al. Person-to-Person Transmission of Nipah Virus in a Bangladeshi Community. *Emerg Infect Dis* 2007;13(7):1031–107.
- [45] Hadley MB, Blum LS, Mujaddid S, Parveen S, Nuremowla S, Haque ME, et al. Why Bangladeshi Nurses Avoid 'Nursing': Social and Structural Factors on Hospital Wards in Bangladesh. *Soc Sci Med* 2007;64(6):1166–77.
- [46] Hampton, Lee M., Francisco Luquero, Alejandro Costa, Anais Legand, and Pierre Formenty. 2023. "Ebola Outbreak Detection and Response since 2013." *The Lancet. Microbe*, June. Doi: 10.1016/S2666-5247(23)00136-2.
- [47] Hansen MC, Potapov PV, Moore R, Hancher M, Turubanova SA, Tyukavina A, et al. High-Resolution Global Maps of 21st-Century Forest Cover Change. *Science* 2013;342(6160):850–83.
- [48] Hossain B, Shi G, Chen Ajiang Md, Sarker NI, Sohail MS, Sun Z, et al. Climate Change Induced Human Displacement in Bangladesh: Implications on the Livelihood of Displaced Riverine Island Dwellers and Their Adaptation Strategies. *Front Psychol* 2022;13(October):964648.
- [49] Hui DS, Azhar EI, Kim Y-J, Menish ZA, Myoung-Don Oh, Zumla A. Middle East Respiratory Syndrome Coronavirus: Risk Factors and Determinants of Primary, Household, and Nosocomial Transmission. *Lancet Infect Dis* 2018;18(8):e217–27.
- [50] Hulland EN, Wiens KE, Shirude S, Morgan JD, Bertozzi-Villa A, Farag TH, et al. Travel Time to Health Facilities in Areas of Outbreak Potential: Maps for Guiding Local Preparedness and Response. *BMC Med* 2019;17(1):232.
- [51] Ithini DG, Buchholz DW, Shahrazad Ezzatpour I, Abrey Monreal Yu, Cong JS, Bangar AS, et al. Multivalent Viral Particles Elicit Safe and Efficient Immunoprotection against Nipah Hendra and Ebola Viruses. *npj Vaccines* 2022;7(1):166.
- [52] Jarrett, Stephen, Sonia Pagliusi, Rachel Park, Taufik Wilmansyah, Suresh Jadhav, Patricia Correa Santana, K. R. Krishnamurthy, and Lingjiang Yang. 2021. "The Importance of Vaccine Stockpiling to Respond to Epidemics and Remediate Global Supply Shortages Affecting Immunization: Strategic Challenges and Risks Identified by Manufacturers." *Vaccine* X 9 (December): 100119.
- [53] Jones KE, Patel NG, Levy MA, Storeygard A, Balk D, Gittleman JL, et al. Global Trends in Emerging Infectious Diseases. *Nature* 2008;451(7181):990–3.
- [54] Keesing F, Belden LK, Daszak P, Andrew Dobson C, Harvell D, Holt RD, et al. Impacts of Biodiversity on the Emergence and Transmission of Infectious Diseases. *Nature* 2010;468(7324):647–52.
- [55] Kiyong'a, Alice N., Elizabeth A. J. Cook, Nisreen M. A. Okba, Velma Kivali, Chantal Reusken, Bart L. Haagmans, and Eric M. Fèvre. Middle East Respiratory Syndrome Coronavirus (MERS-CoV) Seropositive Camel Handlers in Kenya. *Viruses* 2020;12(4). <https://doi.org/10.3390/v12040396>.
- [56] Kramer AM, Tomlin Pulliam J, Alexander LW, Park AW, Rohani P, Drake JM. Spatial Spread of the West Africa Ebola Epidemic. *R Soc Open Sci* 2016;3(8):160294.
- [57] Kumar CP, Girish AP, Sugunan PY, Kurup KK, Aarathie R, Manickam P, et al. Infections among Contacts of Patients with Nipah Virus, India. *Emerg Infect Dis* 2019;25(5):1007–10.
- [58] La Rosa, G., P. Mancini, C. Veneri, G. Bonanno Ferraro, L. Lucentini, M. Iaconelli, and E. Suffredini. 2022. "Detection of Monkeypox Virus DNA in the Wastewater of an Airport in Rome, Italy: Expanding Environmental Surveillance to Emerging Threats." *bioRxiv*. Doi: 10.1101/2022.08.18.22278932.
- [59] Lee JY, Kim Y-J, Chung EH, Kim D-W, Jeong I, Kim Y, et al. The Clinical and Virological Features of the First Imported Case Causing MERS-CoV Outbreak in South Korea, 2015. *BMC Infect Dis* 2017;17(1):498.
- [60] Lerch A, Ten Bosch QA, Jackson ML, Bettis AA, Bernuzzi M, Murphy GAV, et al. Projecting Vaccine Demand and Impact for Emerging Zoonotic Pathogens. *BMC Med* 2022;20(1):202.
- [61] Lesko CR, Fox MP, Edwards JK. A Framework for Descriptive Epidemiology. *Am J Epidemiol* 2022;191(12):2063–70.
- [62] Liljander A, Meyer B, Jores J, Müller MA, Lattwein E, Njeru I, et al. MERS-CoV Antibodies in Humans, Africa, 2013–2014. *Emerg Infect Dis* 2016;22(6):1086–109.
- [63] Loh EH, Zambrana-Torrelío C, Olival KJ, Bogich TL, Johnson CK, Mazet JAK, et al. Targeting Transmission Pathways for Emerging Zoonotic Disease Surveillance and Control. *Vector Borne Zoonotic Dis* 2015;15(7):432–47.
- [64] Luby SP. The Pandemic Potential of Nipah Virus. *Antiviral Res* 2013;100(1):38–43.
- [65] Mahon, Michael B., Alexandra Sack, O. Alejandro Aleuy, Carly Barbera, Ethan Brown, Heather Buelow, David J. Civitello, et al. 2022. "Global Change Drivers and the Risk of Infectious Disease." *bioRxiv*. Doi: 10.1101/2022.07.21.501013.
- [66] Marani M, Katul GG, Pan WK, Parolari AJ. Intensity and Frequency of Extreme Novel Epidemics. In: *Proceedings of the National Academy of Sciences of the United States of America*; 2021. 118 (35). <https://doi.org/10.1073/pnas.2105482118>.
- [67] Matrajt L, Elizabeth Halloran M, Longini Jr IM. Optimal Vaccine Allocation for the Early Mitigation of Pandemic Influenza. *PLoS Comput Biol* 2013;9(3):e1002964.
- [68] McKee CD, Islam A, Luby SP, Salje H, Hudson PJ, Plowright RK, et al. The Ecology of Nipah Virus in Bangladesh: A Nexus of Land-Use Change and Opportunistic Feeding Behavior in Bats. *Viruses* 2021;13(2). <https://doi.org/10.3390/v13020169>.
- [69] Modjarrad K, Roberts CC, Mills KT, Castellano AR, Paolino K, Muthumani K, et al. Safety and Immunogenicity of an Anti-Middle East Respiratory Syndrome Coronavirus DNA Vaccine: A Phase I, Open-Label, Single-Arm, Dose-Escalation Trial. *Lancet Infect Dis* 2019;19(9):1013–22.
- [70] Moghadas, Seyed M., Pratha Sah, Meagan C. Fitzpatrick, Affan Shoukat, Abhishek Pandey, Thomas N. Vilches, Burton H. Singer, Eric C. Schneider, and Alison P. Galvani. n.d. "COVID-19 Deaths and Hospitalizations Averted by Rapid Vaccination Rollout in the United States." Doi: 10.1101/2021.07.07.21260156.
- [71] Munoz, Norma Rocío Forero, Renata L. Muylaert, Stephanie N. Seifert, Gregory Albery, Daniel Becker, Colin J. Carlson, and Timothée Poisot. 2022. "The Coevolutionary Mosaic of Bat Betacoronavirus Emergence Risk." *Virus Evolution*. <https://doi.org/10.1093/ve/vead/079>.
- [72] Musso D, Rodriguez-Morales AJ, Levi JE, Cao-Lormeau V-M, Gubler DJ. Unexpected Outbreaks of Arbovirus Infections: Lessons Learned from the Pacific and Tropical America. *Lancet Infect Dis* 2018;18(11):e355–61.
- [73] Ngeri I, Hunsperger EA, Tong S, Oyugi J, Jaoko W, Harcourt JL, et al. Outbreak of Middle East Respiratory Syndrome Coronavirus in Camels and Probable Spillover Infection to Humans in Kenya. *Viruses* 2022;14(8). <https://doi.org/10.3390/v14081743>.
- [74] Nikolay B, Henrik Salje M, Jahangir Hossain AKM, Khan D, Sazzad HMS, Rahman M, et al. Transmission of Nipah Virus — 14 Years of Investigations in Bangladesh. *N Engl J Med* 2019;380(19):1804–14.
- [75] Olival KJ, Cryan PM, Amman BR, Baric RS, Blehert DS, Brook CE, et al. Possibility for Reverse Zoonotic Transmission of SARS-CoV-2 to Free-Ranging Wildlife: A Case Study of Bats. *PLoS Pathog* 2020;16(9):e1008758.
- [76] Peterson, A. Townsend. 2015. "Mapping Risk of Nipah Virus Transmission across Asia and across Bangladesh." *Asia-Pacific Journal of Public Health / Asia-Pacific Academic Consortium for Public Health* 27 (2): NP824–32.
- [77] Pigott DM, Deshpande A, Letourneau I, Morozoff C, Reiner Jr RC, Kraemer MUG, et al. Local, National, and Regional Viral Haemorrhagic Fever Pandemic Potential in Africa: A Multistage Analysis. *Lancet* 2017;390(10113):2662–72.
- [78] Plowright RK, Becker DJ, Crowley DE, Washburne AD, Tao Huang PO, Nameer ES, et al. Prioritizing Surveillance of Nipah Virus in India. *PLoS Negl Trop Dis* 2019;13(6):e0007393.
- [79] Plowright RK, Parrish CR, McCallum H, Hudson PJ, Ko AI, Graham AL, et al. Pathways to Zoonotic Spillover. *Nat Rev Microbiol* 2017;15(8):502–10.
- [80] Poland GA, Kennedy RB, Toshi PK. Prevention of Monkeypox with Vaccines: A Rapid Review. *Lancet Infect Dis* 2022;22(12):e349–58.
- [81] Potluri R, Kumar A, Oriol-Mathieu V, Van Effelterre T, Metz L, Bhandari H. Model-Based Evaluation of the Impact of Prophylactic Vaccination Applied to Ebola Epidemics in Sierra Leone and Democratic Republic of Congo. *BMC Infect Dis* 2022;22(1):769.
- [82] Pulliam JRC, Epstein JH, Dushoff J, Rahman SA, Bunning M, Jamaluddin AA, et al. Agricultural Intensification, Priming for Persistence and the Emergence of Nipah Virus: A Lethal Bat-Borne Zoonosis. *Journal of the Royal Society, Interface / the Royal Society* 2012;9(66):89–101.
- [83] Ramshaw RE, Letourneau ID, Hong AY, Hon J, Morgan JD, Osborne JCP, et al. A Database of Geopositioned Middle East Respiratory Syndrome Coronavirus Occurrences. *Sci Data* 2019;6(1):318.
- [84] Redding DW, Atkinson PM, Cunningham AA, Iacono GL, Moses LM, Wood JLN, et al. Impacts of Environmental and Socio-Economic Factors on Emergence and Epidemic Potential of Ebola in Africa. *Nat Commun* 2019;10(1):4531.
- [85] Reeves T, Samy AM, Townsend Peterson A. MERS-CoV Geography and Ecology in the Middle East: Analyses of Reported Camel Exposures and a Preliminary Risk Map. *BMC Res Notes* 2015;8(December):801.
- [86] Sazzad HMS, Jahangir Hossain M, Gurley ES, Ameen KMH, Shahana Parveen M, Islam S, et al. Nipah Virus Infection Outbreak with Nosocomial and Corpse-to-Human Transmission, Bangladesh. *Emerg Infect Dis* 2013;19(2):210–27.
- [87] Schroeder S, Mache C, Kleine-Weber H, Corman VM, Muth D, Richter A, et al. Functional Comparison of MERS-Coronavirus Lineages Reveals Increased Replicative Fitness of the Recombinant Lineage 5. *Nat Commun* 2021;12(1):5324.
- [88] Singh MP, Saha KB, Chand SK, Anvikar A. Factors Associated with Treatment Seeking for Malaria in Madhya Pradesh, India. *Tropical Medicine & International Health: TM & IH* 2017;22(11):1377–84.
- [89] Smith KF, Goldberg M, Rosenthal S, Carlson L, Chen J, Chen C, et al. Global Rise in Human Infectious Disease Outbreaks. *J R Soc Interface* 2014. <https://doi.org/10.1098/rsif.2014.0950>.
- [90] Tan CT, Tan KS. Nosocomial Transmissibility of Nipah Virus. *J Infect Dis* 2001;184(10):1367.
- [91] Tebbens RJ, Duintjer MA, Pallansch JP, Alexander, and Kimberly M. Thompson. Optimal Vaccine Stockpile Design for an Eradicated Disease: Application to Polio. *Vaccine* 2010;28(26):4312–27.
- [92] Thiagarajan K. Nipah Virus: India's Kerala State Moves Quickly to Control Fresh Outbreak. *BMJ* 2023;382(September):2117.
- [93] Thompson KM, Duintjer RJ, Tebbens. Framework for Optimal Global Vaccine Stockpile Design for Vaccine-Preventable Diseases: Application to Measles and



- Cholera Vaccines as Contrasting Examples. *Risk Analysis: An Official Publication of the Society for Risk Analysis* 2016;36(7):1487–509.
- [94] Tofighi M, Asgary A, Tofighi G, Najafabadi MM, Arino J, Amiche A, et al. Estimating Social Contacts in Mass Gatherings for Disease Outbreak Prevention and Management: Case of Hajj Pilgrimage. *Tropical Diseases, Travel Medicine and Vaccines* 2022;8(1):19.
- [95] Tracy M, Cerdá M, Keyes KM. Agent-Based Modeling in Public Health: Current Applications and Future Directions. *Annu Rev Public Health* 2018;39(April):77–94.
- [96] Ueno T, Masuda N. Controlling Nosocomial Infection Based on Structure of Hospital Social Networks. *J Theor Biol* 2008;254(3):655–66.
- [97] United Nations International Strategy for Disaster Reduction. 2015. *2015 Global Assessment Report on Disaster Risk Reduction: Making Development Sustainable - The Future of Disaster Risk Reduction*. United Nations.
- [98] Walsh MG. Mapping the Risk of Nipah Virus Spillover into Human Populations in South and Southeast Asia. *Trans R Soc Trop Med Hyg* 2015;109(9):563–71.
- [99] Weiss DJ, Nelson A, Vargas-Ruiz CA, Gligorić K, Bavadekar S, Gabrilovich E, et al. Global Maps of Travel Time to Healthcare Facilities. *Nat Med* 2020;26(12):1835–2188.
- [100] Wells, Chad R., Abhishek Pandey, Alyssa S. Parpia, Meagan C. Fitzpatrick, Lauren A. Meyers, Burton H. Singer, and Alison P. Galvani. 2019. "Ebola Vaccination in the Democratic Republic of the Congo." *Proceedings of the National Academy of Sciences of the United States of America* 116 (20): 10178–83.
- [101] Więcek W. Clinical Trials for Accelerating Pandemic Vaccines. *Oxf Rev Econ Policy* 2022;38(4):797–817.
- [102] Worden, Lee, Rae Wannier, Nicole A. Hoff, Kamy Musene, Bernice Selo, Mathias Mossoko, Emile Okitolonda-Wemakoy, et al. 2019. "Projections of Epidemic Transmission and Estimation of Vaccination Impact during an Ongoing Ebola Virus Disease Outbreak in Northeastern Democratic Republic of Congo, as of Feb. 25, 2019." *PLOS Neglected Tropical Diseases*. Doi: 10.1371/journal.pntd.0007512.
- [103] World Bank. *World Development Indicators 2012*. World Bank 2012. Publications.
- [104] World Health Organization (WHO). 2017a. "Influenza Vaccine Response during the Start of a Pandemic: Report of the Third WHO Informal Consultation." 2017. <https://apps.who.int/iris/bitstream/handle/10665/325973/WHO-WHE-IHM-2019.5-eng.pdf>.
- [105] World Health Organization (WHO). WHO Target Product Profiles for MERS-CoV Vaccines. Accessed May 27, 2024. <https://cdn.who.int/media/docs/default-source/documents/publications/mers-target-product-profile132626f7-b4b1-4757-8056-ae1558311dba.pdf>; 2017.
- [106] World Health Organization (WHO). WHO Target Product Profile for Nipah virus Vaccine. Accessed May 27, 2024. [https://web.archive.org/web/20211122110114/http://www.who.int/blueprint/priority-diseases/key-action/Nipah\\_virus\\_vaccineTPP.pdf?ua=1](https://web.archive.org/web/20211122110114/http://www.who.int/blueprint/priority-diseases/key-action/Nipah_virus_vaccineTPP.pdf?ua=1); 2017.
- [107] Update March 2024".. Accessed May 27, 2024, <https://applications.emro.who.int/docs/WHOEMCSR741E-eng.pdf>.
- [108] Yadav SP. A Study of Treatment Seeking Behaviour for Malaria and Its Management in Febrile Children in Rural Part of Desert, Rajasthan, India. *J Vector Borne Dis* 2010;47(4):235–42.