

1 **SpudDB: A database for accessing potato genomic data**

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15 **Abstract**

16  
17 Potato is a key food crop with a complex, polyploid genome. Advancements in sequencing technologies  
18 coupled with improvements in genome assembly algorithms have enabled generation of phased,  
19 chromosome-scale genome assemblies for cultivated tetraploid potato. The SpudDB database houses  
20 potato genome sequence and annotation, with the doubled monoploid DM 1-3 516 R44 (hereafter DM)  
21 genome serving as the reference genome and haplotype. Diverse annotation data types for DM genes  
22 are provided through a suite of Gene Report Pages including gene expression profiles across 438 potato  
23 samples. To further annotate potato genes based on expression, 65 gene co-expression modules were  
24 constructed that permit identification of tightly co-regulated genes within DM across development and  
25 responses to wounding, abiotic stress, and biotic stress. Genome browser views of DM and 28 other  
26 potato genomes are provided along with a download page for genome sequence and annotation. To link  
27 syntenic genes within and between haplotypes, syntelogs were identified across 25 cultivated potato  
28 genomes. Through access to potato genome sequences and associated annotations, SpudDB can enable  
29 potato biologists, geneticists and breeders to continue to improve this key food crop.

30 **Introduction**

31  
32 *Solanum* Sect Petota contains approximately 100 species including *Solanum tuberosum* L. (cultivated  
33 potato) (Spooner, 2009). Potatoes were domesticated from wild potato species approximately 8,000-  
34 10,000 years ago in the Andes (Spooner et al., 2005) and have since spread throughout the world  
35 serving as a critical food crop. Potato tubers are modified underground stem structures and are clonally  
36 derived. While tubers serve as a mode of asexual reproduction and as a mechanism to overwinter and  
37 evade predation, the reliance on clonal propagation results in high genetic load due to the lack of a  
38 meiotic sieve to remove deleterious and dysfunctional alleles.

39  
40 Most cultivated potato cultivars are autotetraploids ( $2n = 4x = 48$ ) and due to its complex  
41 genome, the first potato genome sequenced was that of *S. tuberosum* Group Phureja DM 1-3 516 R44  
42 (hereafter DM), a doubled monoploid derived from a diploid clone via anther culture (Paz & Veilleux,  
43 1999). Its homozygosity permitted assembly in 2011 of the DM genome via *de novo* assembly of short-  
44 read sequences prior to development of third generation long read sequencing platforms (Potato  
45 Genome Sequencing Consortium et al., 2011). Access to the DM reference genome sequence, albeit a  
46 single haplotype, permitted an explosion of genome-enabled discoveries in potato including  
47 development of SNP-chip genotyping arrays (Felcher et al., 2012; Vos et al., 2015), assessment of potato  
48 structural genome diversity (Hardigan et al., 2015, 2016), understanding genes underlying domestication  
49 (Hardigan et al., 2017), discovery of genes associated with agronomic traits (e.g., (Sharma et al., 2018;  
50 Klaassen et al., 2019; Khlestkin et al., 2020; Prodhomme et al., 2020)), and furthering our knowledge of  
51 potato biology (e.g. (Peterson et al., 2016; Ye et al., 2018; Enciso-Rodriguez et al., 2019; Laimbeer et al.,  
52 2020; Zhang et al., 2020; Eggers et al., 2021; Ma et al., 2021; Ramírez Gonzales et al., 2021)). With  
53 respect to the genome landscape of cultivated tetraploids, whole genome resequencing coupled with  
54 alignments to the DM reference genome revealed a high degree of heterozygosity coupled with  
55 rampant structural variation attributable to mutation and wild species introgressions (Hardigan et al.,  
56 2017).

57  
58 In the last 15 years, the advances in sequencing technologies, enhanced genome assembly  
59 algorithms, and increased computing capacities have resulted in generation of multiple potato genome  
60 sequences, including heterozygous diploid genomes ( $2n = 2x = 24$ ) as well as phased, tetraploid genomes.  
61 Some of the first cultivated potato genomes available subsequent to DM were diploid or dihaploid ( $2n =$   
62  $2x = 24$ ) genomes derived from cultivated tetraploids (van Lieshout et al., 2020; Zhou et al., 2020;  
63 Jayakody et al., 2023). With access to long-read sequencing platforms and improved algorithms that can  
64 phase genome assemblies in the last few years, phased tetraploid genomes are now feasible (Hoopes et  
65 al., 2022; Sun et al., 2022; Bao et al., 2023) confirming earlier estimates of genome heterogeneity, allelic  
66 diversity, structural variation, wild species introgressions, and a high degree of dysfunctional and  
67 deleterious alleles that were derived from short-read sequencing alignments to the DM reference  
68 genome. While most potato genome sequences have been generated from cultivated potato clones, a  
69 subset of wild potato species (Leisner et al., 2018; Yan et al., 2021; Tang et al., 2022; Feng et al., 2024;  
70 Hosaka et al., 2024) have been sequenced. Central to these emerging genome sequences was updating  
71 the DM reference genome to a chromosome-scale using long-read sequences coupled with Hi-C data  
72 that was re-annotated using RNA-sequencing (RNA-seq) and full-length cDNA sequences, greatly  
73 improving the quality of the genome sequence and the gene annotation (Pham et al., 2020).

74  
75 Genome and genetic data for potato are housed in a limited number of databases. The PoMaMo  
76 database included molecular maps, genome sequences, and suite tools (Meyer et al., 2005); however, it  
77 is no longer available. A subset of potato genomic data is available on the Solanaceae Genomics

78 Network (Fernandez-Pozo et al., 2015), yet this database is highly focused on tomato. Potato genome  
79 sequences are also available at the SpudDB (Hirsch et al., 2014) which was created for breeders to mine  
80 genotype and phenotype diversity data primarily derived from North American cultivated potato as part  
81 of the USDA-funded SolCAP project (Douches et al., 2014). With the recent availability of extensive  
82 chromosome-scale genome assemblies, we have updated SpudDB with new content, features, and  
83 access tools.

84

## 85 **Overview and Navigating SpudDB**

86

87 SpudDB provides access to potato genome sequences via genome browsers, search tools, download  
88 pages, and diverse annotation data types for the DM v6.1 reference genome. The home page of SpudDB  
89 (<https://spuddb.uga.edu/index.shtml>) highlights recent updates, a summary of the content of the  
90 database including links to literature associated with large datasets in SpudDB, and a quick search tool  
91 for DM v6.1 genes either by gene identifier or keyword. The menu provides access to the JBrowse2  
92 genome browsers (Diesh et al., 2023), database search tools, dataset download pages, and the results of  
93 various analyses such as gene expression. Archived updates of SpudDB are available on the What's New  
94 page (<https://spuddb.uga.edu/new.shtml>).

95

96 To facilitate access to potato genome sequences, we have deployed a JBrowse2 genome browser  
97 (Diesh et al., 2023) hosting 29 total genomes including the DM v6.1 reference genome (Pham et al.,  
98 2020), phased diploid breeding line RH 89-039-16 (Zhou et al., 2020), phased tetraploid genomes  
99 (Hoopes et al., 2022; Sun et al., 2022; Bao et al., 2023), *S. chacoense* M6--a source of self-compatibility  
100 (Jansky et al., 2011; Eggers et al., 2021; Ma et al., 2021), and *S. candalleanum*, the progenitor of  
101 cultivated potato (Spooner et al., 2005). A suite of search tools for the DM v6.1 gene annotation is  
102 located on the Search Tool Page ([https://spuddb.uga.edu/integrated\\_searches.shtml](https://spuddb.uga.edu/integrated_searches.shtml)) including a BLAST  
103 (v2.2.26) (Altschul et al., 1997) search tool, functional annotation keyword search tool, InterPro  
104 identifier and key word search tool, Gene Ontology identifier and key word search tool, Pfam accession  
105 search tool, and a sequence identifier search tool. On the top menu bar is the Analyses tab which  
106 provides links to DM specific analyses such as gene expression, gene co-expression, and potato  
107 syntelogs. A Contact tab is also present to permit users to send feedback. A Download tab on the top  
108 menu bar contains links to webpages that describe available genome datasets including:

- 109 • DM v6.1 genome assembly, annotation, gene expression matrix, and variant calls
- 110 • Eight phased tetraploid genome assemblies and annotation (cv. Altus, Atlantic, Avenger, Castle  
111 Russet, Colomba, Cooperation-88, Otava, Sputna)
- 112 • Twenty phased dihaploid genome assemblies and annotation from the Potato 2.0 project
- 113 • Diploid RH89-039-16 (v3) genome assembly and annotation
- 114 • Doubled monoploid DM1S1 genome assembly, annotation, and variant calls
- 115 • Updated *S. chacoense* M6 (v5.0) genome assembly and annotation
- 116 • *S. candalleanum* (v1.0) genome assembly and annotation
- 117 • Archived *S. chacoense* M6 (v4.1) genome assembly and annotation
- 118 • Archived DM (PGSC v4.03/v4.04) genome assembly and annotation
- 119 • Tomato (*Solanum lycopersicum*) M82 (SollycM82\_v1) genome assembly and annotation

120

121 In addition, links to the genome browser for each of these assemblies are provided via its download  
122 webpage.

123

## 124 **DM v6.1 as the reference genome for potato**

125  
126 The DM v6.1 reference genome serves as the foundation for the potato community. Not only is it a high-  
127 quality, chromosome-scale genome assembly (Pham et al., 2020), it represents a single haplotype that  
128 serves to link alternative haplotypes present in highly heterozygous, phased diploid and tetraploid  
129 genome assemblies thereby linking alleles and syntelogs. The 741 Mb DM v6.1 genome was scaffolded  
130 into 12 chromosomes and annotated using extensive transcript evidence, including full-length cDNAs,  
131 resulting in 40,652 working protein-coding genes encoding 52,953 gene models and 32,917 high-  
132 confidence protein-coding genes encoding 44,851 gene models (Pham et al., 2020). The DM genes have  
133 been annotated for a suite of annotation data types to aid in understanding gene function which are  
134 available on individual Gene Report Pages for each gene. These include putative functional description  
135 assigned through BLAST searches against the *Arabidopsis thaliana* proteome, Swiss-Prot plant proteins,  
136 and the Pfam database as well as gene expression abundances. Further annotations include Gene  
137 Ontology classifications, BLAST searches against UniRef database, gene co-expression module  
138 assignment, and syntelogs across cultivated potato genomes.  
139

140 The major update to SpudDB was expansion of gene expression profiles to include additional  
141 RNA-sequencing (RNA-seq) datasets from a broader group of developmental stages, tissues, and  
142 treatments. To obtain relevant expression datasets, the National Center for Biotechnology Information  
143 Sequence Read Archive (Sayers et al., 2022) was queried for *S. tuberosum*. Initial filtering for paired end  
144 RNA-seq datasets resulted in 4,571 Sequence Read Archive accessions. These were then filtered for  
145 sequencing platform requiring the Illumina platform, RNA-seq library, minimum of 20 million reads,  
146 paired end sequences, and informative sample description. A subset of 456 accessions were  
147 downloaded and quality checked using FastQC (v0.12.1; (Wingett SW, 2018)) and MultiQC (Ewels et al.,  
148 2016) using default parameters and were then classified based on organ and treatment/conditions  
149 [Organ: fruit, flower, leaf, root, seedling, stem, tuber; Treatment/Conditions: abiotic stress, biotic stress,  
150 development, photoperiod, wounding] based on the BioProject and BioSample description. Expression  
151 abundances were calculated using kallisto quant (Bray et al., 2016) with the parameter -t 8 and  
152 represented as transcripts per million (TPMs). All of the RNA-seq samples downloaded for the gene  
153 expression analysis were clustered to identify mis-labeled samples and 18 accessions were removed  
154 based on their PCC or PCA clustering generated using the R commands: prcomp (R Core Team, 2023)  
155 with default parameters and cor with method option set to pearson, respectively, with aberrant tissue  
156 types. The final RNA-seq dataset has 438 samples. Of the 40,652 DM genes, 39,651 are expressed at  $\geq 0$   
157 TPM in at least one sample. Expression abundances are available for the entire DM genome via the Gene  
158 Expression page (<https://spuddb.uga.edu/expression.shtml>) or individually for each gene via the Gene  
159 Report Page.  
160

## 161 **Gene co-expression**

162  
163 Gene co-expression network modules were generated from all representative working gene models  
164 using Simple Tidy GeneCoEx with default parameters ([Li et al., 2023](#)). Co-expression modules were built  
165 using all 438 RNA-seq libraries (156 samples after replicate averaging) representing a diverse set of  
166 tissues and conditions/treatments including a tuber developmental series, abiotic stress, biotic stress,  
167 and a set of photoperiod conditions (Fig. 1a). To build the correlation network edge table, only edges  
168 with  $r > 0.8$  were used, which corresponded to the top 1% of all edges. A network object was then  
169 constructed using the 'graph\_from\_data\_frame()' function of igraph (Csárdi et al.) with option directed  
170 set to F. Graph based clustering was performed using the Leiden algorithm (implemented as the  
171 'cluster\_leiden()' function in R as part of the igraph package(Csárdi et al.) with a resolution parameter of  
172 4 and objective\_function parameter set to modularity. Of the 40,652 DM genes, 36,025 were placed into

173 co-expression modules based on their expression pattern generating 65 modules containing between 5  
174 and 2,943 genes.

175  
176 The co-expression modules can then be used to identify genes with expression patterns  
177 associated with specific tissues or treatments. Tuber bulking relies on the accumulation of amylopectin  
178 in the amyloplasts that is catalyzed by starch synthases (Nazarian-Firouzabadi & Visser, 2017). Starch  
179 synthase V (Soltu.DM.02G027020.1) was previously identified as vital to tuber bulking (Li et al., 2024). In  
180 the co-expression analyses, starch synthase V is a member of Module 6 which has peak expression in the  
181 tuber short day time course sample collected at 3pm. The expression of starch synthase V was plotted in  
182 red along with the other genes in module 6 showing high expression during later stages of tuber  
183 development and the time course data sets (Fig. 1b). This expression profile is expected based on the  
184 activity of starch synthase V in amylopectin accumulation.

185  
186 Co-expression modules can also be mined to identify additional genes involved in biological  
187 processes. For example, Module 11 has peak expression in wounded tubers after 3 days and generally  
188 high expression in all the wounded tubers after 1 day (Fig. 1a). Of the highly expressed genes, there was  
189 one MYB transcription factor. MYB transcription factors are known to be involved in wound healing  
190 through their regulation of suberin biosynthesis (Han et al., 2024). This uncharacterized MYB  
191 transcription factor (Soltu.DM.04G025530.1) exhibits an expression pattern with high expression after  
192 14 days of wounding and could play an important role in wound healing in potato (Fig. 1c). Co-  
193 expression module membership for the entire DM genome can be obtained via the Gene Co-expression  
194 page (<https://spuddb.uga.edu/coexpression.shtml>) and the Gene Report page for each individual gene.

195  
196 **Potato genome sequences and linking across haplotypes**

197 To facilitate traversing between alleles within and between genome assemblies of cultivated potato, we  
198 determined syntenic relationships between 25 cultivated potato genomes. Using DM as the reference  
199 genome, the representative gene models from four phased tetraploid genomes (Hoopes et al., 2022;  
200 Sun et al., 2022; Bao et al., 2023) and 20 phased dihaploid genomes from the Potato 2.0 project  
201 (<https://potatov2.github.io/>) were input into GENESPACE (Lovell et al., 2022) and syntelogs for each DM  
202 gene were identified. To account for the ploidy of the phased genomes, the ploidy parameter of the  
203 'init\_genespace' function was set to "1" for the DM v6.1 genome, "2" for the 20 dihaploid genomes,  
204 and "4" for the four tetraploid genomes. Syntelogs for each gene are viewable on the Gene Report  
205 Page and as a track on the DM v6.1 JBrowse.

206  
207 **DM v6.1 JBrowse**

208 The DM v6.1 reference genome is available as a JBrowse2 instance (Diesh et al., 2023)(Fig. 2). Tracks  
209 available include reference sequence, loci and gene models with separate tracks for representative high  
210 confidence gene models, high confidence gene models, and working gene models. Gene expression data  
211 is available as RNA-seq coverage tracks of the 438 RNA-seq samples that were generated using HISAT2  
212 (Kim et al., 2019) that are grouped based on classification. Syntelogs from GENESPACE (v1.3.1; (Lovell et  
213 al., 2022) are provided as well. For variant data, SNPs from genotyping-by-sequencing using a set of  
214 57,054 SureSelect baits (Uitdeewilligen et al., 2013) and the SolCAP SNP project that utilized RNA-seq and  
215 draft genome sequence to develop an Affymetrix SNP array platform (Hamilton et al., 2011) are  
216 provided to link positions in the DM genome with widely used genetic markers which are in use in  
217 community-based genotyping platforms. Individual JBrowse instances are also available for 28 other  
218 potato genomes [Tetraploid cultivars: Atlantic, Castle Russet, Cooperation-88, Otava; Diploid/Dihaploids:  
219 RH and 20 dihaploids from the Potato 2.0 project (<https://potatov2.github.io/>), Doubled Monoploid:  
220 DM1S1; Wild species: *S. chacoense* M6, *S. candolleanum*] and available via the top menu bar or via links

221 on their individual Download page.  
222

## 223 **SpudDB Gene Report Pages**

225 For a biologist, access to an array of annotation data types can facilitate understanding gene function.  
226 For each DM gene model, a Gene Report Page is available either through a search via gene identifier,  
227 key word identifier, or from a locus or gene model link in the JBrowse. The SpudDB Gene Report page  
228 (Fig. 3) has a summary of each gene model including putative function description, locus name, and  
229 alternative splice form (gene model). Basic metrics for each gene model include chromosome or scaffold  
230 location, coordinates on the DM v6.1 genome for the mRNA (predicted transcript = gene model), coding  
231 sequence length, and predicted amino acid length. The sequences of the genomic sequence, transcript  
232 sequence, coding sequence, and predicted protein sequence are provided in FASTA format. A link to the  
233 DM v6.1 JBrowse genome browser for each gene model is also included in the Gene Report Page. Gene  
234 ontology classifications are also provided as are searches against UniRef100. To facilitate development  
235 of targeted simple molecular markers, putative Simple Sequence Repeats (SSRs) for the locus are  
236 available with their coordinates.

237  
238 A table of gene expression abundances for each gene model is provided along with its run  
239 identifier from National Center for Biotechnology Information Sequence Read Archive, short description  
240 of the sample, classification of the study, and expression abundance in TPMs. Co-expression module  
241 assignment is also listed with the co-expression module membership and the module peak expression  
242 assignment. To facilitate traversing from DM to other potato genomes, syntelogs from the potato  
243 syntelog analysis are listed for each gene.

244  
245 **Improvements in architecture**  
246 SpudDB has undergone a number of back-end improvements and enhancements since the last release  
247 to support future updates and to continue to provide useful tools to the user community. The entire  
248 SpudDB website has been converted to use HTTPS for increased compatibility and security. The search  
249 tools and Gene Report pages have been migrated from a PostgreSQL instance to SQLite for increased  
250 performance and reliability. The number of genome browsers on SpudDB had grown from the original  
251 MySQL based Gbrowse1 for the legacy DM annotation to include a number of JBrowse1 and JBrowse2  
252 instances for the new and updated genomes added to SpudDB. All of these have been replaced by a  
253 single, unified JBrowse2 instance which is easier to maintain and provides an enhanced user experience.

254  
255 **Future Directions**  
256 As a substantial number of potato researchers are geneticists and breeders, SpudDB serves a key  
257 function in provision of genomic data not only from the DM reference genome but also from new  
258 emerging genome assemblies. We anticipate that more potato genome sequences and annotation will  
259 continue to be generated and made available to the public in the coming years. The back-end  
260 improvements to SpudDB, especially the use of JBrowse 2, will enable streamlined addition of new  
261 genomes to SpudDB. These assemblies, and importantly, the alleles in these genomes can be linked  
262 through synteny with new GENESPACE builds with the addition of new genomes to SpudDB. Continued  
263 development of new functional annotation datatypes will facilitate data-mining the potato genome and  
264 can readily be added to the Gene Report page for each gene. We also anticipate that development of a  
265 pan-genome for potato that captures novel structural variation in cultivated potato will provide new  
266 resources for potato researchers.

267  
268 **Data availability**

269 All data are freely available at SpudDB (<https://spuddb.uga.edu>) for searching and download. We have  
270 also deposited the new gene expression, gene co-expression, and syntelog datasets in Figshare under  
271 <https://doi.org/10.6084/m9.figshare.27471918.v1>.

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277

278 **Conflicts of interest**  
279 The authors declare no conflict of interest.  
280

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451

452 **Figures**

453

454 **Fig. 1. Gene Co-expression Module Expression.** a) Module expression of 438 RNA-sequencing libraries  
455 representing 18 conditions/treatments over seven tissue types. b) Expression of starch synthase V  
456 (Soltu.DM.02G027020). Z-score expression of all genes Module 6 is plotted in grey with starch synthase  
457 V in red. c) Expression of a MYB transcription factor (Soltu.DM.04G025530). Z-score expression of all  
458 genes in Module 11 is plotted in grey with the MYB transcription factor in red.

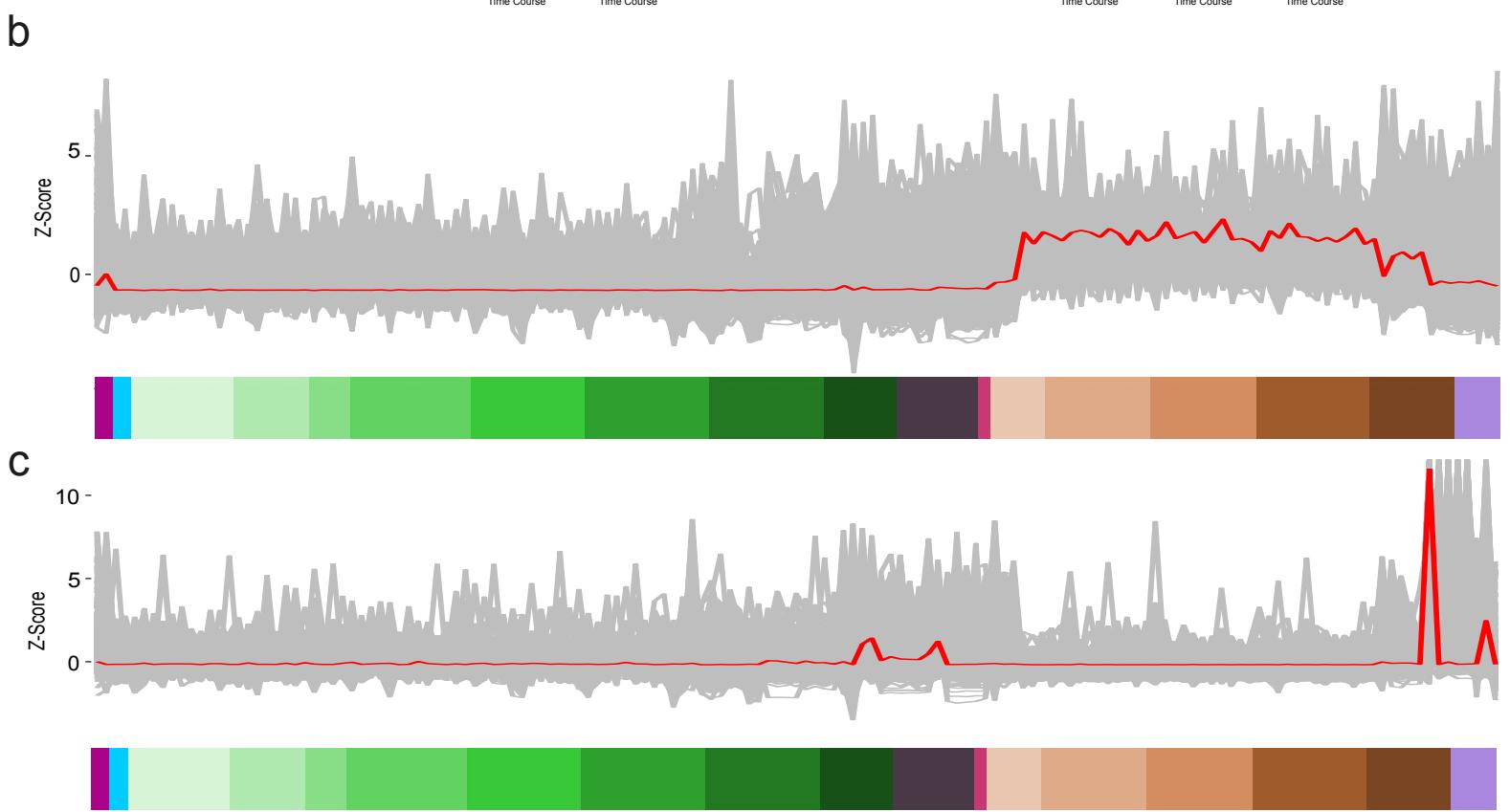
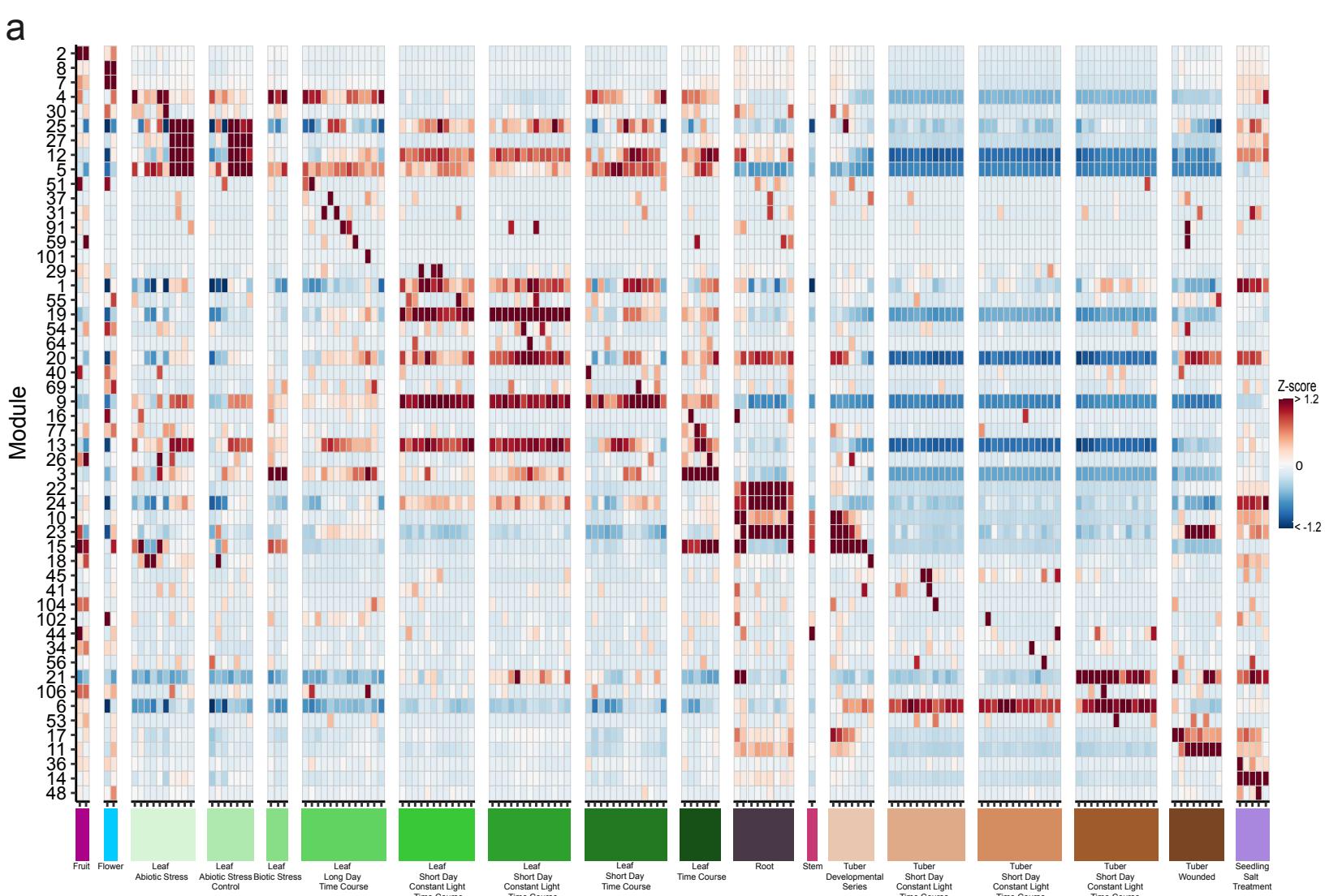
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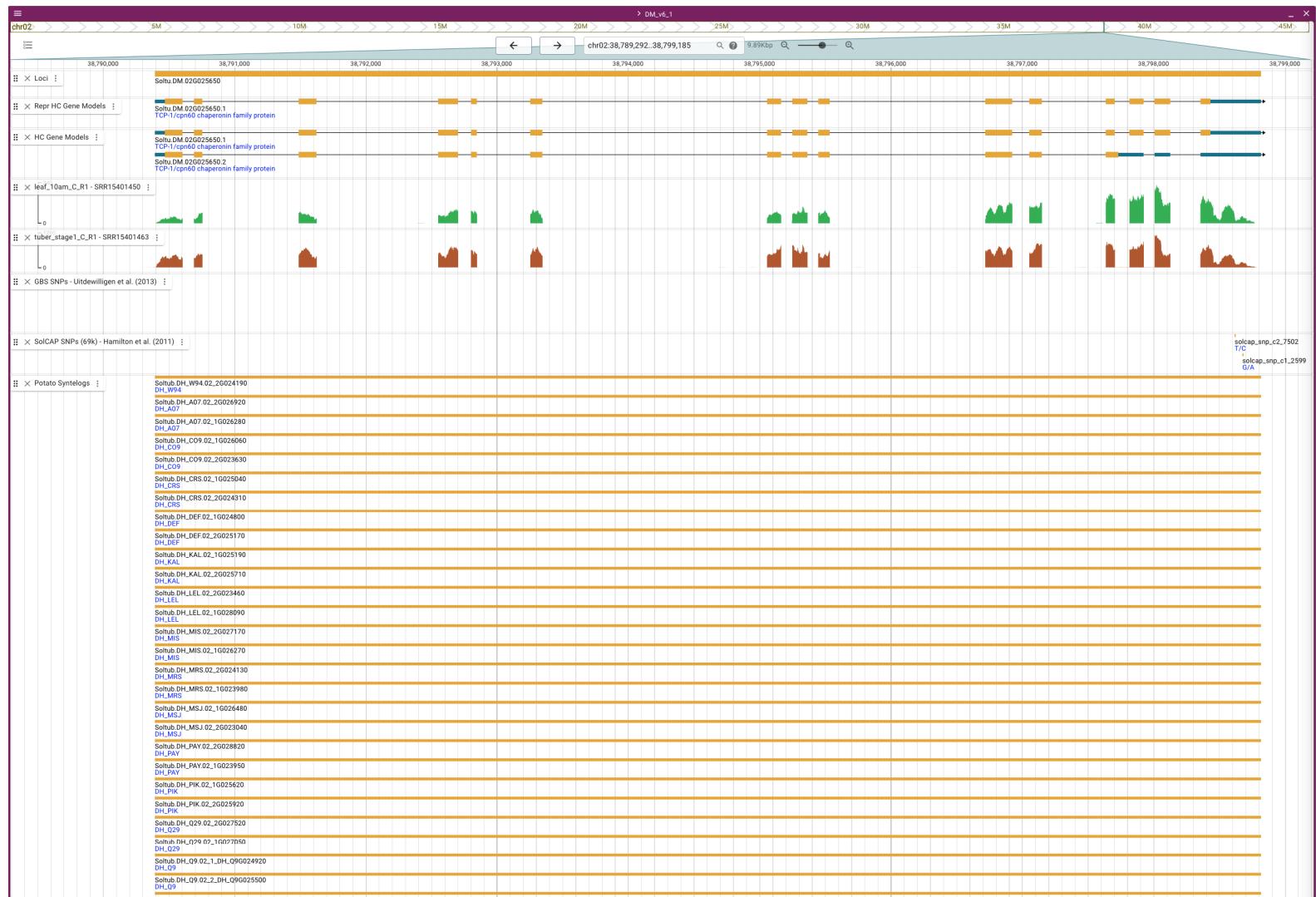
460 **Fig. 2. SpudDB genome browser.** Exemplar JBrowse2 screenshot of the DM v6.1 browser featuring  
461 Soltu.DM.02G025650 which encodes a TCP-1/cpn60 chaperonin family protein. The locus, representative high  
462 confidence gene model (Soltu.DM.02G025650.1), all high confidence gene models, two RNA-sequencing read  
463 alignments (leaf, 10 AM, Rep1 and tuber, stage 1, Rep1), variants from genotyping-by-sequencing and the SolCAP  
464 SNP array, and syntelogs within cultivated potato identified by GENESPACE are shown.

465

466 **Fig. 3. Gene Report Page features in SpudDB.** a) General information regarding Soltu.DM.02G025650.1 including  
467 link to the gene model in the genome browser, putative function, locus name, alternative splice form, and gene  
468 attributes (chromosome, coordinates, CDS length, and amino acid length). b) Gene ontology information. c) BlastP  
469 search of UniRef100 showing accession identifier, percent similarity, percent coverage, description, and p-value.  
470 d) Pfam and InterPro matches including accession identifier, method, name, match positions, and e-value. e)  
471 Potato syntelogs identified through GENESPACE. F) Coexpression module assignment including peak expression  
472 within the module. g) RNA-seq gene expression values in transcripts per million with National Center for

473 Biotechnology Information Sequence Read Archive accession identifier and sample description.  
474





a <a href="#">Soltu.DM.02G025650.1</a>		DM v6.1 Annotation		
<a href="#">Show Soltu.DM.02G025650.1 in the Spud DB Genome Browser</a>				
<b>Gene Identification</b>				
Putative Function: TCP-1/cpn60 chaperonin family protein				
Locus Name: <a href="#">Soltu.DM.02G025650</a>				
Alternative Splice Form: <a href="#">Soltu.DM.02G025650.2</a>				
<b>Gene Attributes</b>				
Scaffold: chr02				
mRNA Genomic Coords (5'-3'): 38790394 - 38798818				
CDS length: 1608 nt				
Protein length: 535 aa				
b Gene Ontology Classification				
GO accession	Type	Name	Code	With
GO:0005515	molecular_function	protein binding	IEA	TAIR:AT3G02530
GO:0042221	biological_process	response to chemical	IEA	TAIR:AT3G02530
GO:0005829	cellular_component	cytosol	IEA	TAIR:AT3G02530
c BlastP Searches (UniRef 100)				
Accession	% Sim	% Cov	Description	P-value
<a href="#">UniRef100_M1AYG3</a>	100	99.8	TCP domain class transcription factor n=1 Tax=Solanum tubero	1e-303
<a href="#">UniRef100_A0A6N2CBJ5</a>	100	99.8	Uncharacterized protein n=1 Tax=Solanum chilense TaxID=4083	3.9e-303
<a href="#">UniRef100_A0A3Q7F9W4</a>	100	99.8	Uncharacterized protein n=1 Tax=Solanum lycopersicum TaxID=4	1.9e-302
d PFAM hits				
Accession	Name	Match Start	Match End	E-value
<a href="#">PF00118</a>	Cpn60_TCP1	29	528	6.1e-165
e Interpro hits				
Interpro Acc	Method	Method Desc	Match Start	Match End
<a href="#">IPR027413</a>	Gene3D	GROEL	1	185
<a href="#">IPR012722</a>	TIGRFAM	chap_CCT_zeta: T-complex protein 1, zeta subunit	2	533
<a href="#">IPR012722</a>	CDD	TCP1_zeta	6	529
f Coexpression Module Assignment				
Module ID	Module Peak Expression			
6	tuber_3pm_SD			
g RNA-Seq Gene Expression Values				
SRA Run Accession	SRA Exp Accession	Sample Desc	Sample Category	TPM
<a href="#">SRR15401435</a>	<a href="#">SRX11703628</a>	closedflower_control_C_R1	Flower	35.2437
<a href="#">SRR15401434</a>	<a href="#">SRX11703629</a>	closedflower_control_C_R2	Flower	32.994
<a href="#">SRR15401433</a>	<a href="#">SRX11703630</a>	closedflower_control_C_R3	Flower	33.8838