

Neuroengineering Approaches Assessing Structural and Functional Changes of Motor Descending Pathways in Stroke

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Abstract

Stroke is a leading cause of adult disability worldwide, with approximately 101 million survivors globally. Over 60% of these individuals suffer from long-term, often lifelong, movement impairments that significantly hinder their ability to perform essential daily activities and maintain independence. Post-stroke movement disabilities are highly associated with structural and functional changes in motor descending pathways, particularly the corticospinal tract (CST) and other indirect motor pathways via the brainstem. For decades, neuroengineers have been working to quantitatively evaluate the post-stroke changes of motor descending pathways, aiming to establish a precision prognosis and tailoring treatments to post-stroke motor impairment. However, a clear and practicable technique has not yet been established as a breakthrough to change the standard of care for current clinical practice. In this review, we outline recent progress in neuroimaging, neuromodulation, and electrophysiological approaches for assessing structural and functional changes of motor descending pathways in stroke. We also discuss their limitations and challenges, arguing the need of artificial intelligence and large multi-modal data registry for a groundbreaking advance to this important topic.

Keywords: Stroke, Motor Descending Pathways, MRI, Transcranial Magnetic Stimulation (TMS), Cortico-muscular connectivity

1. Introduction

Stroke is a leading cause of adult long term disability worldwide¹. There are approximately 101 million stroke survivors in the world with over 60% of them experiencing long-term movement disabilities². These impairments are often life-long and lead to difficulty with activities of daily living and returning to work³. A stroke lesion results in focal damage to motor or sensory cortices and their descending or ascending pathways. Recovery outcomes are highly variable, influenced by the lesion's location and size, associated white matter alterations (e.g., fiber loss or degeneration), and the effectiveness of therapeutic interventions and rehabilitation strategies. These factors determine the degree of functional restoration, highlighting the importance of precise lesion characterization and individualized rehabilitation plans to optimize recovery outcomes⁴.

The somatic motor system is organized into two major descending pathway systems: direct and indirect pathways. The direct pathways, including the corticospinal tract (CST), are primarily responsible for voluntary motor control. In contrast, the indirect pathways, such as the corticoreticulospinal tract (CRST), which is composed of the corticoreticular tract (CRT) and the reticulospinal tract (RST) with synaptic connections in nuclei at the brainstem, focus on reflexive and postural control of musculature⁵. Post-stroke movement disabilities are highly associated with structural and functional changes in these motor control pathways. Reduced CST fiber density severely compromises precise, individuated control of single joints^{6, 7}. CST damage also triggers secondary adaptive changes in the brain and spinal cord, including maladaptive hyperexcitability of the CRST, a key factor in severe post-stroke motor impairments^{8, 9}. This leads to stereotyped, coarse, multi-joint movements, known as pathological limb synergies¹⁰. In the upper limb, this manifests as "flexion synergy," where increased shoulder abduction causes involuntary activation of elbow, wrist, and finger flexors, a hallmark of post-stroke motor dysfunction^{11, 12}.

The mechanisms underlying these motor impairments are closely linked to damage in motor pathways and the plasticity of the nervous system following a stroke. The RST consists of two components: the dorsal and medial RST. The dorsal RST originates from the medullary reticular formation, receives input from the contralateral primary motor cortex, and provides inhibitory input to spinal reflex circuits, descending ipsilaterally to the spinal cord¹³. The medial RST, originating from the pontine reticular formation, receives input from the ipsilateral premotor cortex and supplementary motor areas, descending ipsilaterally to the spinal cord and providing excitatory input to the spinal motor network⁸. Post-stroke, damage to the ipsilesional motor cortex, CST and/or CRT leads to significant motor deficits.

CST damage impairs voluntary limb movement, while CRT damage reduces input to the contralateral medullary reticular formation, resulting in hypoactivity of the contralateral dorsal RST's inhibitory effects on spinal stretch reflexes¹⁴. Concurrently, inputs from the contralateral premotor cortex and supplementary motor areas to the contralateral pontine reticular formation become hyperexcitable, driving spasticity and abnormal flexion synergy^{15, 16}. Spasticity, characterized by hyperactive stretch reflexes, affects up to 65% of stroke patients and is associated with poor motor function^{17, 18}.

Due to the critical role of motor descending pathways in post-stroke movement impairments, specifically flexion synergy and spasticity, neuroengineers are developing methods to quantify changes in these pathways. Techniques such as neuroimaging, neuromodulation, and electrophysiological approaches are being explored to assess these changes. This review discusses the recent advancements in these methods, highlighting current limitations, challenges, and future research directions.

2. Imaging-based approach for detecting and evaluating structural changes of motor descending motor pathways post stroke

2.1 *Lesion load on the corticospinal tract using structural imaging*

The CST, extending from the motor cortex through the internal capsule and cerebral peduncles and decussating at the medulla into the spinal cord, is frequently affected in both ischemic and hemorrhagic strokes. This leads to lasting motor deficits due to disruption of signal transmission between the brain and the extremities¹⁹. Structural magnetic resonance imaging (MRI), particularly T1-weighted imaging (T1WI) and T2-weighted imaging (T2WI) are utilized (**Figure 1**) because these allow for detailed assessment of ischemic and hemorrhagic lesions in cortical and subcortical areas where CST fibers are located^{20, 21}. T1WI is a powerful imaging modality providing high-resolution anatomical details and is particularly useful for detecting primary ischemic or hemorrhagic lesions, as well as assessing atrophy and secondary degeneration. This allows for precise localization of stroke lesions and their relationship to motor pathways. T2WI is highly sensitive to changes in water content and is particularly effective in identifying edema, gliosis, and other pathological changes associated with CST damage²². Each phase of stroke - acute, subacute, and chronic—presents unique structural changes in the CST²³. T1WI and T2WI offer distinct advantages in visualizing these changes, helping clinicians evaluate lesion load, secondary degeneration, and the likelihood of functional

improvement²¹.

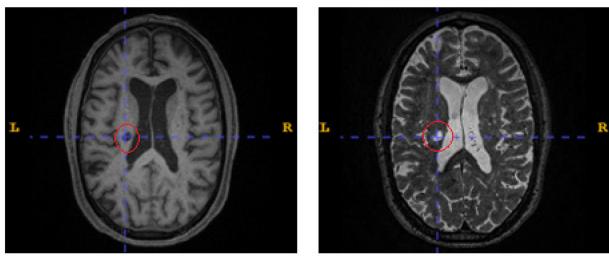


Figure 1. Lesion appears dark on T1-weighted (left) and bright on T2-weighted MRI images (right).

In the acute stage of ischemic stroke, T1WI and T2WI help identify the immediate effects of injury in the CST²⁴. T1WI shows early hypointensity in the infarcted area, allowing for lesion localization. T2WI is especially useful for detecting edema, which appears as hyperintensity in the affected CST^{25, 26}. Edema in the acute stage can expand into surrounding motor-related areas, increasing lesion load and the risk of additional motor deficits²⁷. T1WI and T2WI also help identify the risk of midline shift or compression on the CST, which can worsen motor outcomes. During the subacute stage of ischemic stroke, MRI changes become more pronounced as edema subsides and gliosis begins. T1WI can reveal hypointense regions as the infarcted tissue undergoes necrosis and volume loss. Detecting this degeneration is crucial for assessing prognosis, as it often correlates with motor impairment severity²⁸.

In chronic stages, Wallerian degeneration, gliosis and scarring become prominent, and the CST develops persistent atrophy. Wallerian degeneration is a process of progressive axonal degeneration that occurs following injury to a neuron, often due to stroke, traumatic injury, or demyelinating disease. This degeneration involves the breakdown of the distal portion of the axon and its myelin sheath after a disconnection from the neuron's cell body²⁹⁻³¹. The process is primarily observed in the CST, especially in regions distal to the primary lesion, such as the cerebral peduncle and pons. T1WI reveals hypointense signals and associated atrophy, while T2WI shows hyperintense signal corresponding to ongoing gliosis^{32, 33}. These signal changes and degree of atrophy along the CST indicate sites of irreversible axonal loss and reduced plasticity potential. Identifying these changes can aid in assessing long-term outcomes for motor function in affected patients^{34, 35}.

Quantitative analyses of post stroke lesion on MRI can be predictive of motor outcomes, as larger lesions within or near the CST often correlate with more significant motor impairment^{36, 37}. A study indicated that lesion load to both the primary motor cortex (M1) and ventral premotor cortex derived from T1WI were strongly related to stroke motor severity indexed by Fugl-Meyer Assessment cut-off scores³⁸. A meta-analysis assessed the correlation between MRI-based

lesion size and functional outcomes in patients with stroke. This research included various studies using different techniques to estimate acute lesion size, such as structural MRI, diffusion-weighted imaging (DWI), and perfusion-weighted imaging (PWI)³⁹. Notably, the analysis found that T2WI was more reliable for estimating lesion volume compared to other sequences³⁹. Consequently, T2WI is a preferred modality to approximate final infarct size, offering valuable insights into the relationships between infarct size, functional outcomes, and prognosis⁴⁰.

In recent years, algorithms driven by artificial intelligence (AI) have leveraged structural MRI to improve lesion detection, segmentation, and quantification, thereby aiding in accurate stroke severity assessment and outcome prediction⁴¹. Guo et al. used machine learning algorithms in combination with support vector machine (SVM) classifiers, achieving a Dice coefficient of 0.73 for detecting ischemic stroke lesions on T1WI^{42, 43}. Other studies have illustrated that the Hybrid UNet Transformer (HUT) excelled in single-modality segmentation on the Anatomical Tracings of Lesions After Stroke (ATLAS) dataset, demonstrating an increase of 4.84% in the Dice score and a notable 41% improvement in the Hausdorff Distance score⁴⁴. Classification models have also been developed utilizing decision tree and k-nearest neighbors (kNN) algorithms. A recent study revealed that the decision tree algorithm surpassed kNN in distinguishing between thrombotic, hemorrhagic, and embolic strokes, demonstrating high classification accuracy⁴⁵. With the advancement of novel AI technologies such as SAM, large language models (LLMs), and VMamba, future AI-assisted acute stroke diagnosis is expected to achieve higher levels of accuracy and reliability⁴⁶.

2.2 Integrity assessment of motor pathways using diffusion imaging

To localize the damaged pathways within the CST, diffusion tensor imaging (DTI)—calculated from diffusion-weighted MRI (DWI)—provides a method to examine the integrity of white matter through tractography⁴⁷. By quantifying anisotropic diffusion levels of white matter via fractional anisotropy (FA) values, DTI can assess brain microstructures and white matter integrity in various neurological conditions⁴⁸. FA is sensitive to the relative magnitude of the eigenvalues of the diffusion tensor, which reflect the directionality and extent of water diffusion⁴⁹. Axial diffusivity, the first eigenvalue of the diffusion tensor, reflects the magnitude of diffusivity parallel to the direction of maximal diffusion. In the white matter, axial diffusivity is oriented along the direction of axons, and is thought to primarily relate to axonal integrity.

Given that lesions from stroke often disrupt white matter, lower FA values, especially on the ipsilesional side, are

associated with significant motor deficits⁷. Recent studies have shown that lesion load in the CST can better predict motor impairment on the Fugl-Meyer Assessment, particularly in severely impaired stroke patients. Furthermore, analyzing MRI of the brainstem and spinal cord has revealed that damage to the CST and other sensorimotor pathways, e.g., the reticulospinal and rubrospinal tracts with an increased FA, contributes to motor impairment following stroke^{50, 51}. Changes in white matter integrity in these pathways correlate with the severity of motor deficits, suggesting that abnormal motor synergies and hand impairments may be related to neuroplastic changes in bulbospinal pathways. This suggests that alterations in both FA and diffusivity can provide valuable insight into how motor pathways may be affected by stroke⁴⁶. This process of tracking water diffusion, which reflects the orientation of neural fibers, is known as tractography. FA values are typically used with two main types of tractography algorithms, deterministic and probabilistic, to improve the accuracy of fiber tracking⁵²⁻⁵⁶. However, deterministic tractography has the limitation of assuming a fixed fiber direction at each voxel, without accounting for the inherent uncertainty in fiber orientation. In contrast, probabilistic tractography samples a wider range of possible fiber orientations, generating multiple potential pathways to better capture uncertainty. While this approach is more accurate, it requires more computational resources. Some studies also use CST masks, calculated from probabilistic CST tractography either in healthy control subjects or individually for stroke patients, and overlay them onto the FA images of stroke patients. However, there are growing concerns about the accuracy of overlaying FA images by directly applying only the masks of CST tracts from either controls or stroke patients, due to variability in lesion location and size. In recent years, more detailed subsegments of the motor cortex have been explored, which improve the accuracy of tractography. For example, Derek B. Archer et al. (2018)⁵⁷ proposed a high-resolution sensorimotor area tract template (SMATT) that segments corticofugal tracts by setting seed points at six key cortical regions (HMAT) within the specific sensorimotor areas proposed by Mayka et al. (2006)⁵⁸, using DTI of 100 subjects⁵⁷. The SMATT, along with a probabilistic version that quantifies tract overlap, offers new tools for segmenting and labeling sensorimotor tracts at higher spatial resolution (**Figure 2**). However, concerns remain about the accuracy of directly applying these outcome masks to stroke cases for extracting FA values. To address the potential inaccuracies in fiber tracking in the presence of brain lesions, Qiurong Yu et al. (2023) combined the six subregional masks from SMATT with a transcallosal tract template (TCATT) for frontal tracts, allowing for more precise fiber projections (**Figure 3**)^{59, 60}.

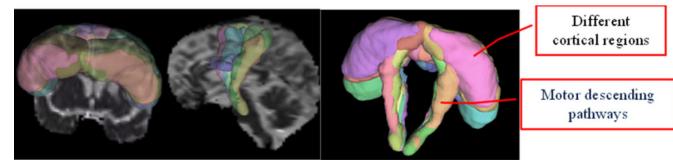


Figure 2. Different cortical regions (HMAT; left) and their underlying motor descending pathways (SMATT; middle).

They also integrated resting-state fMRI to enhance functional connectivity assessments, further improving the accuracy of their approach. Tractography is also highly sensitive to factors such as image quality, scanning parameters, and settings (e.g., diffusion modes, number of directions, and isotropic voxel size). These variables can impact the tractography process, including the step sizes used for tracking, potentially introducing bias and making it more difficult to accurately predict changes in specific motor pathways. From a practical perspective, increasing the number of diffusion directions leads to longer scan times, especially when patients require urgent care. For example, scanning with 90 diffusion directions can take up to 20 minutes⁶¹. Traditionally, clinical practice has used only three directions, though more recently, 6 to 12 directions have become common in neuroimaging. For research purposes, however, more directions are typically used. However, even this increase may still limit the accuracy of fiber tracking, as the eigenvectors and eigenvalues derived from water molecule diffusion may not fully capture the complexity of the white matter pathways.

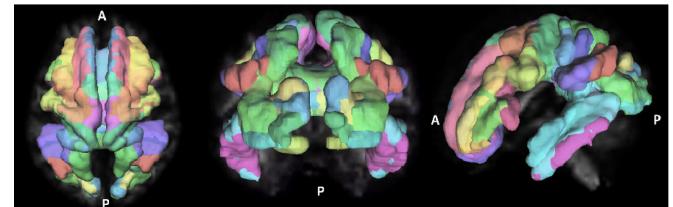


Figure 3. TCATT atlas in transverse, sagittal, and coronal views visualize the neural pathways that connect the left and right hemispheres of the brain through the corpus callosum.

As computational technology has developed, several software programs and algorithms for automatic tractography have been introduced, including DSI Studio (**Figure 4**) and MRtrix⁶². These programs utilize DWI images along with tracking parameters (e.g., angles, minimal step sizes) and predefined atlas regions for seed placement and ROIs. This facilitates the visualization and projection of motor pathway fibers easier by setting the ROI based on the default atlas, including CST and CRST areas. On the other hand, some studies have even adopted AI for fiber tractography, such as DeepDTI. Later, Hongyu Li et al. (2021) proposed SuperDTI, model trained on healthy control data that shows a quantification error of less than 5% in white-matter and gray-matter regions and has even been applied for lesion detection in stroke patients⁶³.

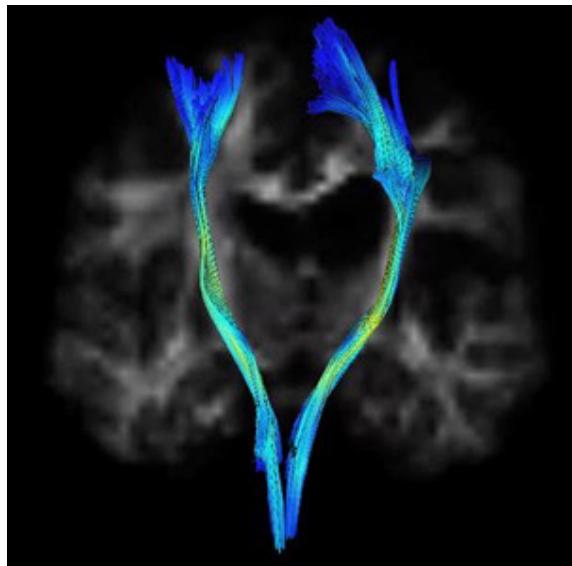


Figure 4. Automatic tractography of the corticospinal tract using the DSI Studio.

In summary, imaging-based approaches such as structural MRI provide detailed anatomical information for clinical and research purposes, helping to identify stroke lesions. Meanwhile, DWI offers a method for assessing white matter integrity through the orientation of water molecules, which is crucial for understanding the status of motor pathways following a stroke. With advancements in technology, AI now assists in lesion delineation and even performs tractography under limited conditions, enhancing our ability to analyze complex neural structures. These developments are paving the way for personalized rehabilitation strategies that are tailored to individual patient needs, ultimately improving functional outcomes.

3. TMS Motor Evoked Potential for assessing functional changes of motor descending motor pathways post stroke

3.1 Motor evoked potential on lesioned corticospinal tract

Transcranial magnetic stimulation (TMS) is a safe, non-invasive, and painless technique that can be used to investigate the excitability of the cortex and motor descending pathways⁶⁴. With TMS, a magnetic stimulus is applied via coil, the effects of the stimulation are dependent on coil shape, size, orientation⁶⁴. Motor-evoked potential (MEP) is a biphasic electromyography (EMG) response to TMS in the target muscles. In humans, the functional integrity of the CST can be assessed after stroke using TMS over ipsilesional M1 to elicit contralateral motor evoked potentials (cMEPs) in paretic upper limb muscles^{65, 66}. The simplest method for evaluation is presence or absence of cMEPs early after stroke to gain information on functional

integrity of the CST^{34, 67}. Previous studies have found that the presence of cMEPs correlated with greater strength and higher FMA scores⁶⁸. Further, if present, the features of the cMEP have gained important scientific and clinical relevance, as they have been associated to motor cortex excitability, and integrity and conduction velocity of the activated fibers of the motor pathway⁶⁹. Specifically, the peak-to-peak amplitude provides insights into cortico-spinal excitability and the latency (or onset) provides information on the conduction within triggered neuronal pathways. Both latency and amplitude of cMEPs have been shown to be significantly correlated with clinical motor assessments post stroke⁷⁰⁻⁷³. This is shown in **Figure 5**.

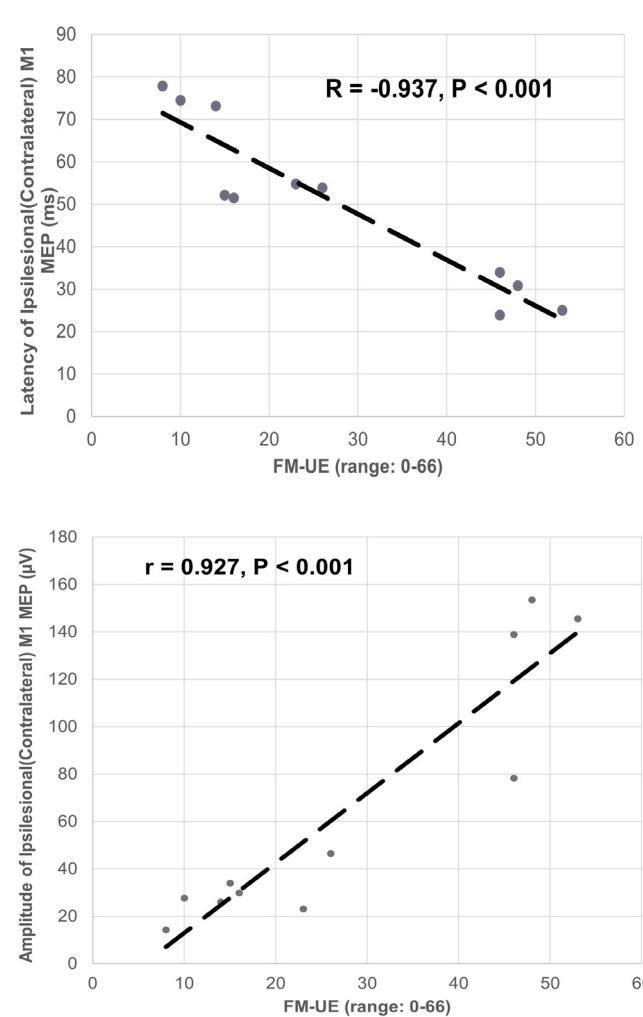


Figure 5. Correlation between the latency and amplitude of ipsilesional (contralateral) M1 MEP and FM-UE. *Adopted from [63], authors received permission from the primary author and colleagues, CC by 4.0*

However, the extraction of features from cMEPs elicited by TMS is performed manually, increasing variability due to observer-dependent subjectivity. To eliminate trials without MEPs, or with poor signal-to-noise ratio, several studies use manual protocols comprising a visual inspection of the EMG

trace from each trial, either during MEP acquisition or in posterior offline processing⁷⁴⁻⁷⁶. Further, the methodology employed for MEP signal processing is heterogeneous throughout the literature^{77, 78}. Therefore, recent research in this field is focused on unbiased methods to automate amplitude and latency detection. Many algorithms have been presented that can accurately detect MEP amplitude and latency in healthy controls⁷⁹⁻⁸¹. However, due to neurological lesion in stroke subjects, there is an increase in the variability of the extracted features⁸². This makes automation of MEP difficult and can cause unreliability even with good signal-to-noise ratio⁸³. A recent method has been proposed by Tecuapetla-Trejo et al (2021). Their algorithm was able to successfully automate MEP selection and feature extraction for stroke subject in the acute phase, with no significant differences from manual measurement performed by three experts⁸⁴.

Another cause of variability in the MEP measures is the activation of the target muscle, a pre-activation will decrease the threshold required to produce a MEP, and the MEP amplitude will be larger than that of a muscle at rest⁸⁵. This can cause intra- and inter-subject MEP variability, as subject-related parameters such as anticipation of stimulation, active thinking about body movements, or action observations have shown to effect MEP characteristics⁸⁶. Due to the high variability of MEPs from stimulation pulse to pulse, definitions of intensity thresholds have been established as an attempt to individualize and standardize the stimulation intensity applied during TMS⁸⁷. The conventional criteria for resting motor threshold (RMT) is the lowest stimulation intensity required to elicit an MEP of 50 μ V in ≥ 5 of 10 trials⁸⁸ and for activated motor threshold (AMT) is defined as the lowest intensity to produce a 200 μ V MEP in ≥ 5 of 10 trials⁸⁹. However, recently, with the advancement of TMS technologies, MEPs that fall below the conventional RMT criteria (responses < 50 μ V) have been shown to be informative and reliable⁹⁰. This is especially important for stroke participants as they have a lower amplitude in the paretic limb, compared to the non-paretic limb and healthy controls^{91, 92}.

3.2 Motor evoked potential on contralateral cortico-reticulospinal tract

Post-stroke movement disorders are not only linked to decreased function in the ipsilateral M1 and its descending CST but are also associated with enhanced activity in the contralateral premotor and supplementary motor areas^{93, 94}. This suggests that post-stroke motor dysfunction involves complex interactions between both hemispheres, impacting motor control and requiring a more comprehensive approach to understanding stroke recovery mechanisms⁹⁵⁻⁹⁷. Most studies aiming to activate the contralateral CRST use previously published methodology for targeting the premotor

cortex, by starting at M1 and moving the coil 1-3 cm medial and 2.5-3 cm anterior of the contralateral M1 and using an anterior-posterior coil orientation⁹⁸⁻¹⁰⁰. This is shown visually in Figure 6.

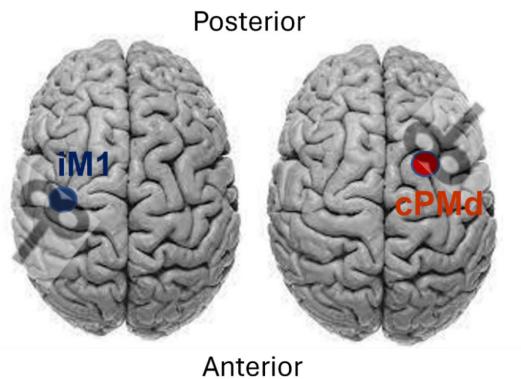


Figure 6. Coil orientation for stimulating ipsilesional primary motor cortex (iM1) and contralateral dorsal premotor area (cPMd), assuming the lesion on the left side. Adapted from [63], authors received permission from the primary author and colleagues, CC by 4.0

Recent research indicates post-stroke changes of CRST innervation in upper limb muscles, particularly biceps muscles, underscoring its significance in upper limb rehabilitation¹⁰¹. In stroke patients, TMS of the contralateral hemisphere (at the premotor cortex and at M1) can elicit responses in ipsilateral muscles of the paretic arm, attributed to the activation of oligosynaptic corticobulbospinal pathways. This response likely reflects hyperexcitability of the CRST¹⁰²⁻¹⁰⁵. While TMS is used to assess CRST excitability, the literature shows variability in stimulation targets, with inconsistent locations and frequent use of anterior-posterior coil orientation¹⁰⁶.

The presence of ipsilateral motor-evoked potentials (iMEPs) from contralateral motor cortex is associated with motor and neurophysiological impairments, with more frequent iMEPs observed in individuals with greater impairment, suggesting maladaptive role of contralateral CRST hyperexcitability. A study found that iMEP occurrence is higher in stroke patients with severe CST damage compared to those with milder damage^{93, 107}. Stronger contralateral CRST projections (reflected by higher iMEP amplitudes) correlate with increased upper limb strength, while stronger ipsilesional CST projections are linked to better motor control and improved muscle individuation¹⁰⁸. The differences in excitatory and inhibitory capacities of the contralateral CRST and ipsilesional CST provide insight into their roles: CST terminals are predominantly excitatory, while CRST terminals include both excitatory and a significant minority of inhibitory connections^{109, 110}. This dual role of the CRST may mediate the muscle suppression needed to support strength, offering a potential mechanism for its contribution to motor recovery¹⁰⁸.

4. Cortico-muscular connectivity and inter-muscular connectivity for brain-muscular communication post stroke

4.1 Cortico-muscular connectivity

Corticomuscular techniques investigate the connections of the cerebral cortex and muscle activation using functional corticomuscular coherence (fCMC)^{111, 112} and corticomuscular coherence (CMC)¹¹³. CMC, first presented by Conway in 1995, emphasizes the importance of cortical neurons in coordinating motor unit output and demonstrates how cortical activation directly affects muscle movements¹¹⁴⁻¹¹⁶. Additionally, these measurements capture the synchronization and coordination required for controlled movement, reflecting both efferent and afferent motor pathways¹¹⁷⁻¹¹⁹.

CMC is most commonly measured by monitoring electroencephalography (EEG) signals and corresponding surface electromyography (EMG), simultaneously. This allows for visualization of real-time motor command projections and sensory feedback during voluntary activities, allowing for the identification of pathway-specific corticomuscular interactions¹²⁰⁻¹²⁴. Large advancement has been made in the field of CMC analysis, methods such as extended partial directed coherence (ePDC)¹²⁵, multiscale transfer entropy (MSTSE)^{126, 127}, multi-spectral phase coherence¹²⁸, wavelet-based coherence¹²⁹⁻¹³¹, and generalized cortico-muscular-cortical network (gCMCN)¹³² have emerged. These methods improve causal coupling assessments across brain signals and can show complicated connections between the motor cortex and peripheral muscles^{126, 128, 133, 134}. Collectively, these new results highlight the importance of fCMC in understanding motor control processes, particularly after stroke, while also emphasizing the need for enhanced techniques that go beyond typical coherence analyses to account for cortical specialization^{125, 126, 135}.

Linear neural connection mostly reflects direct corticospinal tracts and frequently decreases in the ipsilesional hemisphere following a stroke^{119, 136-138}. However, no studies have found a substantial increase in CMC in the contralateral hemisphere during flexion synergy expression^{139, 140}. This finding raises questions about the usefulness of linear coherence for studying contralateral indirect motor pathways, which may operate on a nonlinear basis. Nonlinear connectivity, defined as coupling across different frequencies, is thought to result from the nonlinear characteristic of synaptic connections, which can aggregate across several synapses in indirect motor pathways^{141, 142}. In contrast, the direct corticospinal pathways have fewer synapses¹⁴³. Cross-spectral coherence (CSC) is a method for identifying various nonlinear interactions, including harmonic and intermodulation couplings, which can be

observed in both static and dynamic nonlinearities^{114, 119}. Additionally, the creation of the n:m coherence analysis approach enables a comprehensive measurement of cross-frequency coupling between different frequency components approach^{144, 145}. In chronic hemiparetic stroke, the recruitment of contralateral indirect motor pathways, such as the corticoreticulospinal tract (CRST), has been linked to the expression of flexion synergy, as evidenced by cross-spectral connectivity analyses that show increased nonlinear connectivity during shoulder abduction tasks¹⁶, as shown in Figure 7.

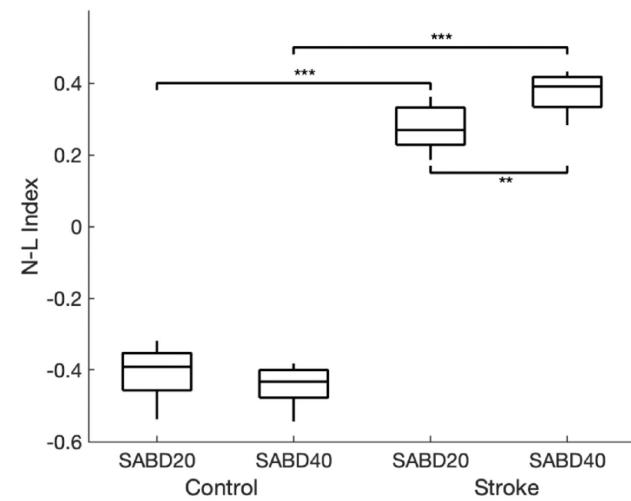


Figure 7. N-L index (nonlinear-over linear connectivity index): $N-L\text{ Index} = (SN-SL) / (SN+SL)$, where SL is the sum of linear connectivity and SN is sum of nonlinear connectivity for control and stroke subjects with different level of shoulder abduction (SABD). Two-sample t-test was applied across groups with same level of SABD, and paired t-test was applied among groups with different levels of SABD. ** for p -value < 0.01 and *** for p -value < 0.001 . Adapted from [16], authors received permission from the corresponding author and colleagues, CC by 4.0

Cortical oscillations between the cortex and the muscle are also direction-dependent^{133, 144, 145}. Previous research has demonstrated that information flow between cortico-cortical regions is directed, and similar directional viewpoints are recognized while analyzing corticomuscular interactions^{119, 133, 141}. The strength of this oscillatory component of the cortical drive can serve as an index of cortical excitability and corticospinal tract integrity^{139, 140, 146, 147}, as a primary cause of movement difficulties in stroke patients is irregular nerve oscillation transmission due to the reduction in the brain's neurological control over muscle movements¹⁴⁸. Additionally, assessing alterations in CMC time delays may also give insight into changes in motor descending pathways post stroke. Stroke participants have been shown to have significantly longer nerve conduction delays between the EEG signal from the ipsilesional motor cortex and the EMG signal from the tested muscles, suggesting the increased delay caused by the absence of direct corticospinal projections¹⁴⁹. One could also assess CMC time delays

between contralesional cortex and the muscles of the paretic limb to evaluate both corticospinal tract's damage and signal transmission in the contralesional cortico-reticulospinal pathway^{15, 150}.

Following a stroke, it is important to highlight that patients have considerably lower mean beta CMC values than healthy controls¹⁵¹. Beta oscillations help to maintain stable load during muscular contractions and relate to higher force output in dynamic tasks¹⁵². Beta-CMC is a significant measure of motor system performance as it is essential for connecting motor cortex activity to muscle function during hand movements, indicating integration and coordination^{112, 120, 146}. Recently, Beta-CMC has been found to be considerably disrupted between ipsilesional motor cortex and the paretic side deltoid muscle in stroke patients, as seen by an increased nerve conduction delay from the motor cortex to the deltoid¹⁴⁹. Chronic stroke patients have also been shown to exhibit significantly lower Beta-CMC during stable force contraction tasks compared to healthy persons, implying that proprioceptive disruptions may reduce beta oscillations, decreasing fCMC and motor control precision^{128, 135, 153}. Measuring Beta-CMC is an effective method for evaluating motor descending pathways and evaluating both linear and nonlinear brain-muscle connection can reveal information about the participation of ipsilesional CST and contralesional CRST. For example, more linearity in Beta-CMC indicates increased contralateral corticospinal activity, whereas higher nonlinearity indicates increased ipsilateral corticoreticulospinal activity, offering a holistic picture of motor pathway functionality¹⁴³.

4.2 Inter-muscular connectivity

Stroke-induced motor impairments disrupt not only the direct brain-muscle pathways but also the coordination among various muscles that are crucial for performing complex and coordinated movements¹⁶. This occurs because the patients' intermuscular coordination abilities, or muscle synergy, become altered or weakened¹⁵⁴. Previous research, using EMG patterns, found that the specific altered upper limb muscle synergies of stroke involve the abnormal coupling of shoulder and elbow muscles during dynamic reaching¹⁵⁵. Further, flexion of paretic wrist and fingers is involuntarily coupled with certain shoulder and elbow movements¹¹. Abnormal coactivation of the three heads of the deltoid muscle has also been observed in stroke¹⁵⁶. These abnormal muscle synergies are associated with motor impairment; the incidence of abnormal muscle co-activation increases as the severity of motor impairment increases^{157, 158}. From this perspective, while traditional stroke rehabilitation often focuses on the strength and recovery of individual muscles, since actual movement occurs through the cooperation of various muscles, the importance of

investigating abnormal muscle coactivation should be emphasized in the stroke neurorehabilitation.

A representative method used to evaluate intermuscular interactions is inter-muscular coherence (IMC). This analytical technique is an effective way to observe neural synchronization between muscles¹⁵⁹, serving as a metric that quantifies the correlation between pairs of electromyographic (EMG) signals across frequency bands^{160, 161}. IMC explains the extent of shared contributions of descending neural activity among the motor neuron pools located in the spinal cord and the strength of neural synchronization at different frequency bands¹⁶², and it signifies the functional relationships between muscles that are activated together in specific patterns or synergies to effectively complete motor tasks^{152, 159}. IMC can be calculated using the following equation:

$$|C_{xy}(f)|^2 = \frac{|P_{xy}(f)|^2}{P_{xx}(f) \cdot P_{yy}(f)}$$

where f denotes the frequency of each EMG signal, $P_{xx}(f)$ and $P_{yy}(f)$ are auto-spectra of the rectified EMG signals of any muscle pair at a given frequency and $P_{xy}(f)$ is the cross-spectrum between them. To measure the importance of the coherence between muscles, a metric known as confidence level (CL) is implemented¹⁶³:

$$CL = 1 - (1 - \alpha)^{\frac{1}{L-1}}$$

Where α signifies the degree of significance ($\alpha = 0.95$) and L indicates the number of data segments utilized for spectrum estimation. The coherence between two muscles is recognized as significant if its value surpasses the confidence level (CL)¹⁶⁴.

A review of previous studies reveals that IMC analysis found significantly lower coherence in the alpha frequency band between the anterior deltoid and triceps brachii muscles in stroke patients compared to healthy controls¹⁶⁵. Watanabe et al. found that using beta-band IMC, which reflects CST activity, intermuscular coherence increased with rising difficulty in postural tasks among young and elderly adults¹⁶⁶. Further, IMC in the beta frequency band can serve as a synchronization index for assessing upper limb motor dysfunction in stroke patients¹⁶⁷. In this way, IMC across each frequency band can serve as a synchronization index for evaluating motor dysfunction in stroke patients, helping to understand patient characteristics.

In the context of motor descending pathways, IMC can serve as a critical indicator of the flexion synergy that results from increased reliance on the corticoreticulospinal tract. As IMC reflects the shared descending neural drive among

motor neuron pools in the spinal cord. IMC plays an essential role in understanding and analyzing these symptoms, providing insights that can inform the development of more sophisticated rehabilitation protocols. Such research is crucial for enhancing the recovery process in stroke patients.

5. Limitation and challenges

Although much work has been done in the field, several challenges remain for accurately quantifying changes in motor descending pathways post stroke. Many of the techniques described use manual processes that are labor-intensive, time consuming, and have inter-observer variability. The analysis highly depends on the experience of the observer, leading to inconsistencies and potential errors. AI-based software is being developed to streamline these processes, but it is not yet widely adopted in clinical practice¹⁶⁸. There are also unique challenges in improving these AI-based methods for stroke. Data quality and availability complicate the development of reliable AI tools. Moreover, the "black box" nature of many AI algorithms, where the decision-making process is not easily interpretable, creates hesitation among clinicians to fully trust AI-based tools. The integration of AI models into clinical workflows remains challenging due to concerns the accuracy.

While there is a large number of randomized control trials evaluating stroke rehabilitation of the upper extremity, research quality continues to be a challenge. Only a small percentage of have multi-center trials and many have limited sample sizes¹⁶⁹. In a recent review of randomized controlled trials for rehabilitation of the upper limb post stroke, the median sample size (start/finish) was found to be 30 (IQR 20-48)/29 (IQR 19-44)¹⁶⁹. Sample size is further hindered by difficulty in recruitment. Recruitment and retention for stroke rehabilitation trials have barriers such as lack of understanding of the trial, burdensome time commitment (work or lack of childcare), and transportation¹⁷⁰. Neither recruitment rate or recruitment efficiency has increased in stroke trials over the past 25 years; if anything, they have decreased¹⁷¹. The small sample sizes within these trials affect the accuracy and hinder the creation of objective methods for assessing motor descending pathways post stroke.

6. Future direction: artificial intelligence and large multi-modal data registry

As mentioned in the previous section, there are significant challenges in the field of neuroengineering; however, the techniques are still evolving. With more publicly available datasets related to stroke, such as The Ischemic Stroke Lesion Segmentation (ISLES), which includes FLAIR, DWI, and Apparent diffusion coefficient (ADC) images, and Anatomical Tracings of Lesions After Stroke (ATLAS), which provides T1 images from patients at various stages

(acute, subacute, and chronic), researchers can now utilize multi-modal MR images^{172, 173}. These images come from different disease stages and even different scanners (e.g., GE, Philips, and Siemens). Despite potential challenges with co-registration and the fact that some images may have only undergone preliminary preprocessing, these datasets reflect conditions that are much closer to real-life scenarios.

Additionally, recently the American Heart Association/American Stroke Association hosted a challenge to merge stroke registry data, which includes patient-level and inpatient data from hospitals across the US¹⁷⁴. This initiative has gained attention from the public, hospitals, and academic institutions, and is forming a trend of integrating multi-modal imaging with non-imaging datasets. Once such comprehensive datasets are established, AI models can be trained with both imaging data and clinical assessments, such as stroke outcomes, complications from treatment, or even muscle assessments. With the integration of non-imaging data, such as functional movement assessments, AI models can be further trained to predict the effectiveness of rehabilitation interventions. These models could identify patterns that link muscle function recovery with brain lesion characteristics, enabling personalized rehabilitation plans that optimize recovery for each patient. Furthermore, AI could help track progress over time, offering real-time feedback to clinicians and patients, improving rehabilitation protocols and outcomes.

With this richer, more holistic dataset, AI models can be developed to be more objective and accurate. These models may even be able to predict patient outcomes based on lesion characteristics, functional impairments, and muscle rehabilitation progress, offering the potential for personalized treatment in clinical practice in the future.

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References

- [1]. Martin SS, Aday AW, Almarzooq ZI, Anderson CA, Arora P, Avery CL, Baker-Smith CM, Barone Gibbs B, Beaton AZ, Boehme

AK. 2024 heart disease and stroke statistics: a report of US and global data from the American Heart Association. *Circulation*. 2024;149(8):e347-e913.

[2]. Owolabi MO, Thrift AG, Martins S, Johnson W, Pandian J, Abd-Allah F, Varghese C, Mahal A, Yaria J, Phan HT, Roth G, Gall SL, Beare R, Phan TG, Mikulik R, Norrving B, Feigin VL. The state of stroke services across the globe: Report of World Stroke Organization-World Health Organization surveys. *Int J Stroke*. 2021;16(8):889-901. Epub 20210527. doi: 10.1177/17474930211019568. PubMed PMID: 33988062; PMCID: PMC8800855.

[3]. Tsao CW, Aday AW, Almarzooq ZI, Anderson CAM, Arora P, Avery CL, Baker-Smith CM, Beaton AZ, Boehme AK, Buxton AE, Commodore-Mensah Y, Elkind MSV, Evenson KR, Eze-Nliam C, Fugar S, Generoso G, Heard DG, Hiremath S, Ho JE, Kalani R, Kazi DS, Ko D, Levine DA, Liu J, Ma J, Magnani JW, Michos ED, Mussolino ME, Navaneethan SD, Parikh NI, Poudel R, Rezk-Hanna M, Roth GA, Shah NS, St-Onge M-P, Thacker EL, Virani SS, Voeks JH, Wang N-Y, Wong ND, Wong SS, Yaffe K, Martin SS. Heart Disease and Stroke Statistics—2023 Update: A Report From the American Heart Association. *Circulation*. 2023;147(8):e93-e621. doi: doi:10.1161/CIR.0000000000001123.

[4]. Nudo RJ, Wise BM, SiFuentes F, Milliken GW. Neural substrates for the effects of rehabilitative training on motor recovery after ischemic infarct. *Science*. 1996;272(5269):1791-4.

[5]. Noback CR, Ruggiero DA, Strominger NL, Demarest RJ. The human nervous system: structure and function: Springer Science & Business Media; 2005.

[6]. Puig J, Blasco G, Daunis-I-Estadella J, Thomalla G, Castellanos M, Figueras J, Remollo S, van Eedenburg C, Sánchez-González J, Serena J. Decreased corticospinal tract fractional anisotropy predicts long-term motor outcome after stroke. *Stroke*. 2013;44(7):2016-8.

[7]. Schaechter JD, Fricker ZP, Perdue KL, Helmer KG, Vangel MG, Greve DN, Makris N. Microstructural status of ipsilesional and contralesional corticospinal tract correlates with motor skill in chronic stroke patients. *Human brain mapping*. 2009;30(11):3461-74.

[8]. Li S, Chen Y-T, Francisco GE, Zhou P, Rymer WZ. A unifying pathophysiological account for post-stroke spasticity and disordered motor control. *Frontiers in neurology*. 2019;10:468.

[9]. Li S. Spasticity, Motor Recovery, and Neural Plasticity after Stroke. *Front Neurol*. 2017;8:120. Epub 20170403. doi: 10.3389/fneur.2017.00120. PubMed PMID: 28421032; PMCID: PMC5377239.

[10]. McMorland AJ, Runnalls KD, Byblow WD. A neuroanatomical framework for upper limb synergies after stroke. *Frontiers in human neuroscience*. 2015;9:82.

[11]. Miller LC, Dewald JP. Involuntary paretic wrist/finger flexion forces and EMG increase with shoulder abduction load in individuals with chronic stroke. *Clinical neurophysiology*. 2012;123(6):1216-25.

[12]. Brunnstrom S. Movement therapy in hemiplegia. A neurophysiological approach. 1970.

[13]. Jang SH, Seo JP. The distribution of the cortical origin of the corticoreticular pathway in the human brain: a diffusion tensor imaging study. *Somatosensory & Motor Research*. 2014;31(4):204-8.

[14]. Ko S-H, Kim T, Min JH, Kim M, Ko H-Y, Shin Y-I. Corticoreticular pathway in post-stroke spasticity: a diffusion tensor imaging study. *Journal of Personalized Medicine*. 2021;11(11):1151.

[15]. McPherson JG, Chen A, Ellis MD, Yao J, Heckman C, Dewald JP. Progressive recruitment of contralesional cortico-reticulospinal pathways drives motor impairment post stroke. *The Journal of physiology*. 2018;596(7):1211-25.

[16]. Tian R, Dewald JP, Yang Y. Assessing the usage of indirect motor pathways following a hemiparetic stroke. *IEEE Transactions on Neural Systems and Rehabilitation Engineering*. 2021;29:1568-72.

[17]. Lance J. What is spasticity? *The Lancet*. 1990;335(8689):606.

[18]. Ivanhoe CB, Reistetter TA. Spasticity: the misunderstood part of the upper motor neuron syndrome. *American journal of physical medicine & rehabilitation*. 2004;83(10):S3-S9.

[19]. Møller M, Frandsen J, Andersen G, Gjedde A, Vestergaard-Poulsen P, Østergaard L. Dynamic changes in corticospinal tracts after stroke detected by fibretracking. *Journal of Neurology, Neurosurgery & Psychiatry*. 2007;78(6):587-92.

[20]. Wey H-Y, Desai VR, Duong TQ. A review of current imaging methods used in stroke research. *Neurological research*. 2013;35(10):1092-102.

[21]. Fiebach JB, Schellinger PD, Sartor K. *Stroke MRI*: Springer Science & Business Media; 2003.

[22]. González R, Hirsch J, Koroshetz W, Lev M, Schaefer P. *Acute ischemic stroke*: Springer2011.

[23]. El-Koussy M, Schroth G, Brekenfeld C, Arnold M. Imaging of acute ischemic stroke. *European neurology*. 2014;72(5-6):309-16.

[24]. Kloska SP, Wintermark M, Engelhorn T, Fiebach JB. Acute stroke magnetic resonance imaging: current status and future perspective. *Neuroradiology*. 2010;52:189-201.

[25]. Lövblad K-O, Altrichter S, Pereira VM, Vargas M, Gonzalez AM, Haller S, Sztajzel R. Imaging of acute stroke: CT and/or MRI. *Journal of Neuroradiology*. 2015;42(1):55-64.

[26]. Kim BJ, Kang HG, Kim H-J, Ahn S-H, Kim NY, Warach S, Kang D-W. Magnetic resonance imaging in acute ischemic stroke treatment. *Journal of stroke*. 2014;16(3):131.

[27]. von Kummer R, Back T. Magnetic resonance imaging in ischemic stroke. *AMERICAN JOURNAL OF NEURORADIOLOGY*. 2007;28(3):596.

[28]. Allen LM, Hasso AN, Handwerker J, Farid H. Sequence-specific MR imaging findings that are useful in dating ischemic stroke. *Radiographics*. 2012;32(5):1285-97.

[29]. Domi T, devebeur G, Shroff M, Kouzmancheva E, MacGregor DL, Kirton A. Corticospinal tract pre-wallerian degeneration: a novel outcome predictor for pediatric stroke on acute MRI. *Stroke*. 2009;40(3):780-7.

[30]. Hustings N, Lemmerling M. MRI of Wallerian degeneration in the brainstem: a pictorial essay. *Journal of the Belgian Society of Radiology*. 2021;105(1).

[31]. Thomalla G, Glauche V, Weiller C, Röther J. Time course of wallerian degeneration after ischaemic stroke revealed by diffusion tensor imaging. *Journal of Neurology, Neurosurgery & Psychiatry*. 2005;76(2):266-8.

[32]. Jiang Q, Zhang ZG, Chopp M. MRI of stroke recovery. *Stroke*. 2010;41(2):410-4.

[33]. MacIntosh BJ, Graham SJ. Magnetic resonance imaging to visualize stroke and characterize stroke recovery: a review. *Frontiers in neurology*. 2013;4:60.

[34]. Stinear CM, Barber PA, Smale PR, Coxon JP, Fleming MK, Byblow WD. Functional potential in chronic stroke patients depends on corticospinal tract integrity. *Brain*. 2007;130(1):170-80.

[35]. Sagnier S, Sibon I. The new insights into human brain imaging after stroke. *Journal of Neuroscience Research*. 2022;100(5):1171-81.

[36]. Farr TD, Wegener S. Use of magnetic resonance imaging to predict outcome after stroke: a review of experimental and clinical evidence. *Journal of Cerebral Blood Flow & Metabolism*. 2010;30(4):703-17.

[37]. Feng W, Wang J, Chhatbar PY, Doughty C, Landsittel D, Lioutas VA, Kautz SA, Schlaug G. Corticospinal tract lesion load: An imaging biomarker for stroke motor outcomes. *Ann Neurol*. 2015;78(6):860-70. Epub 20151031. doi: 10.1002/ana.24510. PubMed PMID: 26289123; PMCID: PMC4715758.

[38]. Ito KL, Kim B, Liu J, Soekadar SR, Winstein C, Yu C, Cramer SC, Schweighofer N, Liew S-L. Corticospinal tract lesion load originating from both ventral premotor and primary motor cortices are associated with post-stroke motor severity. *Neurorehabilitation and neural repair*. 2022;36(3):179-82.

[39]. Schiemannck S, Kwakkel G, Post M, Prevo A. Predictive value of ischemic lesion volume assessed with magnetic resonance imaging for neurological deficits and functional outcome poststroke: a critical review of the literature. *Neurorehabilitation and Neural Repair*. 2006;20(4):492-502.

[40]. Kane I, Sandercock P, Wardlaw J. Magnetic resonance perfusion diffusion mismatch and thrombolysis in acute ischaemic stroke: a systematic review of the evidence to date. *Journal of Neurology, Neurosurgery & Psychiatry*. 2007;78(5):485-91.

[41]. Cui L, Fan Z, Yang Y, Liu R, Wang D, Feng Y, Lu J, Fan Y. Deep learning in ischemic stroke imaging analysis: a comprehensive review. *BioMed Research International*. 2022;2022(1):2456550.

[42]. Guo D, Zheng K, Wang S, editors. *Lesion detection using T1-weighted MRI: a new approach based on functional cortical ROIs*. 2017 IEEE International Conference on Image Processing (ICIP); 2017: IEEE.

[43]. Dice LR. Measures of the Amount of Ecologic Association Between Species. *Ecology*. 1945;26(3):297-302. doi: 10.2307/1932409.

[44]. Soh WK, Yuen HY, Rajapakse JC. HUT: Hybrid UNet transformer for brain lesion and tumour segmentation. *Helion*. 2023;9(12).

[45]. Adam SY, Yousif A, Bashir MB. Classification of ischemic stroke using machine learning algorithms. *International Journal of Computer Applications*. 2016;149(10):26-31.

[46]. Wang Z, Yang W, Li Z, Rong Z, Wang X, Han J, Ma L. A 25-Year Retrospective of the Use of AI for Diagnosing Acute Stroke: Systematic Review. *Journal of Medical Internet Research*. 2024;26:e59711.

[47]. Meyer JW, Makris N, Bates JF, Caviness Jr VS, Kennedy DN. MRI-based topographic parcellation of human cerebral white matter: I. Technical foundations. *Neuroimage*. 1999;9(1):1-17.

[48]. Panton L, Garcia JH, Gutierrez JA. Cerebral white matter is highly vulnerable to ischemia. *Stroke*. 1996;27(9):1641-7.

[49]. Beaulieu C. The basis of anisotropic water diffusion in the nervous system—a technical review. *NMR in Biomedicine: An International Journal Devoted to the Development and Application of Magnetic Resonance In Vivo*. 2002;15(7-8):435-55.

[50]. Karbasforoushan H, Cohen-Adad J, Dewald J. Brainstem and spinal cord MRI identifies altered sensorimotor pathways post-stroke (vol 10, 3524, 2019). *NATURE COMMUNICATIONS*. 2020;11(1).

[51]. Owen M, Ingo C, Dewald J. Upper extremity motor impairments and microstructural changes in bulbospinal pathways in chronic hemiparetic stroke. *Frontiers in neurology*. 2017;8:258123.

[52]. Basser PJ, Pajevic S, Pierpaoli C, Duda J, Aldroubi A. In vivo fiber tractography using DT-MRI data. *Magnetic resonance in medicine*. 2000;44(4):625-32.

[53]. Yeh F-C, Verstynen TD, Wang Y, Fernández-Miranda JC, Tseng W-YI. Deterministic diffusion fiber tracking improved by quantitative anisotropy. *PloS one*. 2013;8(11):e80713.

[54]. Conturo TE, Lori NF, Cull TS, Akbudak E, Snyder AZ, Shimony JS, McKinstry RC, Burton H, Raichle ME. Tracking neuronal fiber pathways in the living human brain. *Proceedings of the National Academy of Sciences*. 1999;96(18):10422-7.

[55]. Behrens TE, Berg HJ, Jbabdi S, Rushworth MF, Woolrich MW. Probabilistic diffusion tractography with multiple fibre orientations: What can we gain? *neuroimage*. 2007;34(1):144-55.

[56]. Tournier JD, Calamante F, Connelly A. MRtrix: diffusion tractography in crossing fiber regions. *International journal of imaging systems and technology*. 2012;22(1):53-66.

[57]. Archer DB, Vaillancourt DE, Coombes SA. A template and probabilistic atlas of the human sensorimotor tracts using diffusion MRI. *Cerebral cortex*. 2018;28(5):1685-99.

[58]. Mayka MA, Corcos DM, Leurgans SE, Vaillancourt DE. Three-dimensional locations and boundaries of motor and premotor cortices as defined by functional brain imaging: a meta-analysis. *Neuroimage*. 2006;31(4):1453-74.

[59]. Yu Q, Yin D, Kaiser M, Xu G, Guo M, Liu F, Li J, Fan M. Pathway-specific mediation effect between structure, function, and motor impairment after subcortical stroke. *Neurology*. 2023;100(6):e616-e26.

[60]. Archer DB, Coombes SA, McFarland NR, DeKosky ST, Vaillancourt DE. Development of a transcallosal tractography template and its application to dementia. *Neuroimage*. 2019;200:302-12.

[61]. Ranzenberger LR, Snyder T. *Diffusion tensor imaging2019*.

[62]. Tournier J-D, Smith R, Raffelt D, Tabbara R, Dhollander T, Pietsch M, Christiaens D, Jeurissen B, Yeh C-H, Connelly A. MRtrix3: A fast, flexible and open software framework for medical image processing and visualisation. *Neuroimage*. 2019;202:116137.

[63]. Li H, Liang Z, Zhang C, Liu R, Li J, Zhang W, Liang D, Shen B, Zhang X, Ge Y, Zhang J, Ying L. SuperDTI: Ultrafast DTI and fiber tractography with deep learning. *Magn Reson Med*. 2021;86(6):3334-47. Epub 20210726. doi: 10.1002/mrm.28937. PubMed PMID: 34309073.

[64]. Stinear CM, Byblow WD. The Role of TMS for Predicting Motor Recovery and Outcomes After Stroke. In: Lapchak PA, Yang G-Y, editors. *Translational Research in Stroke*. Singapore: Springer Singapore; 2017. p. 537-53.

[65]. Schambra HM, Xu J, Branscheidt M, Lindquist M, Uddin J, Steiner L, Hertler B, Kim N, Berard J, Harran MD. Differential poststroke motor recovery in an arm versus hand muscle in the absence of motor evoked potentials. *Neurorehabilitation and neural repair*. 2019;33(7):568-80.

[66]. Byblow WD, Stinear CM, Barber PA, Petoe MA, Ackerley SJ. Proportional recovery after stroke depends on corticomotor integrity. *Annals of neurology*. 2015;78(6):848-59.

[67]. Talelli P, Greenwood R, Rothwell J. Arm function after stroke: neurophysiological correlates and recovery mechanisms assessed by transcranial magnetic stimulation. *Clinical neurophysiology*. 2006;117(8):1641-59.

[68]. Schambra HM, Xu J, Branscheidt M, Lindquist M, Uddin J, Steiner L, Hertler B, Kim N, Berard J, Harran MD, Cortes JC, Kitago T, Luft A, Krakauer JW, Celnik PA. Differential Poststroke Motor Recovery in an Arm Versus Hand Muscle in the Absence of Motor Evoked Potentials. *Neurorehabilitation and Neural Repair*. 2019;33(7):568-80. doi: 10.1177/1545968319850138. PubMed PMID: 31170880.

[69]. Chen R, Cros D, Curra A, Di Lazzaro V, Lefaucheur J-P, Magistris MR, Mills K, Rösler KM, Triggs WJ, Ugawa Y. The clinical diagnostic utility of transcranial magnetic stimulation: report of an IFCN committee. *Clinical neurophysiology*. 2008;119(3):504-32.

[70]. Karatzetou S, Tsipitsios D, Terzoudi A, Aggeloussis N, Vadikolias K. Transcranial magnetic stimulation implementation on stroke prognosis. *Neurological Sciences*. 2022;43(2):873-88. doi: 10.1007/s10072-021-05791-1.

[71]. Escudero JV, Sancho J, Bautista D, Escudero M, López-Trigo J. Prognostic Value of Motor Evoked Potential Obtained by Transcranial Magnetic Brain Stimulation in Motor Function Recovery in Patients With Acute Ischemic Stroke. *Stroke*. 1998;29(9):1854-9. doi: 10.1161/01.STR.29.9.1854.

[72]. Li P, Chen C, Huang B, Jiang Z, Wei J, Zeng J. Altered excitability of motor neuron pathways after stroke: more than upper motor neuron impairments. *Stroke and Vascular Neurology*. 2022;7(6):518-26. doi: 10.1136/svn-2022-001568.

[73]. Williamson JN, James SA, He D, Li S, Sidorov EV, Yang Y. High-definition transcranial direct current stimulation for upper extremity rehabilitation in moderate-to-severe ischemic stroke: a pilot study. *Frontiers in Human Neuroscience*. 2023;17:1286238.

[74]. Blesneag A, Slăvoacă D, Popa L, Stan A, Jemna N, Moldovan FI, Mureşanu D. Low-frequency rTMS in patients with subacute ischemic stroke: clinical evaluation of short and long-term outcomes and neurophysiological assessment of cortical excitability. *Journal of medicine and life*. 2015;8(3):378.

[75]. Borich MR, Wheaton LA, Brodie SM, Lakhani B, Boyd LA. Evaluating interhemispheric cortical responses to transcranial magnetic stimulation in chronic stroke: a TMS-EEG investigation. *Neuroscience letters*. 2016;618:25-30.

[76]. Ackerley SJ, Stinear CM, Barber PA, Byblow WD. Priming sensorimotor cortex to enhance task-specific training after subcortical stroke. *Clinical Neurophysiology*. 2014;125(7):1451-8.

[77]. Groppe S, Oliviero A, Eisen A, Quartarone A, Cohen L, Mall V, Kaelin-Lang A, Mima T, Rossi S, Thickbroom G. A practical guide to diagnostic transcranial magnetic stimulation: report of an IFCN committee. *Clinical Neurophysiology*. 2012;123(5):858-82.

[78]. McDonnell MN, Stinear CM. TMS measures of motor cortex function after stroke: a meta-analysis. *Brain stimulation*. 2017;10(4):721-34.

[79]. Bigoni C, Cadic-Melchior A, Vassiliadis P, Morishita T, Hummel FC. An automatized method to determine latencies of motor-evoked potentials under physiological and pathophysiological conditions. *Journal of neural engineering*. 2022;19(2):024002.

[80]. Šoda J, Vidaković MR, Lorincz J, Jerković A, Vujović I. A novel latency estimation algorithm of motor evoked potential signals. *IEEE Access*. 2020;8:193356-74.

[81]. Milardovich D, Souza VH, Zubarev I, Tugin S, Nieminen JO, Bigoni C, Hummel FC, Korhonen JT, Aydogan DB, Lioumis P. DELMEP: a deep learning algorithm for automated annotation of motor evoked potential latencies. *Scientific Reports*. 2023;13(1):8225.

[82]. Butler AJ, Kahn S, Wolf SL, Weiss P. Finger extensor variability in TMS parameters among chronic stroke patients. *Journal of neuroengineering and rehabilitation*. 2005;2:1-13.

[83]. Rábago CA, Lancaster JL, Narayana S, Zhang W, Fox PT. Automated-parameterization of the motor evoked potential and cortical silent period induced by transcranial magnetic stimulation. *Clinical neurophysiology*. 2009;120(8):1577-87.

[84]. Tecuapetla-Trejo JE, Cantillo-Negrete J, Carrillo-Mora P, Valdés-Cristerna R, Ortega-Robles E, Arias-Carrion O, Carino-Escobar RI. Automatic selection and feature extraction of motor-evoked potentials by transcranial magnetic stimulation in stroke patients. *Med Biol Eng Comput*. 2021;59(2):449-56. Epub 20210126. doi: 10.1007/s11517-021-02315-z. PubMed PMID: 33496910.

[85]. Devanne H, Lavoie B, Capaday C. Input-output properties and gain changes in the human corticospinal pathway. *Experimental brain research*. 1997;114:329-38.

[86]. Bestmann S, Harrison LM, Blankenburg F, Mars RB, Haggard P, Friston KJ, Rothwell JC. Influence of uncertainty and surprise on human corticospinal excitability during preparation for action. *Current Biology*. 2008;18(10):775-80.

[87]. Sollmann N, Tanigawa N, Bulubas L, Sabih J, Zimmer C, Ringel F, Meyer B, Krieg SM. Clinical Factors Underlying the Inter-individual Variability of the Resting Motor Threshold in Navigated Transcranial Magnetic Stimulation Motor Mapping. *Brain Topography*. 2017;30(1):98-121. doi: 10.1007/s10548-016-0536-9.

[88]. Rossini PM, Burke D, Chen R, Cohen LG, Daskalakis Z, Di Iorio R, Di Lazzaro V, Ferreri F, Fitzgerald PB, George MS, Hallett M, Lefaucheur JP, Langguth B, Matsumoto H, Miniussi C, Nitsche MA, Pascual-Leone A, Paulus W, Rossi S, Rothwell JC, Siebner HR, Ugawa Y, Walsh V, Ziemann U. Non-invasive electrical and magnetic stimulation of the brain, spinal cord, roots and peripheral nerves: Basic principles and procedures for routine clinical and research application. An updated report from an I.F.C.N. Committee. *Clinical Neurophysiology*. 2015;126(6):1071-107. doi: <https://doi.org/10.1016/j.clinph.2015.02.001>.

[89]. Vucic S, Stanley Chen K-H, Kiernan MC, Hallett M, Benninger DH, Di Lazzaro V, Rossini PM, Benussi A, Berardelli A, Currà A, Krieg SM, Lefaucheur J-P, Long Lo Y, Macdonell RA, Massimini M, Rosanova M, Picht T, Stinear CM, Paulus W, Ugawa Y, Ziemann U, Chen R. Clinical diagnostic utility of transcranial magnetic stimulation in neurological disorders. Updated report of an IFCN committee. *Clinical Neurophysiology*. 2023;150:131-75. doi: <https://doi.org/10.1016/j.clinph.2023.03.010>.

[90]. Li Z, Peterchev AV, Rothwell JC, Goetz SM. Detection of motor-evoked potentials below the noise floor: rethinking the motor stimulation threshold. *Journal of neural engineering*. 2022;19(5):056040.

[91]. Di Lazzaro V, Profice P, Pilato F, Capone F, Ranieri F, Pasqualetti P, Colosimo C, Pravatà E, Cianfoni A, Dileone M. Motor Cortex Plasticity Predicts Recovery in Acute Stroke. *Cerebral Cortex*. 2009;20(7):1523-8. doi: 10.1093/cercor/bhp216.

[92]. Lewis GN, Signal N, Taylor D. Reliability of lower limb motor evoked potentials in stroke and healthy populations: How many responses are needed? *Clinical Neurophysiology*. 2014;125(4):748-54. doi: <https://doi.org/10.1016/j.clinph.2013.07.029>.

[93]. Mooney RA, Anaya MA, Stilling JM, Celnik PA. Heightened Reticulospinal Excitability after Severe Corticospinal Damage in Chronic Stroke. *Annals of Neurology*. 2024.

[94]. Trompetto C, Assini A, Buccolieri A, Marchese R, Abbruzzese G. Motor recovery following stroke: a transcranial magnetic stimulation study. *Clinical Neurophysiology*. 2000;111(10):1860-7. doi: [https://doi.org/10.1016/S1388-2457\(00\)00419-3](https://doi.org/10.1016/S1388-2457(00)00419-3).

[95]. Buetefisch CM. Role of the contralesional hemisphere in post-stroke recovery of upper extremity motor function. *Frontiers in neurology*. 2015;6:214.

[96]. Gerloff C, Bushara K, Sailer A, Wassermann EM, Chen R, Matsuoka T, Waldvogel D, Wittenberg GF, Ishii K, Cohen LG. Multimodal imaging of brain reorganization in motor areas of the contralesional hemisphere of well recovered patients after capsular stroke. *Brain*. 2006;129(3):791-808.

[97]. Rehme AK, Fink GR, Von Cramon DY, Grefkes C. The role of the contralesional motor cortex for motor recovery in the early days after stroke assessed with longitudinal fMRI. *Cerebral cortex*. 2011;21(4):756-68.

[98]. Stephan MA, Brown R, Lega C, Penhune V. Melodic Priming of Motor Sequence Performance: The Role of the Dorsal Premotor Cortex. *Front Neurosci.* 2016;10:210. Epub 20160510. doi: 10.3389/fnins.2016.00210. PubMed PMID: 27242414; PMCID: PMC4862034.

[99]. Caramia M, Telera S, Palmieri M, Wilson-Jones M, Scalise A, Iani C, Giuffre R, Bernardi G. Ipsilateral motor activation in patients with cerebral gliomas. *Neurology.* 1998;51(1):196-202.

[100]. Tscherpel C, Hensel L, Lemberg K, Vollmer M, Volz LJ, Fink GR, Grefkes C. The differential roles of contralateral frontoparietal areas in cortical reorganization after stroke. *Brain Stimul.* 2020;13(3):614-24. Epub 20200201. doi: 10.1016/j.brs.2020.01.016. PubMed PMID: 32289686.

[101]. Taga M, Charalambous CC, Raju S, Lin J, Zhang Y, Stern E, Schambra HM. Corticoreticulospinal tract neurophysiology in an arm and hand muscle in healthy and stroke subjects. *J Physiol.* 2021;599(16):3955-71. doi: 10.1113/jp281681. PubMed PMID: 34229359; PMCID: PMC8942144.

[102]. Hammerbeck U, Tyson SF, Samraj P, Hollands K, Krakauer JW, Rothwell J. The Strength of the Corticospinal Tract Not the Reticulospinal Tract Determines Upper-Limb Impairment Level and Capacity for Skill-Acquisition in the Sub-Acute Post-Stroke Period. *Neurorehabilitation and Neural Repair.* 2021;35(9):812-22. doi: 10.1177/15459683211028243. PubMed PMID: 34219510.

[103]. Ziemann U, Ishii K, Borgheresi A, Yaseen Z, Battaglia F, Hallett M, Cincotta M, Wassermann EM. Dissociation of the pathways mediating ipsilateral and contralateral motor-evoked potentials in human hand and arm muscles. *The Journal of physiology.* 1999;518(3):895-906.

[104]. Hammerbeck U, Hoad D, Greenwood R, Rothwell JC. The unsolved role of heightened connectivity from the unaffected hemisphere to paretic arm muscles in chronic stroke. *Clinical Neurophysiology.* 2019;130(5):781-8.

[105]. Jankowska E, Edgley SA. How can corticospinal tract neurons contribute to ipsilateral movements? A question with implications for recovery of motor functions. *The Neuroscientist.* 2006;12(1):67-79.

[106]. Bradnam LV, Stinear CM, Byblow WD. Ipsilateral motor pathways after stroke: implications for non-invasive brain stimulation. *Frontiers in human neuroscience.* 2013;7:184.

[107]. Bestmann S, Swayne O, Blankenburg F, Ruff CC, Teo J, Weiskopf N, Driver J, Rothwell JC, Ward NS. The role of contralateral dorsal premotor cortex after stroke as studied with concurrent TMS-fMRI. *Journal of Neuroscience.* 2010;30(36):11926-37.

[108]. Taga M, Hong YNG, Charalambous CC, Raju S, Hayes L, Lin J, Zhang Y, Shao Y, Houston M, Zhang Y, Mazzoni P, Roh J, Schambra HM. Corticospinal and corticoreticulospinal projections benefit motor behaviors in chronic stroke. *bioRxiv.* 2024;2024.04.04.588112. doi: 10.1101/2024.04.04.588112.

[109]. Sinopoulou E, Rosenzweig ES, Conner JM, Gibbs D, Weinholtz CA, Weber JL, Brock JH, Nout-Lomas YS, Ovruchesky E, Takashima Y. Rhesus macaque versus rat divergence in the corticospinal projectome. *Neuron.* 2022;110(18):2970-83. e4.

[110]. Du Beau A, Shrestha SS, Bannatyne B, Jalicy S, Linnen S, Maxwell D. Neurotransmitter phenotypes of descending systems in the rat lumbar spinal cord. *Neuroscience.* 2012;227:67-79.

[111]. Yang Q, Fang Y, Sun C-K, Siemionow V, Ranganathan VK, Khoshnabi D, Davis MP, Walsh D, Sahgal V, Yue GH. Weakening of functional corticomuscular coupling during muscle fatigue. *Brain research.* 2009;1250:101-12.

[112]. Ibáñez J, Del Vecchio A, Rothwell J, Baker S, Farina D. Only the fastest corticospinal fibers contribute to β corticomuscular coherence. *Journal of Neuroscience.* 2021;41(22):4867-79.

[113]. Liu J, Sheng Y, Liu H. Corticomuscular coherence and its applications: a review. *Frontiers in human neuroscience.* 2019;13:100.

[114]. Grosse P, Guerrini R, Parmeggiani L, Bonanni P, Pogosyan A, Brown P. Abnormal corticomuscular and intermuscular coupling in high-frequency rhythmic myoclonus. *Brain.* 2003;126(2):326-42.

[115]. Stokkermans M, Solis-Escalante T, Cohen MX, Weerdesteyn V. Distinct cortico-muscular coupling between step and stance leg during reactive stepping responses. *Frontiers in Neurology.* 2023;14:1124773.

[116]. Conway B, Halliday D, Farmer S, Shahani U, Maas P, Weir A, Rosenberg J. Synchronization between motor cortex and spinal motoneuronal pool during the performance of a maintained motor task in man. *The Journal of physiology.* 1995;489(3):917-24.

[117]. Hellwig B, Häußler S, Schelter B, Lauk M, Guschlbauer B, Timmer J, Lücking C. Tremor-correlated cortical activity in essential tremor. *The Lancet.* 2001;357(9255):519-23.

[118]. Airaksinen K, Lehti T, Nurminen J, Luoma J, Helle L, Taulu S, Pekkonen E, Mäkelä JP. Cortico-muscular coherence parallels coherence of postural tremor and MEG during static muscle contraction. *Neuroscience letters.* 2015;602:22-6.

[119]. Fang Y, Daly JJ, Sun J, Hvorat K, Fredrickson E, Pundik S, Sahgal V, Yue GH. Functional corticomuscular connection during reaching is weakened following stroke. *Clinical neurophysiology.* 2009;120(5):994-1002.

[120]. Kristeva R, Patino L, Omlor W. Beta-range cortical motor spectral power and corticomuscular coherence as a mechanism for effective corticospinal interaction during steady-state motor output. *Neuroimage.* 2007;36(3):785-92.

[121]. Artoni F, Fanciullacci C, Bertolucci F, Panarese A, Makeig S, Micera S, Chisari C. Unidirectional brain to muscle connectivity reveals motor cortex control of leg muscles during stereotyped walking. *Neuroimage.* 2017;159:403-16.

[122]. Riddle CN, Baker SN. Manipulation of peripheral neural feedback loops alters human corticomuscular coherence. *The Journal of physiology.* 2005;566(2):625-39.

[123]. Salenius S, Portin K, Kajola M, Salmelin R, Hari R. Cortical control of human motoneuron firing during isometric contraction. *Journal of neurophysiology.* 1997;77(6):3401-5.

[124]. Groß J, Tass P, Salenius S, Hari R, Freund H, Schnitzler A. Cortico-muscular synchronization during isometric muscle contraction in humans as revealed by magnetoencephalography. *The Journal of Physiology.* 2000;527(Pt 3):623.

[125]. Liu J, Wang J, Tan G, Sheng Y, Chang H, Xie Q, Liu H. Correlation evaluation of functional corticomuscular coupling with abnormal muscle synergy after stroke. *IEEE Transactions on Biomedical Engineering.* 2021;68(11):3261-72.

[126]. Chen X, Zhang Y, Cheng S, Xie P. Transfer spectral entropy and application to functional corticomuscular coupling. *IEEE Transactions on Neural Systems and Rehabilitation Engineering.* 2019;27(5):1092-102.

[127]. Xi X, Ding J, Wang J, Zhao Y-B, Wang T, Kong W, Li J. Analysis of functional corticomuscular coupling based on multiscale transfer spectral entropy. *IEEE Journal of Biomedical and Health Informatics.* 2022;26(10):5085-96.

[128]. Yang Y, Solis-Escalante T, Yao J, Daffertshofer A, Schouten AC, Van Der Helm FC. A general approach for quantifying nonlinear connectivity in the nervous system based on phase coupling. *International journal of neural systems.* 2016;26(01):1550031.

[129]. Xu Y, McClelland VM, Cvetković Z, Mills KR. Corticomuscular coherence with time lag with application to delay estimation. *IEEE Transactions on Biomedical Engineering.* 2016;64(3):588-600.

[130]. Yang Q, Siemionow V, Yao W, Sahgal V, Yue GH. Single-trial EEG-EMG coherence analysis reveals muscle fatigue-related progressive alterations in corticomuscular coupling. *IEEE transactions on neural systems and rehabilitation engineering*. 2010;18(2):97-106.

[131]. Xi X, Sun Z, Hua X, Yuan C, Zhao Y-B, Miran SM, Luo Z, Lü Z. Construction and analysis of cortical-muscular functional network based on EEG-EMG coherence using wavelet coherence. *Neurocomputing*. 2021;438:248-58.

[132]. Liu J, Wang J, Tan G, Sheng Y, Feng L, Tang T, Li X, Xie Q, Liu H, Wei Y. A generalized cortico-muscular-cortical network to evaluate the effects of three-week brain stimulation. *IEEE Transactions on Biomedical Engineering*. 2023.

[133]. Witham CL, Riddle CN, Baker MR, Baker SN. Contributions of descending and ascending pathways to corticomuscular coherence in humans. *The Journal of physiology*. 2011;589(15):3789-800.

[134]. Chen X, Xie P, Zhang Y, Chen Y, Yang F, Zhang L, Li X. Multiscale information transfer in functional corticomuscular coupling estimation following stroke: a pilot study. *Frontiers in neurology*. 2018;9:287.

[135]. Xie P, Cheng S, Zhang Y, Liu Z, Liu H, Chen X, Li X. Direct interaction on specific frequency bands in functional corticomuscular coupling. *IEEE Transactions on Biomedical Engineering*. 2019;67(3):762-72.

[136]. Mima T, Toma K, Koshy B, Hallett M. Coherence between cortical and muscular activities after subcortical stroke. *Stroke*. 2001;32(11):2597-601.

[137]. Negro F, Farina D. Linear transmission of cortical oscillations to the neural drive to muscles is mediated by common projections to populations of motoneurons in humans. *The Journal of physiology*. 2011;589(3):629-37.

[138]. Mima T, Hallett M. Corticomuscular coherence: a review. *Journal of clinical neurophysiology*. 1999;16(6):501.

[139]. Guo Z, Qian Q, Wong K, Zhu H, Huang Y, Hu X, Zheng Y. Altered corticomuscular coherence (CMCoh) pattern in the upper limb during finger movements after stroke. *Frontiers in Neurology*. 2020;11:410.

[140]. Zheng Y, Peng Y, Xu G, Li L, Wang J. Using corticomuscular coherence to reflect function recovery of paretic upper limb after stroke: a case study. *Frontiers in neurology*. 2018;8:728.

[141]. Yang Y, Dewald JP, van der Helm FC, Schouten AC. Unveiling neural coupling within the sensorimotor system: directionality and nonlinearity. *European journal of neuroscience*. 2018;48(7):2407-15.

[142]. Negro F, Farina D. Decorrelation of cortical inputs and motoneuron output. *Journal of neurophysiology*. 2011;106(5):2688-97.

[143]. Sinha N, Dewald JP, Heckman CJ, Yang Y. Cross-frequency coupling in descending motor pathways: Theory and simulation. *Frontiers in Systems Neuroscience*. 2020;13:86.

[144]. Yang Y, Solis-Escalante T, Van de Ruit M, Van der Helm FC, Schouten AC. Nonlinear coupling between cortical oscillations and muscle activity during isotonic wrist flexion. *Frontiers in computational neuroscience*. 2016;10:126.

[145]. Young CK, Eggermont JJ. Coupling of mesoscopic brain oscillations: recent advances in analytical and theoretical perspectives. *Progress in neurobiology*. 2009;89(1):61-78.

[146]. Rossiter HE, Eaves C, Davis E, Boudrias M-H, Park C-h, Farmer S, Barnes G, Litvak V, Ward NS. Changes in the location of cortico-muscular coherence following stroke. *NeuroImage: Clinical*. 2013;2:50-5.

[147]. Larsen LH, Zibrandtsen IC, Wienecke T, Kjaer TW, Christensen MS, Nielsen JB, Langberg H. Corticomuscular coherence in the acute and subacute phase after stroke. *Clinical Neurophysiology*. 2017;128(11):2217-26.

[148]. Choi S-M. Movement disorders following cerebrovascular lesions in cerebellar circuits. *Journal of movement disorders*. 2016;9(2):80.

[149]. Parmar N, Sirpal P, Sikora WA, Dewald JPA, Refai HH, Yang Y. Beta-band cortico-muscular phase coherence in hemiparetic stroke. *Biomedical Signal Processing and Control*. 2024;97:106719. doi: <https://doi.org/10.1016/j.bspc.2024.106719>.

[150]. Vasudeva B, Tian R, Wu DH, James SA, Refai HH, Ding L, He F, Yang Y. Multi-phase locking value: A generalized method for determining instantaneous multi-frequency phase coupling. *Biomedical signal processing and control*. 2022;74:103492.

[151]. Xu R, Zhang H, Shi X, Liang J, Wan C, Ming D. Lower-limb motor assessment with corticomuscular coherence of multiple muscles during ankle dorsiflexion after stroke. *IEEE Transactions on Neural Systems and Rehabilitation Engineering*. 2022;31:160-8.

[152]. Aumann TD, Prut Y. Do sensorimotor β -oscillations maintain muscle synergy representations in primary motor cortex? *Trends in neurosciences*. 2015;38(2):77-85.

[153]. Kristeva-Feige R, Fritsch C, Timmer J, Lücking C-H. Effects of attention and precision of exerted force on beta range EEG-EMG synchronization during a maintained motor contraction task. *Clinical Neurophysiology*. 2002;113(1):124-31.

[154]. McCrea PH, Eng JJ, Hodgson AJ. Saturated muscle activation contributes to compensatory reaching strategies after stroke. *Journal of neurophysiology*. 2005;94(5):2999-3008.

[155]. Cheung VC, Turolla A, Agostini M, Silvoni S, Bennis C, Kasi P, Paganoni S, Bonato P, Bizzi E. Muscle synergy patterns as physiological markers of motor cortical damage. *Proceedings of the national academy of sciences*. 2012;109(36):14652-6.

[156]. Roh J, Rymer WZ, Perreault EJ, Yoo SB, Beer RF. Alterations in upper limb muscle synergy structure in chronic stroke survivors. *Journal of neurophysiology*. 2013;109(3):768-81.

[157]. Sheng Y, Tan G, Liu J, Chang H, Wang J, Xie Q, Liu H. Upper limb motor function quantification in post-stroke rehabilitation using muscle synergy space model. *IEEE Transactions on Biomedical Engineering*. 2022;69(10):3119-30.

[158]. Pan B, Sun Y, Xie B, Huang Z, Wu J, Hou J, Liu Y, Huang Z, Zhang Z. Alterations of muscle synergies during voluntary arm reaching movement in subacute stroke survivors at different levels of impairment. *Frontiers in Computational Neuroscience*. 2018;12:69.

[159]. Laine CM, Valero-Cuevas FJ. Intermuscular coherence reflects functional coordination. *Journal of neurophysiology*. 2017;118(3):1775-83.

[160]. Yamanaka E, Goto R, Kawakami M, Tateishi T, Kondo K, Nojima I. Intermuscular Coherence during Quiet Standing in Sub-Acute Patients after Stroke: An Exploratory Study. *Brain Sciences*. 2023;13(12):1640.

[161]. Farmer S. Rhythmicity, synchronization and binding in human and primate motor systems. *The Journal of physiology*. 1998;509(1):3-14.

[162]. Obata H, Abe MO, Masani K, Nakazawa K. Modulation between bilateral legs and within unilateral muscle synergists of postural muscle activity changes with development and aging. *Experimental brain research*. 2014;232:1-11.

[163]. Fisher R, Galea M, Brown P, Lemon R. Digital nerve anaesthesia decreases EMG-EMG coherence in a human precision grip task. *Experimental brain research*. 2002;145:207-14.

[164]. Liu H, Gao Y, Huang W, Li R, Houston M, Benoit JS, Roh J, Zhang Y. Inter-muscular coherence and functional coordination in

the human upper extremity after stroke. *Math Biosci Eng.* 2022;19:4506-25.

[165]. Kisiel-Sajewicz K, Fang Y, Hrovat K, Yue GH, Siemionow V, Sun C-K, Jaskólska A, Jaskólski A, Sahgal V, Daly JJ. Weakening of synergist muscle coupling during reaching movement in stroke patients. *Neurorehabilitation and neural repair.* 2011;25(4):359-68.

[166]. Watanabe T, Saito K, Ishida K, Tanabe S, Nojima I. Coordination of plantar flexor muscles during bipedal and unipedal stances in young and elderly adults. *Experimental Brain Research.* 2018;236:1229-39.

[167]. Fisher KM, Zaaimi B, Williams TL, Baker SN, Baker MR. Beta-band intermuscular coherence: a novel biomarker of upper motor neuron dysfunction in motor neuron disease. *Brain.* 2012;135(9):2849-64.

[168]. Yin J, Ngiam KY, Teo HH. Role of Artificial Intelligence Applications in Real-Life Clinical Practice: Systematic Review. *J Med Internet Res.* 2021;23(4):e25759. Epub 20210422. doi: 10.2196/25759. PubMed PMID: 33885365; PMCID: PMC8103304.

[169]. Saikaley M, McIntyre A, Pauli G, Teasell R. A systematic review of randomized controlled trial characteristics for interventions to improve upper extremity motor recovery post stroke. *Top Stroke Rehabil.* 2023;30(4):323-32. Epub 20220214. doi: 10.1080/10749357.2022.2035578. PubMed PMID: 35156561.

[170]. Boden-Albala B, Carman H, Southwick L, Parikh NS, Roberts E, Waddy S, Edwards D. Examining Barriers and Practices to Recruitment and Retention in Stroke Clinical Trials. *Stroke.* 2015;46(8):2232-7. doi: doi:10.1161/STROKEAHA.114.008564.

[171]. Feldman WB, Kim AS, Chiong W. Trends in Recruitment Rates for Acute Stroke Trials, 1990–2014. *Stroke.* 2017;48(3):799-801. doi: doi:10.1161/STROKEAHA.116.014458.

[172]. Hernandez Petzsche MR, de la Rosa E, Hanning U, Wiest R, Valenzuela W, Reyes M, Meyer M, Liew S-L, Kofler F, Ezhov I. ISLES 2022: A multi-center magnetic resonance imaging stroke lesion segmentation dataset. *Scientific data.* 2022;9(1):762.

[173]. Collaboration A. The ATLAS experiment at the CERN large hadron collider. *Journal of Instrumentation.* 2008.

[174]. Challenge SD. American Heart Association; 2023. Available from: [Www.heart.org.https://www.heart.org/en/professional/quality-improvement/quality-research-and-publications/stroke-data-challenge](https://www.heart.org/en/professional/quality-improvement/quality-research-and-publications/stroke-data-challenge).