

TITLE

Parenthood and gene expression of oxytocin receptors and vasopressin receptors in sensory cortices of the male California mouse (*Peromyscus californicus*)

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ABSTRACT

The onset of parental care is associated with shifts in parents' perception of sensory stimuli from infants, mediated by neural plasticity in sensory systems. In new mothers, changes in auditory and olfactory processing have been linked to plasticity at several points along both sensory pathways, including cortical changes that are modulated, at least in part, by oxytocin. In males of biparental species, vasopressin, in addition to oxytocin, is important for modulating parental behavior; however, little is known about sensory plasticity in new fathers. We examined variation in the mRNA expression of oxytocin and vasopressin receptors (*Oxtr* and *Avpr1a*) in sensory cortices of virgin males, paired nonbreeding males, and new fathers in the biparental California mouse (*Peromyscus californicus*), and variation among cortices using the visual cortex for comparison. Reproductive status did not affect gene expression for either receptor, but compared to the visual cortex, expression of both receptors was higher in the left auditory cortex and lower in the anterior olfactory nucleus. Additionally, expression for both receptors was higher in the left auditory cortex compared to the right auditory cortex. While oxytocin and vasopressin receptor expression may remain stable across reproductive stages in male California mice, our findings provide support for auditory cortex lateralization, with the left auditory cortex possibly displaying higher sensitivity to both oxytocin and vasopressin compared to the right.

KEY WORDS

Oxytocin, vasopressin, parenthood, olfactory plasticity, auditory plasticity, cortex, California mouse

INTRODUCTION

Mammals that exhibit parental care experience dramatic shifts in their detection, perception, and responses to infant-related stimuli during the transition to parenthood (Horrell et al. 2019; Numan 2020; Wilson et al. 2023). The valence of infant stimuli, such as odors and vocalizations, changes from being aversive to attractive around the time of parturition (Fleming et al. 1993; Fleming et al. 2002; González-Mariscal and Poindron 2002; Lévy et al. 2004). For example, CBA/CaJ and NMRI house mouse (*Mus musculus*) mothers are more sensitive to and better able to discriminate pup vocalizations than virgin females (Galindo-Leon et al. 2009; Liu et al. 2006; Rothschild et al. 2013; Shepard et al. 2013), and NMRI males with paternal experience prefer tones that are a similar frequency to pup calls, compared to lower-frequency tones, while males without paternal experience show no preference (Ehret 2005; Ehret and Koch 1989). Similarly, in both Sprague-Dawley rats (*Rattus norvegicus*) and house mice, new mothers are more attracted to pup-related odors than are virgin females (Kinsley and Bridges 1990; Lévy et al. 2004; Lévy and Keller 2009), and regardless of mating status, male prairie voles (*Microtus ochrogaster*) that exhibit paternal behavior prefer pup odors to control odors (Yamoah et al. 2008). Some of this sensory plasticity is associated with changes in neuroendocrine signaling in the brain (Fleming et al. 1989; Ziegler and Sosa 2016), specifically, along sensory pathways (Miranda and Liu 2009; Wilson et al. 2023).

The neuropeptides oxytocin (OXT) and arginine vasopressin (AVP) facilitate the onset of parental behavior, largely through actions in integrative forebrain regions (Bales and Saltzman 2016; Horrell et al. 2019; Numan 2020; Saltzman and Ziegler 2014). For example, soon after the birth of their first litter, female rats have higher OXT receptor (*Oxtr*) mRNA expression in integrative regions important for parental care (medial preoptic area [MPOA] and bed nucleus of

the stria terminalis [BNST]) compared to virgin females (Meddle et al. 2007), and injections of either an AVP or OXT antagonist into the MPOA of female rats soon after parturition reduce parental care (Pedersen et al. 1994). Similar findings have been reported for males in species in which new fathers spontaneously care for offspring. In mandarin voles (*Microtus mandarinus*), injecting an OXT antagonist into the MPOA reduces paternal behavior in new fathers (Yuan et al. 2019). Additionally, in biparental California mice (*Peromyscus californicus*), fathers have lower mRNA expression for *Oxtr* and AVP 1a receptor (*Avpr1a*) in the BNST compared to virgin males (Perea-Rodriguez et al. 2015), and fathers display increased responsiveness to newborns following intranasal administration of OXT (Guoynes and Marler 2022). Similarly, in the facultatively biparental meadow (*Microtus pennsylvanicus*), AVP injection to the lateral ventricles increases parenting behavior in virgin males (Parker and Lee 2001).

Oxytocin can also modulate sensory plasticity during the transition to motherhood (Bester-Meredith et al. 2015; Numan 2020; Valtcheva and Froemke 2019; Wilson et al. 2023). Oxytocin receptors have been identified in the auditory (AC), piriform (Pir), visual (VC) and somatosensory cortices as well as in the anterior olfactory nucleus (AON) of mouse mothers, virgin females and virgin males (Mitre et al. 2016) and in sensory association areas of virgin male and female prairie voles (Duchemin et al. 2017). Primiparous female rats have higher *Oxtr* mRNA expression in the olfactory bulbs compared to virgin females (Meddle et al. 2007), and elevated OXT enhances maternal behavior in response to pup calls in mice (Banerjee and Lui 2013; Marlin et al. 2015; Yoshihara et al. 2018). Interestingly, Marlin et al. (2015) found that OXT infusion into the left auditory cortex (L-AC), but not the right (R-AC), reduced latency to retrieve pups in primiparous mouse mothers. AVP may also modulate sensory plasticity, since AVP 1a receptors have been identified in cortical regions of rats of both sexes including the

AON and Pir (Wacker and Ludwig 2019). However, it remains unclear whether expression of *Oxtr* and *Avpr1a* in sensory cortical regions changes during the transition to parenthood.

The distribution of *Oxtr* and *Avpr1a* in the sensory cortices of fathers relative to reproductively inexperienced males is, to our knowledge, unknown. However, evidence suggests that OXT and AVP can act in sensory pathways of males. Male C57BL/6 house mice have receptors for OXT and AVP in their vomeronasal organs (VNOs), and i.p. injection of OXT reduces VNO activity and pup-directed aggression (Nakahara et al. 2020). Additionally, i.p. injection of AVP raises auditory brainstem response thresholds in virgin male Wistar rats, indicating reduced ability to detect auditory stimuli (Naganuma et al. 2014).

In the present study, we examined the effects of fatherhood on *Oxtr* and *Avpr1a* mRNA expression in the auditory and olfactory cortices of male California mice, a monogamous, biparental rodent in which fathers provide extensive care for their offspring (Gubernick and Alberts 1987). The onset of parenthood in this species alters males' behavioral and neural (as measured by expression of Fos, the product of the immediate early gene *c-fos*) responses to pups as well as to isolated pup odors and vocalizations (Arquilla et al. 2023; de Jong et al. 2009; Wilson et al. 2022). Preliminary findings indicate that electrophysiological responses of the auditory cortex to pup vocalizations differ between fathers and virgin males (Deane, K.E., Saltzman, W., Razak, K.A., unpub). In addition, preliminary data suggest that treatment with an OXTR antagonist mildly inhibits parental care in California mouse fathers (Hussein, M., Unal, A., Saltzman, W., unpub. data), and fathers have lower levels of both *Oxtr* and *Avpr1a* in the BNST than virgin males (Perea-Rodriguez et al. 2015). However, effects of fatherhood on *Oxtr* and *Avpr1a* in brain regions associated with sensory processing have not been evaluated. Therefore, we quantified mRNA expression for both *Oxtr* and *Avpr1a* in four cortical regions

involved in acoustic (L-AC, R-AC) or olfactory (Pir and AON) processing, and as a control, in the visual cortex (VC). The VC was used as a control because rodents rely more heavily on olfactory and acoustic pathways for processing social cues, compared to visual pathways (Chen and Hong 2018), and previous work by Mitre et al. (2016) showed that the percent of VC cells containing *Oxtr* does not differ between mothers and virgins in C57BL/6 house mice. Because AVP and OXT impact the saliency of a range of social cues (Rigney et al 2022), two control groups (males paired with another male and males paired with a tubally ligated female) were employed in order to address potential effects not only of fatherhood but also of cohabitation with a female.

METHODS

Animal housing and care

California mice were bred at the University of California, Riverside (UCR) and were descendants of mice purchased from the Peromyscus Genetic Stock Center (University of South Carolina, Columbia, USA). All animals were housed in 44 × 24 × 20 cm polycarbonate cages with aspen shavings for bedding, cotton for nesting material, and ad libitum access to food (Purina 5001 Rodent Chow) and water. The lights were on a 14:10 h cycle with lights on at 2300 h. Ambient temperature was maintained at approximately 23° C, and humidity was around 65%. All procedures were approved by UCR's Institutional Animal Care and Use Committee and were conducted in accordance with the recommendations of the *Guide for the Care and Use of Laboratory Animals*. UCR is accredited by the Association for Assessment and Accreditation of Laboratory Animal Care.

Mice were housed with both parents until weaned at 27-31 days of age. They were then housed in single-sex groups with 1-3 other age-matched mice until they were used for this study. Male mice were assigned to three groups, each with 8 males: fathers, paired nonbreeders, and virgins. Fathers and paired nonbreeders were housed with an unrelated (less closely related than first cousins), age-matched female, and virgins were housed with a male from their original group of juveniles to reduce the potential of aggressive interactions between cage mates (Trainor and Marler 2001).

Prior to pair formation, females to be paired with fathers underwent sham tubal ligation (see below), and females to be paired with nonbreeding males underwent tubal ligation (see below). No surgeries were performed on males. In total, male subjects were from 16 different families, with no more than 3 males used from the same family. When siblings were used, they were assigned to different groups.

Surgeries and Pairing

Before being paired with a male, females were housed in same-sex groups of 2-4 mice until 75 - 91 days of age ($X \pm SD = 82.5 \pm 6.0$), at which time they underwent tubal ligation or sham tubal ligation following previously established protocols (Zhao et al. 2018). Briefly, females were anesthetized with 2.5% isoflurane vapor, a midline incision (approximately 1 cm) was made across the lower abdomen, and the fallopian tubes were located. For tubal ligation surgeries, each fallopian tube was tied in two places using absorbable sutures and cut between the ties. For sham tubal ligations, the fallopian tubes were left intact. The abdominal muscle layer was closed using absorbable sutures, and the skin was sealed using tissue glue. Lidocaine was applied topically at the site of the incision, and females were given s.c. buprenorphine (Hospira Inc., Lake Forest, IL,

138 USA) every 7-10 hours for 48 hours and Carprofen (Carprieve [Norbrook Laboratories;
139 Overland Park, KS, USA]) every 24 hours for 48 hours, with the first dose of both given
140 immediately before surgery.

141 Following surgery, females were allowed to recover in isolation for 7 days, reunited with
142 their original female cage mates for an additional 7 days, and then paired with a male mate. At
143 the time of pairing, males were between 88 and 138 days old and age did not differ between
144 groups (t-test $P > 0.35$; $X \pm SD$: fathers = 106 ± 6.4 ; nonbreeding = 114.4 ± 5.8).

145

146 **Brain collection**

147 Brains were collected from breeding males 2-3 days after the birth of their first litter and from
148 nonbreeders and virgins on the same day in an age-matched manner. The length of time mice
149 were paired prior to brain collection did not differ between breeding and nonbreeding pairs (t-test
150 $P < 0.67$, X and SD : breeding = 51.4 ± 7.2 ; nonbreeding = 55 ± 4.4). Brain collection from virgin
151 males occurred at a younger age than males paired with a female (ANOVA, model $P = 0.007$, F
152 = 6.5, post-hoc P 's < 0.04 . X and SD : virgin = 135.75 ± 6.7 ; fathers = 157.4 ± 7.1 ; nonbreeding
153 = 169.4 ± 6.7).

154 Each mouse was removed from its home cage between 0900 and 1000 h, placed into a
155 DecapiCone (Braintree Scientific; Braintree, MA, USA), and immediately decapitated using a
156 guillotine. The brain was then rapidly dissected from the skull. Following a previously described
157 protocol (Duchemin et al. 2017), the two cortical hemispheres were separated, flattened and
158 placed on dry ice. Cortical punches (1 mm diameter) were collected and pooled from the left and
159 right VC, Pir and AON. Punches were also collected from the right and left ACs but were kept

separate based on the possibility of lateralization (Marlin et al. 2015). All samples were stored at -80° C.

qPCR

Quantitative PCR was performed following previously established procedures (Laredo et al. 2014). In brief, we extracted RNA from each punch sample using Trizol (Fisher Scientific) and assessed RNA quality using spectrographic analyses on a Nanodrop. For each sample, 1 µg of RNA was used for reverse transcription using iScript (BioRad). We performed duplicate real-time PCR reactions for *Oxtr* (Genbank accession: MN265350.1, FisherSci Catalog number 43-320-78) and *Avpr1a* (Genbank accession: XM_052753487.1, FisherSci Catalog number 43-320-78), and 18s ribosomal RNA (FisherSci Catalog number: 43-108-93E) was used for a reference transcript. All samples were run using Taqman chemistry (FisherSci catalog number: 44-49-63) on an Applied Biosystems 7500 detection system (Applied Biosystems). For each plate, relative expression levels were calculated for each sample using the $\Delta\Delta CT$ method. To compare samples across plates, we made a pool of cDNA from each brain region. Samples of this pool was run for each transcript on every plate. For each sample, the expression value for each transcript was divided by the expression value from the pool.

Statistical Analyses

Linear mixed-effect models were used to determine whether receptor mRNA expression varied with male reproductive status (virgin, nonbreeding and father). All models included reproductive status as the main effect and male age as a covariate. Age was removed from models when it did not predict mRNA expression ($\alpha \leq 0.05$). Since not all variables could be transformed effectively

using the same method of transformation (see below), non-parametric Kruskal-Wallis tests were used to evaluate whether mRNA expression differed among cortices. . Significant results were further evaluated using Dunn's tests.

STATA 17 (StataCorp LP, College Station, TX, USA) was used for all analyses. Assumptions of normality were assessed using Shapiro-Wilk analyses and quantile-quantile plots. Data were either log-transformed (VC, Pir, L-AC and R-AC) or inverse square-root-transformed (AON) to meet assumptions of normality for LMMs. Within each receptor type for each brain region, transformed mRNA expression data were analyzed for outliers, which were considered to be values that were ≥ 1.5 interquartile ranges above or below the 75th and 25th quartiles, respectively. Across all data, 12 outlier values were identified and removed prior to analyses (VC *Oxtr*: n = 3; VC *Avpr1a*: n = 1; AON *Oxtr*: n = 5; AON *Avpr1a*: n = 3). For all tests, the critical P-value was set at 0.05 (two-tailed).

RESULTS

Expression of *Oxtr* and *Avpr1a* mRNA varied widely among individual mice within each cortical region, but this variation was not explained by male reproductive status. We found no differences in mRNA expression of *Oxtr* or *Avpr1a* among male California mice housed with a male (virgins), a tubally ligated female (nonbreeding males), or a sham-tubally ligated female and their first litter of pups (fathers) in any of the cortical regions examined (LMM, model *P*'s > 0.18; Table 1). The covariate of male age was non-significant and, thus, was removed from all models except for *Oxtr* expression in the AON (LMM, model *P* = 0.17, male reproductive status *P* = 0.18, male age *P* = 0.03; Table 1).

Expression of both *Oxtr* and *Avpr1a* mRNA differed significantly among cortical regions. Specifically, *Oxtr* mRNA expression was significantly lower in the AON compared to the Pir, VC and L-AC, and significantly higher in the L-AC compared to all other regions (Kruskal-Wallis test and Dunn's post-hoc test, Table 2, Fig. 1A). Similarly, expression of *Avpr1a* mRNA was significantly lower in the AON compared to all other regions, and significantly higher in the L-AC compared to all other regions (Kruskal-Wallis test and Dunn's post-hoc test, Table 2, Fig. 1B). Notably, both *Oxtr* and *Avpr1a* mRNA levels were significantly higher in the L-AC compared to the R-AC (paired t-tests, *Oxtr*: $P = 0.004$, *Avpr1a*: $P = 0.0007$, Fig. 1).

228 TABLES

229 Table 1. Expression of oxytocin (*Oxtr*) and vasopressin (*Avpr1a*) receptor mRNA in cortical
 230 brain regions of fathers, virgin males and nonbreeding males. Parental status did not impact
 231 mRNA expression in any region, and family identity did not contribute significantly to any
 232 model. Reported means and confidence intervals are for transformed data. LMMs, log-
 233 transformed: visual cortex [VC], piriform cortex Pir], left auditory cortex [L-AC] and right
 234 auditory cortex [R-AC]; inverse square root-transformed: anterior olfactory nucleus [AON].

Brain region	<i>Oxtr</i>					<i>Avpr1a</i>				
	Mean	95% CI	N	χ^2	P	Mean	95% CI	N	χ^2	P
VC			19	0.49	0.783			21	2.98	0.225
Virgin	1.83	1.10 to 2.57	6			3.62	2.31 to 4.93	7		
Nonbreeding	1.50	0.86 to 2.13	8			2.33	1.10 to 3.55	8		
Father	1.73	0.92 to 2.53	5			3.78	2.36 to 5.20	6		
Pir			23	0.76	0.684			23	1.42	0.491
Virgin	2.20	1.01 to 3.38	7			2.98	1.63 to 4.32	7		
Nonbreeding	2.48	1.37 to 3.59	8			3.33	2.07 to 4.59	8		
Father	1.78	0.67 to 2.89	8			2.26	1.01 to 3.52	8		
AON ^a			16	5.11	0.164			18	0.80	0.671
Virgin	0.84	0.70 to 0.99	6			0.54	0.44 to 0.65	7		
Nonbreeding	0.63	0.49 to 0.77	6			0.47	0.36 to 0.59	6		
Father	0.71	0.56 to 0.85	4			0.53	0.40 to 0.66	5		
L-AC			19	0.65	0.724			19	0.97	0.614
Virgin	3.45	2.02 to 4.87	7			5.81	4.27 to 7.34	7		
Nonbreeding	4.31	2.77 to 5.85	6			6.92	5.26 to 8.58	6		
Father	3.79	2.26 to 5.33	6			6.11	4.45 to 7.77	6		
R-AC			18	3.09	0.214			18	2.37	0.306
Virgin	0.72	-0.08 to 1.52	7			1.98	0.91 to 3.06	7		
Nonbreeding	1.52	0.72 to 2.32	7			2.90	1.83 to 3.98	7		
Father	1.78	0.72 to 2.84	4			3.25	1.83 to 4.67	4		

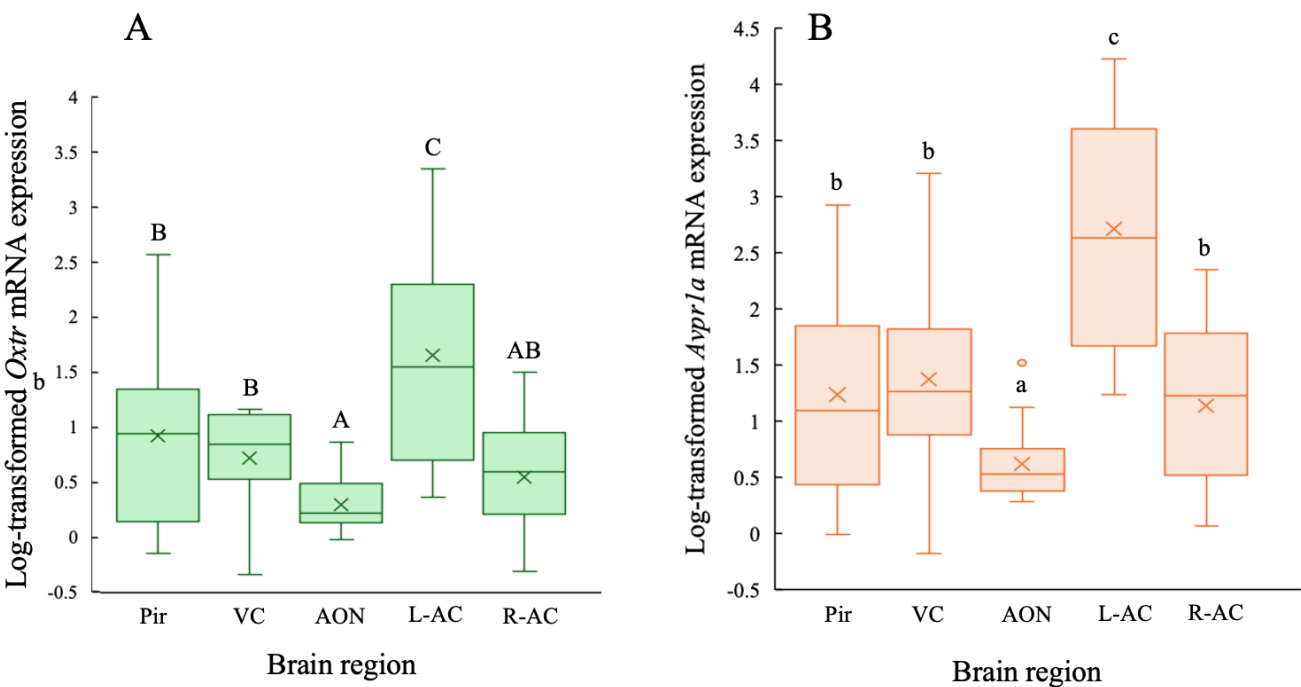
235 ^a Male age varied positively with *Oxtr* receptor mRNA ($z = 2.20$; $P = 0.028$).

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Table 2. Comparison of expression of oxytocin (*Oxtr*) and vasopressin (*Avpr1a*) receptor mRNA among sensory cortical regions of male California mice from all three reproductive groups combined. Kruskal-Wallis tests (top row of results) followed by post-hoc Dunn's tests. Pir – piriform cortex, VC – visual cortex, AON – anterior olfactory nucleus, L-AC– left auditory cortex, R-AC – right auditory cortex. P-values <0.05 are in bold.

Brain region	<i>Oxtr</i>			<i>Avpr1a</i>		
	N	χ^2	P	N	χ^2	P
		30.26	0.0001		38.35	0.0001
Pir versus:	23			23		
VC	19		0.252	21		0.295
AON	16		0.001	18		0.008
L-AC	19		0.007	19		< 0.0001
R-AC	18		0.056	18		0.414
VC versus:						
AON			0.009			0.002
L-AC			0.002			0.0004
R-AC			0.186			0.236
AON versus:						
L-AC			< 0.0001			< 0.0001
R-AC			0.0692			0.019
L-AC versus:						
R-AC			0.0001			< 0.0001



253

254 Fig. 1: Expression of (A) oxytocin (*Oxt*) and (B) vasopressin (*Avpr1a*) receptor mRNA in

255 sensory cortical brain regions of male California mice from all three reproductive groups

256 compared to the visual cortex. Box plots show median, 1st and 3rd quartiles. Error bars show

257 minimum and maximum values. Letters denote significant ($P < 0.05$) differences from post-hoc

258 Dunn's test following Kruskal-Wallis tests. Pir – piriform cortex, VC – visual cortex, Aon –

259 anterior olfactory nucleus, L-AC – left auditory cortex, R-AC – right auditory cortex.

DISCUSSION

The transition to parenthood can induce plasticity in sensory cortical regions in new mothers, which facilitates the expression of maternal care (Valtcheva and Froemke 2019). However, cortical plasticity in new fathers is poorly studied and, for both sexes, it is unclear whether plasticity involves changes in neuropeptide signaling in sensory brain regions. We found that neither *Oxtr* nor *Avpr1a* mRNA expression in the sensory cortices differed between males housed with another male and males housed with a nonbreeding female, or between either of these controls and fathers. Across reproductive groups, however, levels of both *Oxtr* and *Avpr1a* mRNA differed significantly across brain regions, with expression highest in the L-AC and lowest in the AON. These differences across regions may provide further insight into the role of different sensory cortices in modulating behavioral changes observed in new parents.

Previous studies have identified receptors for OXT and AVP in sensory cortices and sensory association areas of adult rodents (Duchemin et al. 2017; Mitre et al. 2016; Vaccari et al. 1998; Wacker and Ludwig 2019), and expression of OXTR and *Avpr1a* along sensory pathways can be modulated by early-life experience (Bester-Meredith and Marler 2001; Zeng et al. 2014). Our findings suggest that, unlike early-life events, the transition to parenthood in males does not induce plasticity in sensory cortices through variation in *Oxtr* or *Avpr1a* expression, which aligns with previous findings that OXTR labeling in sensory cortices did not differ between house mouse mothers and virgin females (Mitre et al. 2016).

On the other hand, changes in central concentrations of OXT and AVP might provide a mechanism through which sensory processing in the cortex is altered in parents. In some rodents, including the biparental prairie vole and Mandarin vole, the transition to fatherhood can result in increased synthesis of OXT and AVP in the paraventricular nucleus (PVN) and supraoptic

nucleus (SON) of the hypothalamus and of AVP in the bed nucleus of the stria terminalis and medial amygdala (Bales and Saltzman 2016; Song et al. 2010; Wang et al. 2000; Zimmermann-Peruzatto et al. 2015), which facilitates increased central release of OXT and AVP. Although *Avp* mRNA expression in the PVN and number of OXT- and AVP-stained neurons in the PVN and bed nucleus of the stria terminalis do not differ based on reproductive state in male California mice (De Jong et al. 2009, 2013), other potential sources of AVP (such as the SON) have not, to our knowledge, been explored. Thus, even though receptor expression may not change, the effects of OXT and AVP on synaptic activity in sensory cortices might be altered in fathers compared to virgin males.

This suggestion is consistent with the role of OXT in maternal cortical plasticity proposed by Valtcheva and Froemke (2019), whereby sensory inputs stimulate synthesis of OXT in, for example, oxytocin neurons in the PVN, which then modulates changes in sensory cortices that alter saliency of sensory stimuli from pups. Studies that demonstrate a connection between elevated levels of OXT and changes in cortical activity in females support the suggestion that cortical plasticity is driven by increased nonapeptide binding, and that the L-AC is specifically important for processing pup acoustic stimuli. Synaptic inhibition in the L-AC and Pir, as measured by whole-cell recordings from brain slices, was reduced for virgin female house mice in the presence of OXT compared to baseline inhibitory post-synaptic potentials (Mitre et al. 2016). Additionally, infusion of an OXT antagonist into the L-AC, but not the R-AC, of maternally experienced mice resulted in faster pup retrieval, and topical administration of OXT to virgin female mice resulted in neuronal responses to pup calls that were comparable to new mothers (Marlin et al. 2015). Interestingly, lateralization of *Oxtr* in the AC was observed in female mice, regardless of maternal status, but not males (Mitre et al. 2016). In our study,

biparental male California mice, regardless of paternal status, displayed the same lateralization of *Oxtr* and *Avpr1a* in their auditory cortices: males had higher expression of mRNA for both receptors in the L-AC compared to the R-AC. Thus, it is possible that the nonapeptides play a similar role in modulating responses to pup vocalizations for males of a biparental species and females of a uniparental species. Lateralization of *Oxtr* (and *Avpr1a*) in the auditory cortex could reflect increased receptor expression in L-AC within cell types that also express the receptors in R-AC. Alternatively or additionally, it is possible that some cell types express *Oxtr* (or *Avpr1a*) in L-AC but not in R-AC. These possibilities could be evaluated by using single-nucleus RNA sequencing.

We found that expression of both *Oxtr* and *Avpr1a* mRNA differed across sensory cortices in male California mice. The highest levels of receptor mRNA were found in the L-AC, while the lowest were found in the AON, which is important for olfactory memory and social behavior (Johnson and Young 2017; Oettl and Kelsch 2018). These findings suggest that the olfactory, auditory, and visual cortices may differ in the extent to which they are modulated by OXT and AVP. However, the functional significance of these differences remains to be determined. To our knowledge, the relative expression of OXT and AVP receptors, or the extent of cortical modulation by OXT and AVP, has not been compared systematically across sensory cortices in other species (but see Duchemin et al. 2017).

In conclusion, we found no evidence that either paternal status or cohabitation with a female influences expression of *Oxtr* and *Avpr1a* in sensory cortices of male California mice. It is possible, however, that fatherhood and/or cohabitation with a female alters receptor expression in subcortical regions of sensory pathways, or that *Oxtr* and/or *Avpr1a* expression undergoes transient changes during the onset of fatherhood or pair-bonding that were

not apparent at the time points used in our study. Further research into potential changes in OXT and AVP signaling within sensory systems will provide better context for the results reported here and may expand our understanding of the mechanisms by which these neuropeptides influence the onset of paternal care.

REFERENCES

Arquilla, A.M., Wilson, K.M., Razak, K.A., Saltzman, W. 2023. Fatherhood increases attraction to sensory stimuli from unrelated pups in male California mice, *Peromyscus californicus*. Anim. Behav. 198, 131-140. <https://doi.org/10.1016/j.anbehav.2023.02.001>

Bales, K. L., Saltzman, W. 2016. Fathering in rodents: Neurobiological substrates and consequences for offspring. Horm. Behav. 77, 249–259. <https://doi.org/10.1016/j.yhbeh.2015.05.021>

Banerjee, S.B., Liu, R.C. 2013. Storing maternal memories: hypothesizing an interaction of experience and estrogen on sensory cortical plasticity to learn infant cues. Front. Neuroendocrinol. 34, 300-314. <https://doi.org/10.1016/j.yfrne.2013.07.008>

Bester-Meredith, J. K., Marler, C. A. 2001. Vasopressin and aggression in cross-fostered California mice (*Peromyscus californicus*) and white-footed mice (*Peromyscus leucopus*). Horm. Behav. 40, 51–64. <https://doi.org/10.1006/hbeh.2001.1666>

Bester-Meredith, J. K., Fancher, A. P., Mammarella, G. E. 2015. Vasopressin proves essential: Vasopressin and the modulation of sensory processing in mammals. *Frontiers Endocrinol.* 6, 5. <https://doi.org/10.3389/fendo.2015.00005>

Charitidi, K., Meltser, I., Canlon, B. 2012. Estradiol treatment and hormonal fluctuations during the estrous cycle modulate the expression of estrogen receptors in the auditory system and the prepulse inhibition of acoustic startle response. *Endocrinology.* 153, 4412-4421. <https://doi.org/10.1210/en.2012-1416>

Chen, P., Hong, W. 2018. Neural circuit mechanisms of social behavior. *Neuron.* 98, 16–30. <https://doi.org/10.1016/j.neuron.2018.02.026>

de Jong, T. R., Chauke, M., Harris, B. N., Saltzman, W. 2009. From here to paternity: Neural correlates of the onset of paternal behavior in California mice (*Peromyscus californicus*). *Horm. Behav.* 56, 220–231. <https://doi.org/10.1016/j.yhbeh.2009.05.001>

de Jong, T. R., Harris, B. N., Perea-Rodriguez, J. P., Saltzman, W. 2013. Physiological and neuroendocrine responses to chronic variable stress in male California mice (*Peromyscus californicus*): Influence of social environment and paternal state. *Psychoneuroendocrinology.* 38, 2023–2033. <https://doi.org/10.1016/j.psyneuen.2013.03.006>

378 Duchemin, A., Seelke, A. M. H., Simmons, T. C., Freeman, S. M., Bales, K. L. 2017.
 379 Localization of oxytocin receptors in the prairie vole (*Microtus ochrogaster*) neocortex.
 380 Neuroscience. 348, 201–211. <https://doi.org/10.1016/j.neuroscience.2017.02.017>
 381
 382 Dumais, K. M., Veenema, A. H. 2016. Vasopressin and oxytocin receptor systems in the brain:
 383 Sex differences and sex-specific regulation of social behavior. Front. Neuroendocrinol. 40, 1–23.
 384 <https://doi.org/10.1016/j.yfrne.2015.04.003>
 385
 386 Ehret, G. 2005. Infant rodent ultrasounds – a gate to the understanding of sound communication.
 387 Behav. Genet. 3, 19–29. <https://doi.org/10.1007/s10519-004-0853-8>
 388
 389 Ehret, G., Koch, M. 1989. Ultrasound-induced parental behaviour in house mice is controlled by
 390 female sex hormones and parental experience. Ethology. 80, 81–93.
 391 <https://doi.org/10.1111/j.1439-0310.1989.tb00731.x>
 392
 393 Fleming, A. S., Cheung, U., Myhal, N., Kessler, Z. 1989. Effects of maternal hormones on
 394 ‘timidity’ and attraction to pup-related odors in female rats. Physiol. Behav. 46, 449–453.
 395 [https://doi.org/10.1016/0031-9384\(89\)90019-X](https://doi.org/10.1016/0031-9384(89)90019-X)
 396
 397 Fleming, A.S., Corter, C., Franks, P., Surbey, M., Schneider, B., Steiner, M. 1993. Postpartum
 398 factors related to mother’s attraction to newborn infant odors. Dev. Psychobiol. 26, 115 – 132.
 399 <https://doi.org/10.1002/dev.420260204>
 400

401 Fleming, A.S., Corter, C., Stallings, J., Steiner, M. 2002. Testosterone and prolactin are
 402 associated with emotional responses to infant cries in new fathers. *Horm. Behav.* 42, 399–413.
 403 doi:10.1006/hbeh.2002.1840.

404

405 Galindo-Leon, E.E., Lin, F.G., Liu, R.C. 2009. Inhibitory plasticity in a lateral band improves
 406 cortical detection of natural vocalizations. *Neuron* 62, 705-716.
 407 <https://doi.org/10.1016/j.neuron.2009.05.001>

408

409 González-Mariscal, G., Poindron, P., 2002. Parental care in mammals: immediate internal and
 410 sensory factors of control, in *Hormones, Brain and Behavior*. Academic Press. pp. 215-298.
 411 <https://doi.org/10.1016/B978-012532104-4/50005-6>

412 Gubernick, D.J., Alberts, J.R. 1987. The biparental care system of the California mouse,
 413 *Peromyscus californicus*. *J. Comp. Psychol.* 101, 169.

414

415 Guoynes, C. D., Marler, C. A. 2022. Intranasal oxytocin reduces pre-courtship aggression and
 416 increases paternal response in California mice (*Peromyscus californicus*). *Physiol. Behav.* 249,
 417 113773. <https://doi.org/10.1016/j.physbeh.2022.113773>

418

419 Horrell, N. D., Hickmott, P. W., Saltzman, W. 2019. Neural regulation of paternal behavior in
 420 mammals: Sensory, neuroendocrine, and experiential influences on the paternal brain, in:
 421 Coolen, L. M., Grattan D. R. (Eds.), *Neuroendocrine Regulation of Behavior*. Springer, Cham.
 422 pp. 111–160. https://doi.org/10.1007/7854_2018_55

423

424 Johnson, Z. V., Young, L. J. 2017. Oxytocin and vasopressin neural networks: Implications for
 425 social behavioral diversity and translational neuroscience. *Neurosci. Biobehav. Rev.* 76, 87–98.
 426 <https://doi.org/10.1016/j.neubiorev.2017.01.034>
 427
 428 Kinsley, C.H., Bridges, R.S. 1990. Morphine treatment and reproductive condition alter olfactory
 429 preferences for pup and adult male odors in female rats. *Dev. Psychobiol.* 23, 331-347.
 430 doi:10.1002/dev.420230405
 431
 432 Laredo, S. A., Orr, V. N., McMackin, M. Z., Trainor, B. C. 2014. The effects of exogenous
 433 melatonin and melatonin receptor blockade on aggression and estrogen-dependent gene
 434 expression in male California mice (*Peromyscus californicus*). *Physiol. Behav.* 128, 86–91.
 435 <https://doi.org/10.1016/j.physbeh.2014.01.039>
 436
 437 Lévy, F., Keller, M. 2009. Olfactory mediation of maternal behavior in selected mammalian
 438 species. *Behav. Brain Res.* 200, 336-345. <https://doi.org/10.1016/j.bbr.2008.12.017>
 439
 440 Lévy, F., Keller, M., Poindron, P. 2004. Olfactory regulation of maternal behavior in mammals.
 441 *Horm. Behav.* 46, 284-302. <https://doi.org/10.1016/j.yhbeh.2004.02.005>
 442
 443 Liu, R.C., Linden, J.F., Schreiner, C.E. 2006. Improved cortical entrainment to infant
 444 communication calls in mothers compared with virgin mice. *Eur. J. Neurosci.* 23, 3087-3097.
 445 <https://doi.org/10.1111/j.1460-9568.2006.04840.x>
 446

447 Marlin, B. J., Mitre, M., D'amour, J. A., Chao, M. V., Froemke, R. C. 2015. Oxytocin enables
 448 maternal behaviour by balancing cortical inhibition. *Nature*. 520, 499–504.
 449 <https://doi.org/10.1038/nature14402>
 450
 451 Meddle, S. L., Bishop, V. R., Gkoumassi, E., van Leeuwen, F. W., Douglas, A. J. 2007.
 452 Dynamic changes in oxytocin receptor expression and activation at parturition in the rat brain.
 453 *Endocrinology*. 148, 5095–5104. <https://doi.org/10.1210/en.2007-0615>
 454
 455 Miranda, J. A., Liu, R. C. 2009. Dissecting natural sensory plasticity: Hormones and experience
 456 in a maternal context. *Hear. Res.* 252, 21–28. <https://doi.org/10.1016/j.heares.2009.04.014>
 457
 458 Mitre, M., Marlin, B. J., Schiavo, J. K., Morina, E., Norden, S. E., Hackett, T. A., Aoki, C. J.,
 459 Chao, M. V., Froemke, R. C. 2016. A distributed network for social cognition enriched for
 460 oxytocin receptors. *J. Neurosci.* 36, 2517–2535. [https://doi.org/10.1523/JNEUROSCI.2409-](https://doi.org/10.1523/JNEUROSCI.2409-15.2016)
 461 [15.2016](https://doi.org/10.1523/JNEUROSCI.2409-15.2016)
 462
 463 Naganuma, H., Kawahara, K., Tokumasu, K., Satoh, R., Okamoto, M. 2014. Effects of arginine
 464 vasopressin on auditory brainstem response and cochlear morphology in rats. *Auris Nasus*
 465 *Larynx*. 41, 249–254. <https://doi.org/10.1016/j.anl.2013.12.004>
 466
 467 Nakahara, T.S., Camargo, A.P., Magalhães, P.H., Souza, M.A., Ribeiro, P.G., Martins-Netto,
 468 P.H., Carvalho, V.M., José, J., Papes, F. 2020. Peripheral oxytocin injection modulates

vomeroneasal sensory activity and reduces pup-directed aggression in male mice. *Sci. Rep.* 10, 19943. <https://doi.org/10.1038/s41598-020-77061-7>

Numan M. 2020. *The Parental Brain: Mechanisms, Development, and Evolution*. Oxford University Press.

Oettl, L.-L., Kelsch, W. 2018. Oxytocin and olfaction, in: Hurlemann, R., Grinevich, V. (Eds.), *Behavioral Pharmacology of Neuropeptides: Oxytocin*. *Current Topics in Behavioral Neurosciences* 35. Springer, Cham, pp. 55–75.

Parker, K. J., Lee, T. M. 2001. Central vasopressin administration regulates the onset of facultative paternal behavior in *Microtus pennsylvanicus* (meadow voles). *Horm. Behav.* 39, 285–294. <https://doi.org/10.1006/hbeh.2001.1655>

Pedersen, C. A., Caldwell, J. D., Walker, C., Ayers, G., Mason, G. A. 1994. Oxytocin activates the postpartum onset of rat maternal behavior in the ventral tegmental and medial preoptic areas. *Behav. Neurosci.* 108, 1163–1171. <https://doi.org/10.1037/0735-7044.108.6.1163>

Perea-Rodriguez, J. P., Takahashi, E. Y., Amador, T. M., Hao, R. C., Saltzman, W., Trainor, B. C. 2015. Effects of reproductive experience on central expression of progesterone, oestrogen α , oxytocin and vasopressin receptor mRNA in male California mice (*Peromyscus californicus*). *J. Neuroendocrinol.* 27, 245–252. <https://doi.org/10.1111/jne.12264>

492 Rigney, N., De Vries, G. J., Petrulis, A., Young, L. J. 2022. Oxytocin, vasopressin, and social
 493 behavior: from neural circuits to clinical opportunities. *Endocrinology*. 163, bqac111.
 494 <https://doi.org/10.1210/endo/bqac111>
 495
 496 Rothschild, G., Cohen, L., Mizrahi, A., Nelken, I. 2013. Elevated correlations in neuronal
 497 ensembles of mouse auditory cortex following parturition. *J. Neurosci.* 33, 12851-12861.
 498 <https://doi.org/10.1523/JNEUROSCI.4656-12.2013>
 499
 500 Saltzman, W., Ziegler, T. E. 2014. Functional significance of hormonal changes in mammalian
 501 fathers. *J. Neuroendocrinol.* 26, 685–696. <https://doi.org/10.1111/jne.12176>
 502
 503 Shepard, K.N., Kilgard, M.P., Liu, R.C. 2013. Experience-dependent plasticity and auditory
 504 cortex, in: Cohen, Y.E., Popper, A.N., Fay, R.R. (Eds). *Neural Correlates of Auditory Cognition*
 505 *Springer Handbook of Auditory Research*. Springer, Cham. pp. 293-327.
 506
 507 Song, Z., Tai, F., Yu, C., Wu, R., Zhang, X., Broders, H., He, F., Guo, R. 2010. Sexual or
 508 paternal experiences alter alloparental behavior and the central expression of ER α and OT in
 509 male mandarin voles (*Microtus mandarinus*). *Behav. Brain Res.* 214, 290–300.
 510 <https://doi.org/10.1016/j.bbr.2010.05.045>
 511
 512 Trainor, B. C., Marler, C. A. 2001. Testosterone, paternal behavior, and aggression in the
 513 monogamous California mouse (*Peromyscus californicus*). *Horm. Behav.* 40, 32–42.
 514 <https://doi.org/10.1006/hbeh.2001.1652>

515

516 Vaccari, C., Lolait, S. J., Ostrowski, N. L. 1998. Comparative distribution of vasopressin V1b
 517 and oxytocin receptor messenger ribonucleic acids in brain. *Endocrinology*. 139, 5015–5033.
 518 <https://doi.org/10.1210/endo.139.12.6382>

519

520 Valtcheva, S., Froemke, R. C. 2019. Neuromodulation of maternal circuits by oxytocin. *Cell*
 521 *Tissue Res.* 375, 57–68. <https://doi.org/10.1007/s00441-018-2883-1>

522

523 Wacker, D., Ludwig, M. 2019. The role of vasopressin in olfactory and visual processing. *Cell*
 524 *Tissue Res.* 375, 201–215. <https://doi.org/10.1007/s00441-018-2867-1>

525

526 Wang, Z. X., Liu, Y., Young, L. J., Insel, T. R. 2000. Hypothalamic vasopressin gene expression
 527 increases in both males and females postpartum in a biparental rodent. *J. Neuroendocrinol.* 12,
 528 111–120. <https://doi.org/10.1046/j.1365-2826.2000.00435.x>

529

530 Wilson, K. M., Arquilla, A. M., Rosales-Torres, K. M., Hussein, M., Chan, M. G., Razak, K. A.,
 531 Saltzman, W. 2022. Neural responses to pup calls and pup odors in California mouse fathers and
 532 virgin males. *Behav. Brain Res.* 434, 114024. <https://doi.org/10.1016/j.bbr.2022.114024>

533

534 Wilson, K. M., Arquilla, A. M., Saltzman, W. 2023. The parental umwelt: Effects of parenthood
 535 on sensory processing in rodents. *J. Neuroendocrinol.* 35, e13237.
 536 <https://doi.org/10.1111/jne.13237>

537

538 Yamoah, D., Williams-Baginski, K., Bamshad, M. 2008. Changes in response to odors during
 539 the reproductive period in male and female prairie voles (*Microtus ochrogaster*). Can. J. Zool.
 540 86, 224–230. <https://doi.org/10.1139/Z07-133>
 541
 542 Yoshihara, C., Numan, M. and Kuroda, K.O., 2018. Oxytocin and parental behaviors, in
 543 Hurlemann, R., Grinevich, V. (Eds.), Behavioral Pharmacology of Neuropeptides: Oxytocin.
 544 Current Topics in Behavioral Neurosciences 35. Springer, Cham., pp.119-153.
 545 https://doi.org/10.1007/7854_2017_11
 546 Yuan, W., He, Z., Hou, W., Wang, L., Li, L., Zhang, J., Yang, Y., Jia, R., Qiao, H., Tai, F. 2019.
 547 Role of oxytocin in the medial preoptic area (MPOA) in the modulation of paternal behavior in
 548 mandarin voles. Horm. Behav. 110, 46–55. <https://doi.org/10.1016/j.yhbeh.2019.02.014>
 549
 550 M. Zhao, T. Garland Jr., M.A. Chappell, J.R. Andrew, B.N. Harris, W. Saltzman. 2018. Effects
 551 of a physical and energetic challenge on male California mice (*Peromyscus californicus*):
 552 modulation by reproductive condition, J. Exp. Biol. 221, jeb168559.
 553 <https://doi.org/10.1242/jeb.168559>.
 554
 555 Zheng, J.J., Li, S.J., Zhang, X.D., Miao, W.Y., Zhang, D., Yao, H., Yu, X. 2014. Oxytocin
 556 mediates early experience–dependent cross-modal plasticity in the sensory cortices. Nat.
 557 Neurosci, 17, 391–399. <https://doi.org/10.1038/nn.3634>
 558

559 Ziegler, T.E., Sosa, M.E. 2016. Hormonal stimulation and paternal experience influence
560 responsiveness to infant distress vocalizations by adult male common marmosets, *Callithrix*
561 *jacchus*. Horm. Behav. 78, 13-19. <https://doi.org/10.1016/j.yhbeh.2015.10.004>
562
563 Zimmermann-Peruzatto, J. M., Lazzari, V. M., de Moura, A. C., Almeida, S., Giovenardi, M.
564 2015. Examining the role of vasopressin in the modulation of parental and sexual behaviors.
565 Front. Psychiatry. 6, 130. <https://doi.org/10.3389/fpsyt.2015.00130>
566